SPECIAL ISSUE EDITORIAL



Overcoming adversity through diversity: aquatic carbon concentrating mechanisms

Carbon concentrating mechanism (CCM) systems, associated with evolutionarily diverse aquatic photosynthetic organisms, make a major contribution to global net primary productivity and marine carbon sequestration. Here, an overview of these global contributions is presented from their evolutionary origins, including a possible trigger for their diversification when the aqueous O_2/CO_2 ratio rose above parity, and a re-definition of the paradox of phytoplankton. The reviews and research in the special issue also include molecular physiology and ecology of CCMs, through to future potential applications for sustaining carbon sequestration and supporting terrestrial crop productivity.

The inorganic carbon substrate supply needed for photosynthesis in the aquatic milieu is limited by inorganic carbon solubility and diffusion across the boundary layer, cell wall and multiple membranes to the primary carboxylase Rubisco. Various biophysical carbon concentrating mechanism (CCM) systems are found in many aquatic phytoplankters and have overcome these chemical and physical limitations. Such CCMs deliver the appropriate inorganic carbon species demanded by Rubisco (CO₂), at an enhanced concentration which compensates for the enzyme's low substrate affinity and competitive inhibition from oxygen.

Despite this adversity, aquatic organisms clearly punch above their weight of biomass relative to terrestrial plants. The instantaneous standing biomass crop of aquatic plants (primarily microorganisms) is 3 PgC (i.e. 10^{15} g carbon) relative to the 610 PgC usually quoted for terrestrial plant above-ground biomass. The paradox of how phytoplankton deliver an annual net primary productivity of 47.5 PgC, relative to the 56.4 PgC of their terrestrial counterparts (Field *et al.*, 1998), has long intrigued researchers. In addition, the oceanic sink for net carbon sequestration is equal to that of land plants (2.3 PgC per year), such that marine organisms also facilitate the absorption of over 25% of annual anthropogenic CO₂ emissions (Pan *et al.*, 2011).

The original paradox of the phytoplankton was thought to reflect phylogenetic diversity in competition for limiting light and inorganic resources. The high net primary productivity, identified by Field *et al.* (1998), could be explained by the interaction between ecological and environmental factors across space and time to prevent the dominance of any one phytoplankton group. Despite this contention, it is with some amusement we note that each authority tends to claim preeminence for the contribution made by their particular phytoplankton clade to net primary productivity!

However, the past few decades have seen several historical paradigms overturned – such as photosynthetic acclimation to light increasing the depth of the photic zone (Richardson *et al.*, 1983; Raven *et al.*, 2017), the breadth of productivity across oceanic gyres (Johnson *et al.*, 2006; Partensky and Garczarek, 2010), and the molecular basis of niche differentiation found within cyanobacterial and eukarotic picoplankton populations in coastal and equatorial waters (Not *et al.*, 2012; Biller *et al.*, 2015). Additionally, we now recognize that more than 80% of marine primary productivity will be facilitated by some form of CCM (Raven and Beardall, 2016; Raven *et al.*, 2017).

This special issue provides a comprehensive update on aquatic carbon concentrating mechanisms, as well as reflection on how the field has progressed since the 1980s (Kaplan, 2017) together with the latest new research (see Box 1).

Box 1. Pioneering contributions over 40 years

The diversity of papers presented in this special issue reflects the range of contributions made at CCM9 in 2016, the ninth International Symposium on Inorganic Carbon Uptake by Aquatic Photosynthetic Organisms (Cambridge, UK; a satellite meeting following the 17th International Congress on Photosynthesis in Maastricht, The Netherlands). At the meeting, we were able to celebrate pioneering contributions over the past 40 years in person with Joe Berry, Aaron Kaplan and John Raven, and also recognize the outstanding technical and theoretical innovations made throughout this period by Murray Badger. A series of special publications has historically accompanied previous CCM Symposia, starting with the pioneering ASPP (American Society of Plant Physiologists) 'Green Book' proceedings from the first meeting in Asilomar (CA, USA) (Lucas and Berry, 1985), and through to that summarized by Moroney and Wee (2014). In the current special issue, we capture this progression with the highly personalized account by Aaron Kaplan of CCM research developments during those early years (Kaplan, 2017).

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Palaeohistorical and environmental drivers for CCM origins and diversity

We have more certainty about the timeline for the diversification of prokaryotic and eukaryotic clades, and their contrasting endosymbiotic exchanges, than for the origins and diversity of CCMs. Using projections based on the current Rubisco content and kinetic properties of extant cyanobacteria, Raven *et al.* (2017) suggest that the atmospheric CO₂ concentrations ranging from ×10 current to ×4.5 current, between 1.6 to 0.6 Ga (i.e. 10^9 years ago), could have been associated with CCM activity (see also Riding, 2006). Raven *et al.* (2017) also speculate that were *Gloeobacter* to be representative of a basal cyanobacterium, as accorded by some phylogenetic studies, then the CCM could even extend back to the Great Oxidation Event at 2.4 Ga.

Eukaryotic CCMs are generally thought to be homoplastic, with independent origins in each lineage (Raven et al., 2017). The primary endosymbiosis which led to the earliest oxygenic eukaryotes was likely to have been 1.0-1.6 Ga, with green, red and glaucophyte lineages subsequently diversifying via secondary and tertiary endosymbioses (Leliaert et al., 2012). The green algal lineage leading to the Chlorophyta is thought to have diverged via the Prasinophyceae from the Streptophyta by some 0.5-0.75 Ga. The red lineage (Rhodophyta) gave rise to Chromist algae (e.g. Haptophyta, Cryptophyta and Dinophyta), with haptophytes diversifying around 0.5 Ga and diatoms around 0.2 Ga (Heureux et al., 2017; Young and Hopkinson, 2017). Whilst the Rubisco large subunit has provided key phylogenetic insights for this progression (Badger and Price, 2003; Price et al., 2013), the coevolution of Rubisco variants, their kinetic properties and responsiveness to CO₂ and O₂ remain a critical element in CCM evolution.

Rubisco is not only sensitive to CO_2 , rather the CO_2/O_2 ratio at its active site, and there is some evidence that oxygen exclusion may be an important component in many biophysical CCM systems (Meyer and Griffiths, 2013; Heureux *et al.*, 2017; Meyer *et al.*, 2017). Against the backdrop of generally declining CO_2 and increasing O_2 over geological history, it is an interesting thought-experiment to explore at what point rising aqueous O_2 overtook CO_2 to become more dominant in seawater. It is worth noting that such a chemical event would occur at different times in the atmosphere and ocean. CO_2 is approximately 30 times more soluble than O_2 in seawater (Fig. 1), so atmospheric CO_2 can be 30 times less concentrated than O_2 in the atmosphere, but the two species will be equimolar in the ocean.

In the early stages of the oxygenation of the atmosphere during the Great Oxidation Event (2.4 Ga), oxygen is estimated to have risen to between 10^{-2} and 10^{-1} PAL (present atmospheric level), i.e. between 210 ppmV and 2100 ppmV (Lyons *et al.*, 2014). The best estimates of CO₂ at this time suggest that it was probably in excess of 100 PAL (i.e. 35 000 ppmV) (Young *et al.*, 2012), so orders of magnitude more abundant than O₂. Even if oxygen levels persisted at 0.1 to 0.2 PAL (21 000 ppmV) through the 'boring billion' (approximately 2 to 1 Ga) until close to the Precambrian/Cambrian boundary, CO₂ levels would need to be 2-4 PAL (i.e. 7-1400 ppmV) to be equimolar with oxygen in the ocean, much lower than current estimates for this time. The timing then of parity between O₂ and CO₂ concentrations in the oceans is determined by the point at which O_2 rose from around 0.2 PAL to close to modern values, and CO₂ was sufficiently low to be equimolar. The most recent estimates of this O₂ rise through 0.5 PAL (10.5%), around 450 Ma (Lenton et al., 2016), requires an atmospheric concentration of 10 PAL CO_2 (3500 ppm) to provide equimolar dissolved CO_2 and O_2 in marine waters, a CO₂ level which is well within range of coincident atmospheric estimates (Fig. 2). Therefore, the environmental threshold of O₂ overtaking CO₂ in surface waters, driving marine organisms to provide a mechanism to boost the CO_2/O_2 ratio at the site of Rubisco, is likely to date to the invasion of land by the earliest plants around the late Silurian/early Devonian. Such a timing seems to agree well with an analysis of a limited number of Rubisco large subunit sequences which also finds a number of events of positive selection up to 410 Ma. This indicates that emergence of CCMs around this time indeed left a footprint in the Rubisco protein (Young et al., 2012). Other, more conservative estimates, put the origins of cyanobacterial and eukaryotic CCMs following the 'Devonian Drop' and during the Carboniferous (Badger and Price, 2003).

There appears to be an evolutionary progression of a higher Rubisco specificity factor (selectivity for CO₂ over O₂) from cyanobacterial, chlorophyte and then to higher plant Form 1B Rubisco (Meyer and Griffiths, 2013). Considering the Form 1D in the marine algae, and Rhodophyta as the endosymbiont, the evolutionary trend is towards decreased Rubisco specificity factor and lower carbon affinity (higher K_c). This has been interpreted as an evolutionary response to improved CCM activity that allows a relaxation of substrate affinity and faster catalytic turnover of the enzyme (Tcherkez *et al.*, 2006) in rhodophytes Young *et al.*, 2012; Heureux *et al.*, 2017) as well as in cyanobacteria and chlorophytes (Meyer and

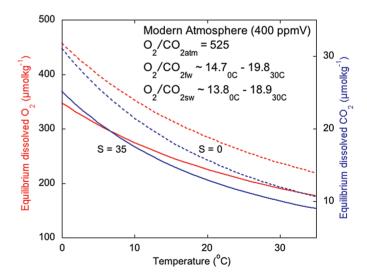


Fig. 1. (a) The sensitivity of equilibrium dissolved CO₂ (blue; Weiss, 1974) and O₂ (red; Benson and Krause, 1984) concentrations to temperature and salinity (S; 0, dashed line, and 35 ppt, solid line) and the modern range (with an atmosphere of 400 ppmV) of dissolved O₂/CO₂ ratios for freshwater (fw) and seawater (sw) between 0 and 30 °C.

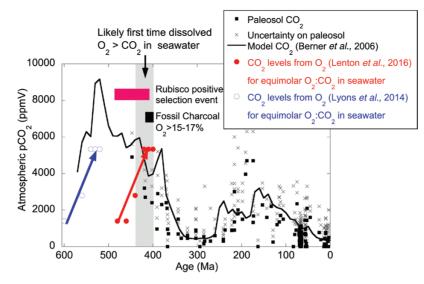


Fig. 2. An estimate of the threshold when the concentration of dissolved aqueous O_2 rose to a higher concentration than CO_2 in seawater as a trigger for the emergence of CCMs (grey bar). The CO_2 concentrations that would be equimolar with O_2 reconstructions (blue open circles – Lyons *et al.*, 2014; red closed circles – Lenton *et al.*, 2016) are based on an approximate 30-fold difference in solubilities. The Phanerozoic history of atmospheric CO_2 is compiled from paleosols (black squares with crosses to show the uncertainty; Royer 2006) and the GEOCARB III model (solid black line; Berner and Kothavala, 2001). Also shown are the first appearance of fossil charcoal evidence that $O_2 > 15 - 17\%$ (black bar, Glasspool *et al.*, 2004), and a Rubisco positive selection event (magenta bar, Young *et al.*, 2012).

Griffiths, 2013). An apparent breakdown in the canonical trade-off between affinity for carbon and turnover in modern diatom Form 1D Rubisco (Young *et al.*, 2016) challenges our understanding of this evolutionary progression to lower carbon affinities. It may be necessary to consider the impact of other Rubisco catalytic parameters on Rubisco performance.

As well as the correlations between Rubisco affinity and turnover rate for both substrates, O₂ and CO₂ (Heureux et al., 2017), the K_c and K_o of many Rubiscos also appear to be linked (Fig. 3). The trade-off of a relaxed affinity for oxygen (higher K_{0} , which accompanies a more efficient CCM and a higher K_c , could improve Rubisco performance in an increasingly oxygenated environment. Whether this relationship between K_c and K_0 is a constraint imposed by the structure of the enzyme (Savir *et al.*, 2010), or a response to the O_2/CO_2 ratio at the active site of the Rubisco remains an open question. What is curious is that the ratio of the affinities of Rubisco for O2 and $CO_2 (K_o/K_c = \sim 16)$ of many Form 1D-containing marine algal Rubiscos, including most diatoms, and the Form 1B-containing green algae (e.g. Chlamydomonas), appears to match the modern dissolved O_2/CO_2 ratio of natural waters at equilibrium with an atmosphere containing 400 ppmV CO₂ (16 at a representative sea surface temperature of 12 °C; see Fig. 3, and Heureux *et al.*, 2017). The affinity of these Rubiscos for O_2 is 16 times lower than that for CO_2 . This implies that the activity of a CCM to elevate the Rubisco K_c values above the environmental availability of CO₂ also compensates perfectly for the environmental excess of O_2 over CO_2 . Yet there is a bimodality to the Rubisco K_0/K_c ratio. The C₃ plants and some C₄ plants tend to fall on a line with a K_0/K_c gradient of 35. Such a ratio is much higher than the dissolved O_2/CO_2 ratio of any modern natural waters (see Fig. 1). However, during Pleistocene glacial periods, atmospheric CO₂ fell to ~ 180 ppmV (Lüthi *et al.*,

2008), but atmospheric O_2 remained constant so the environmental dissolved O₂/CO₂ ratio was more than doubled relative to the modern (i.e. ~35). Modern Rubiscos in plants and algae, therefore, appear to be tuned in terms of O_2/CO_2 affinities to compensate for either glacial or modern O_2/CO_2 such that the Rubisco experiences a 1:1 competition between O_2 and CO_2 at the active site. This underpins the concept that it was the rising of environmental dissolved O_2/CO_2 above 1 that triggered the emergence of a CCM. It further demonstrates that the Rubisco enzyme, traditionally thought to be an inefficient relic of ancient environments, is highly dynamic. Rubisco appears to evolve its kinetics in response to environmental change over timescales of at least tens of kyrs, if not hundreds of years. The K_o/K_c data appear better tuned to anthropogenic conditions rather than average interglacial atmospheric compositions. At any event, we urgently need additional analyses of Rubisco kinetic properties and sequence specificity for Chromists (Young et al., 2016, 2017) as well as across the green algal lineages (Goudet, 2016) to fully understand the evolutionary history, rate of change and current diversity of carbon handling across the photosynthesizers.

A final consideration is that the origin and maintenance of CCMs might reflect environmental limitations, in addition to external inorganic carbon supply. Interactions between nitrogen availability may relate to a reduced requirement for catalytic protein following CCM induction in *Chlorella* (Beardall *et al.*, 1982), although other evidence is equivocal (Ruan *et al.*, 2017). Consistent with the earlier observation, the proportion of Rubisco of total soluble protein is low in cells with a higher CCM efficiency in chromists, as evidenced by raised K_c of the Rubisco (Young *et al.*, 2016; Heureux *et al.*, 2017). Low energetic availability (as light or P) tends to reduce CCM activity (Maberly and Gontero, 2017), and interactions with low temperature may also have been significant, whether directly in

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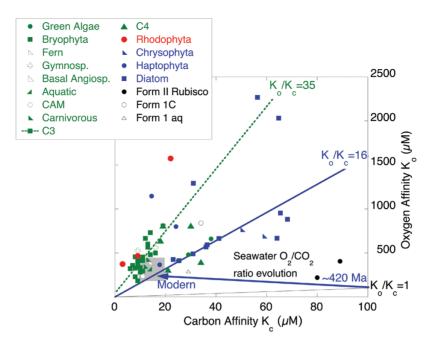


Fig. 3. Compiled Rubisco K_c (µmol kg⁻¹) versus K_o (µmol kg⁻¹) (Galmés *et al.*, 2014; Young *et al.*, 2016; Heureux *et al.*, 2017) for a range of plant and algal species with Form 1B Rubisco-containing organisms in green, and Form 1D Rubisco-containing organisms in red (rhodophyta) and blue (diatoms, chrysophytes and haptophytes). Lines are plotted that indicate a K_c/K_o ratio of 1 (black), 16 (blue, equivalent to modern dissolved O_2/CO_2 ratio of seawater) and 35 (an apparent line of best fit to the C_3 data). The range of environmental aqueous concentrations is indicated by the grey box. The evolution of the dissolved concentrations and O_2/CO_2 ratio from ~1 at 420 Ma (see Fig. 2) to 16 today ('modern') is indicated by the blue arrow. Note that the $O_2:CO_2$ dissolved ratio of natural waters at glacial maxima of the Pleistocene was ~35.

terms of survival during the Cryogenian snowball earth, or at higher latitudes in terrestrial and marine algae (Raven *et al.*, 2017). One additional driver for a CCM could also occur when inorganic carbon becomes locally depleted within a dense algal bloom, and so may be to some extent independent of equilibration with ambient air (Maberly and Gontero, 2017).

Convergence in CCM form and function

The three pillars usually invoked to support a CCM (Meyer and Griffiths, 2013) include:

- biophysical inorganic transporters, operating in parallel across adjacent membranes, raising the inorganic carbon pool by some 40-fold (Chlorophyte) to 400-fold (Cyanobacteria) and determining overall affinity and effectiveness of the CCM;
- (ii) a suite of strategically placed carbonic anhydrases
 (CA) and CA-like moieties, adjacent to the inorganic transporters, to assist in bicarbonate interconversion or regeneration (or recapture) of CO₂ close to Rubisco;
- (iii) a microcompartment within which Rubisco aggregates, and from which CO₂ leakage is minimized, such as the carboxysome in cyanobacteria and pyrenoid associated with most eukaryotic CCM systems.

Papers in this special issue report on the latest developments in identifying key cyanobacterial and chlorophyte CCM components (Rae *et al.*, 2017), albeit in the context of their potential for introduction into higher plants to augment productivity (see also Price *et al.*, 2013). Cyanobacterial α - and β -carboxysomes regulate the influx of bicarbonate and other metabolite exchanges via pore structures in the proteinaceous shell, with CO₂ converted internally by CA systems (for details see Rae *et al.*, 2017). The detailed variations between the two cyanobacterial lineages were described by Price *et al.* (2008), but in α -carboxysomes, Form-1A Rubisco is attached to the highly disordered CsoS2 protein, and in β -carboxysomes Form IB Rubisco is integrated via small subunit substitutions to the full-length CcmM protein in an ordered array (Rae *et al.*, 2017).

Additional insights for the carboxysome shell proteins are provided by Sommer *et al.* (2017), who have undertaken a bioinformatic survey of β -carboxysome shell proteins, which suggest that variations in carboxysome structure allow plasticity in response to changing environmental conditions. Meanwhile, Larsson *et al.* (2017) provide crystallographic structural insights for regulation of metabolite exchange by gating of the CcmP protein in the β -carboxysome shell of *Synechococcus elongatus* PCC7942.

The diversity of CCM systems in most photosynthetic eukaryotics mostly requires all three physiological pillars indicated above, and those few which lack an identifiable pyrenoid show reduced capacity for carbon accumulation (Giordano *et al.*, 2005). In this special issue, the comparative evolution of pyrenoids in chlorophytes and chromists is discussed in terms of the commonalities seen in mode of Rubisco aggregation, usually in association with some specialized thylakoid membrane organization (Meyer *et al.*, 2017), although others may be stalked, and in some dinoflagellates Rubisco aggregation is more transient, forming centrally under circadian control (Nassoury *et al.*, 2001).

For chlorophytes, the CCM in *Chlamydomonas* is the best-defined system from a molecular perspective (Meyer and Griffiths, 2013), with hierarchical models presented for regulatory processes leading to CCM induction

(Mitchell et al., 2017). Various mutagenic screens and genetic manipulations have helped to characterize components of the Chlamydomonas CCM (Li et al., 2016; Machingura et al., 2017), including recent observations on the protein elements associated with Rubisco aggregation (Mackinder et al., 2016; Mitchell et al., 2017), and a new potential thylakoid bicarbonate transporter (Machingura et al., 2017). The observation that specific elements of Rubisco small subunits (SSU) were integral to the aggregation mechanism (Meyer et al., 2012) has led to insights into the function of a possible linker protein (EPYC1, formerly LCI5: Mackinder et al., 2016), and also now for the hierarchical organization of the pyrenoid (Meyer et al., 2017). The Chlamydomonas SSU mutants retain the knotted thylakoid tubules that intersect at the heart of the usual pyrenoid location, with growth and photosynthesis restored under elevated CO₂ supply (Caspari et al., 2017). Such observations suggest that the pyrenoidassociated starch sheath and additional external regulatory elements (LCIB/C) are dependent upon Rubisco aggregation, and the spatial segregation of PSII (normally excluded from within the pyrenoid matrix) does not compromise overall energetic efficiency (Caspari et al., 2017).

For diatoms, whilst the specific details of CCM processes are less well understood than for Chlamydomonas, we have more detailed comparative insights into contrasting CCM systems for a wider range of species across the clade. Here, the four layers of thylakoid membranes, associated with secondary plastid endosymbiosis in the Dinophyceae, offer a range of options for concentrating inorganic carbon. Young and Hopkinson (2017) highlight the contrasting trade-offs which seem to have occurred in terms of investment in Rubisco relative to altered Rubisco kinetic properties for contrasting marine habitats. Matsuda et al. (2017; see also Tsuji et al., 2017) outline the contrasting modes of carbon uptake and conversion thought to operate in diatoms, dependent on either diffusive entry of CO₂ or active transport, with one mechanism supported by the more detailed observations seen for the role of CAH1 in Nannochloropsis oceanica (Gee et al., 2017).

Further insights are provided by the co-evolution of inorganic carbon transporters (SLC4) in diatoms, used in combination with contrasting CA species (Shen *et al.*, 2017), and also by the use of an intra-thylakoid CA to regenerate CO_2 adjacent to the aggregated Rubisco (Tsuji *et al.*, 2017), suggesting convergence with the mechanism also proposed for Chlorophytes (Meyer and Griffiths, 2013).

From molecular diversity to overcoming ecological adversity

The ecological implications of CCM systems are also addressed from an experimental perspective in a number of papers in this special issue. Evolutionary origins (Raven *et al.*, 2017) are complemented by a more detailed comparison of ecological drivers in marine, freshwater and terrestrial habitats by Maberly and Gontero (2017). This leads to a highly original analysis of competitive interactions between cyanobacterial and chlorophyte cells (Ji *et al.*, 2017; see also the Insight article by Beardall and Raven, 2017). These observations are consistent with notions that CCMs help to overcome adversity, as defined above in terms of nutrient availability or local depletion of inorganic carbon within blooms. Thus, chlorophyte algae endure under low ambient CO_2 equilibration, relative to cyanobacteria, despite their 'less effective' CCM; whilst cyanobacteria may thrive under future elevated CO_2 conditions (Ji *et al.*, 2017; Beardall and Raven, 2017).

The major contribution made by diatoms to biogeochemical cycles, as reviewed by Young and Hopkinson (2017), is further characterized experimentally by a comparison of the effectiveness of the various inorganic carbon accumulation mechanisms (Clement *et al.*, 2017). And finally, although macrophytes make a relatively small contribution to marine net primary productivity (1 PgC per year: Field *et al.*, 1998), the issue contains papers analyzing the mechanisms of inorganic carbon uptake in seagrasses (Larkum *et al.*, 2017) and Antarctic macrophytes (Iñiguez *et al.*, 2017).

The future

The papers in this special issue convey a renewed sense of excitement and impetus in the field of aquatic carbon concentrating mechanisms, and include contributions from many young scientists with an astonishing breadth of skills, encompassing structural biology, novel molecular manipulations and bioinformatic approaches which are now augmenting traditional physiological and ecological experimentation. Globally, we may face uncertainty, but the potential for CCM systems to enhance marine carbon sequestration (Heureux *et al.*, 2017; Raven *et al.*, 2017; Young and Hopkinson, 2017) or terrestrial crop productivity (Rae *et al.*, 2017), informed by ongoing cutting-edge research programmes, provide some hope, and much promise, for the future.

Howard Griffiths^{*,1}, Moritz T. Meyer^{1,2} and Rosalind E.M. Rickaby³

 ¹ Department of Plant Sciences, University of Cambridge, Cambridge CB2 3EA, UK
 ² Department of Molecular Biology, Princeton University, Princeton, NJ 08544-1014
 ³ University of Oxford, Department of Earth Science, Oxford OX1 3AN, UK

* Correspondence: hg230@cam.ac.uk

References

Badger MR, Price GD. 2003. CO_2 concentrating mechanisms in cyanobacteria: molecular components, their diversity and evolution. Journal of Experimental Botany **54**, 609–622.

Beardall J, Griffiths H, Raven JA. 1982. Carbon isotope discrimination and the CO₂ accumulating mechanism in *Chlorella pyrenoidosa*. Journal of Experimental Botany **33**, 729–737.

Beardall J, Raven J. 2017. Cyanobacteria vs green algae: which group has the edge? Journal of Experimental Botany **68**, 3697–3699.

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Benson BB, Krause D. 1984. The concentration and isotopic fractionation of oxygen dissolved in freshwater and seawater in equilibrium with the atmosphere. Limnology and Oceanography **29,** 620–632.

Berner RA, Kothavala Z. 2001. GEOCARB III: a revised model of atmospheric CO_2 over Phanerozoic time. Americal Journal of Science **301**, 182–204.

Biller SJ, Berube PM, Lindell D, Chisholm SW. 2015. Prochlorococcus: the structure and function of collective diversity. Nature Reviews Microbiology **13**, 13–27.

Caspari OD, Meyer MT, Tolleter D, Wittkopp TM, Cunniffe NJ, Lawson T, Grossman AR, Griffiths H. 2017. Pyrenoid loss in *Chlamydomonas reinhardtii* causes limitations in CO₂ supply, but not thylakoid operating efficiency. Journal of Experimental Botany **68**, 3903–3913.

Clement R, Jensen E, Prioretti L, Maberly SC, Gontero B. 2017. Diversity of CO_2 concentrating mechanisms and responses to CO_2 concentration in marine and freshwater diatoms. Journal of Experimental Botany **68**, 3925–3935.

Field CB, Behrenfeld MJ, Randerson JT, Falkowski P. 1998. Primary production of the biosphere: integrating terrestrial and oceanic components. Science **281**, 237–240.

Galmés J, Kapralov MV, Andralojc PJ, Conesa MÀ, Keys AJ, Parry MA, Flexas J. 2014. Expanding knowledge of the Rubisco kinetics variability in plant species: environmental and evolutionary trends. Plant, Cell & Environment **37**, 1989–2001.

Gee CW, Nyogi KK. 2017. The carbonic anhydrase CAH1 is an essential component of the carbon-concentrating mechanism in *Nannochloropsis oceanica*. Proceedings of the National Academy of Sciences, USA **114**, 4537–4542.

Giordano M, Beardall J, Raven JA. 2005. CO₂ concentrating mechanisms in algae: mechanisms, environmental modulation, and evolution. Annual Review of Plant Biology **56**, 99–131.

Glasspool IJ, Edwards D, Axe L. 2004. Charcoal in the Silurian as evidence for the earliest wildfire. Geology **32**, 381–383.

Goudet MMMG. 2016. Evolutionary history of ribulose-1,5- biphosphate carboxylase/oxygenase and the origin of the algal pyrenoid. MPhil Thesis. University of Cambridge.

Han X, Sun N, Xu M, Mi H. 2017. Co-ordination of NDH and Cup proteins in CO₂ uptake in cyanobacterium *Synechocystis* sp. PCC 6803. Journal of Experimental Botany **68**, 3869–3877.

Heureux AMC, Young JN, Whitney SM, Eason-Hubbard MR, Lee RBY, Sharwood RE, Rickaby REM. 2017. The role of Rubisco kinetics and pyrenoid morphology in shaping the CCM of haptophyte microalgae. Journal of Experimental Botany **68**, 3959–3969.

Iñiguez C, Heinrich S, Harms L, Gordillo FJL. 2017. Increased temperature and CO_2 alleviate photoinhibition in *Desmarestia anceps*: from transcriptomics to carbon utilization. Journal of Experimental Botany **68**, 3971–3984.

Ji X, Verspagen JMH, Stomp M, Huisman J. 2017. Competition between cyanobacteria and green algae at low *versus* elevated CO₂: who will win, and why? Journal of Experimental Botany **68**, 3815–3828.

Johnson ZI, Zinser ER, Coe A, McNulty NP, Woodward EM, Chisholm SW. 2006. Niche partitioning among Prochlorococcus ecotypes along ocean-scale environmental gradients. Science **311**, 1737–1740.

Kaplan A. 2017. On the cradle of CCM research: discovery, development, and challenges ahead. Journal of Experimental Botany **68**, 3785–3796.

Larkum AWD, Davey PA, Kuo J, Ralph PJ, Raven JA. 2017. Carbonconcentrating mechanisms in seagrasses. Journal of Experimental Botany 68, 3773–3784.

Larsson AM, Hasse D, Valegård K, Andersson I. 2017. Crystal structures of β -carboxysome shell protein CcmP: ligand binding correlates with the closed or open central pore. Journal of Experimental Botany **68**, 3857–3867.

Leliaert F, Smith DR, Moreau H, Herron MD, Verbruggen H, Delwiche CF, De Clerck O. 2012. Phylogeny and molecular evolution of the green algae. Critical Reviews in Plant Sciences **31**, 1–46.

Lenton TM, Dahl TW, Daines SJ, Mills BJ, Ozaki K, Salzmann MR, Porada P. 2016. Earliest land plants created modern levels of atmospheric oxygen. Proceedings of the National Academy of Sciences, USA **113**, 9704–9709. Li X, Zhang R, Patena W, et al. 2016. An indexed, mapped mutant library enables reverse genetics studies of biological processes in *Chlamydomonas reinhardtii*. The Plant Cell **28**, 367–387.

Lucas WJ, Berry JA. 1985. *Inorganic carbon uptake by aquatic photosynthetic organisms*. Rockville, MD: American Society of Plant Physiologists.

Lüthi D, Le Floch M, Bereiter B, *et al.* 2008. High-resolution carbon dioxide concentration record 650 000–800 000 years before present. Nature **453**, 379–382.

Lyons TW, Reinhard CT, Planavsky NJ. 2014. The rise of oxygen in Earth's early ocean and atmosphere. Nature **506**, 307–315.

Maberly SC, Gontero B. 2017. Ecological imperatives for aquatic carbon dioxide-concentrating mechanisms. Journal of Experimental Botany **68**, 3797–3814.

Machingura MC, Bajsa-Hirschel J, Laborde SM, Schwartzenburg JB, Mukherjee B, Mukherjee A, Pollock SV, Förster B, Price GD, Moroney JV. 2017. Identification and characterization of a solute carrier, CIA8, involved in inorganic carbon acclimation in *Chlamydomonas reinhardtii*. Journal of Experimental Botany **68**, 3879–3890.

Mackinder LC, Meyer MT, Mettler-Altmann T, et al. 2016. A repeat protein links Rubisco to form the eukaryotic carbon-concentrating organelle. Proceedings of the National Academy of Sciences, USA **113**, 5958–5963.

Meyer M, Griffiths H. 2013. Origins and diversity of eukaryotic CO_2 -concentrating mechanisms: lessons for the future. Journal of Experimental Botany **64**, 769–786.

Meyer MT, Genkov T, Skepper JN, et al. 2012. Rubisco small-subunit α -helices control pyrenoid formation in *Chlamydomonas*. Proceedings of the National Academy of Sciences, USA **109**, 19474–19479.

Meyer MT, Whittaker C, Griffiths H. 2017. The algal pyrenoid: key unanswered questions. Journal of Experimental Botany **68**, 3739–3749.

Mitchell MC, Metodieva G, Metodiev MV, Griffiths H, Meyer MT. 2017. Pyrenoid loss impairs carbon-concentrating mechanism induction and alters primary metabolism in *Chlamydomonas reinhardtii*. Journal of Experimental Botany **68**, 3891–3902.

Moroney JV, Wee JL. 2014. CCM8: the eighth international symposium on inorganic carbon uptake by aquatic photosynthetic organisms. Photosynthesis Research **121**, 107–110.

Nassoury N, Fritz L, Morse D. 2001. Circadian changes in ribulose-1,5-bisphosphate carboxylase/oxygenase distribution inside individual chloroplasts can account for the rhythm in dinoflagellate carbon fixation. The Plant Cell **13**, 923–934.

Not F, Siano R, Kooistra WHCF, *et al.* 2012. Diversity and ecology of eukaryotic marine phytoplankton. Advances in Botanical Research **64**, 1–53.

Pan Y, Birdsey RA, Fang J, et al. 2011. A large and persistent carbon sink in the world's forests. Science **333**, 988–993.

Partensky F, Garczarek L. 2010. Prochlorococcus: advantages and limits of minimalism. Annual Review of Marine Science 2, 305–331.

Price GD, Badger MR, Woodger FJ, Long BM. 2008. Advances in understanding the cyanobacterial CO₂-concentrating-mechanism (CCM): functional components, Ci transporters, diversity, genetic regulation and prospects for engineering into plants. Journal of Experimental Botany **59**, 1441–1461.

Price GD, Pengelly JJ, Forster B, Du J, Whitney SM, von Caemmerer S, Badger MR, Howitt SM, Evans JR. 2013. The cyanobacterial CCM as a source of genes for improving photosynthetic CO₂ fixation in crop species. Journal of Experimental Botany **64**, 753–768.

Rae BD, Long BM, Förster B, Nguyen ND, Velanis CN, Atkinson N, Hee WY, Mukherjee B, Price GD, McCormick AJ. 2017. Progress and challenges of engineering a biophysical carbon dioxide-concentrating mechanism into higher plants. Journal of Experimental Botany **68**, 3717–3737.

Raven JA, Beardall J. 2016. The ins and outs of CO_2 . Journal of Experimental Botany **67**, 1–13.

Raven JA, Beardall J, Sánchez-Baracaldo P. 2017. The possible evolution, and future, of CO₂-concentrating mechanisms. Journal of Experimental Botany **68**, 3701–3716.

Richardson K, Beardall J, Raven JA. 1983. Adaptation of unicellular algae to irradiance: an annlysis of strategies. New Phytologist **93**, 157–191.

Riding R. 2006. Cyanobacterial calcification, carbon dioxide concentrating mechanisms, and Proterozoic–Cambrian changes in atmospheric composition. Geobiology **4**, 299–316.

Royer D. 2006. CO_2 -forced climate thresholds during the Phanerozoic. Geochimica et Cosmochimica Acta **70**, 5665–5675.

Ruan Z, Raven JA, Giordano M. 2017. In *Synechococcus* sp. competition for energy between assimilation and acquisition of C and those of N only occurs when growth is light limited. Journal of Experimental Botany **68**, 3829–3839.

Savir Y, Noor E, Milo R, Tlusty T. 2010. Cross-species analysis traces adaptation of Rubisco toward optimality in a low-dimensional landscape. Proceedings of the National Academy of Sciences, USA **107**, 3475–3480.

Shao H, Gontero B, Maberly SC, Jiang HS, Cao Y, Li W, Huang WM. 2017. Responses of *Ottelia alismoides*, an aquatic plant with three CCMs, to variable CO₂ and light. Journal of Experimental Botany **68**, 3985–3995.

Shen C, Dupont CL, Hopkinson BM. 2017. The diversity of carbon dioxide-concentrating mechanisms in marine diatoms as inferred from their genetic content. Journal of Experimental Botany **68**, 3937–3948.

Sommer M, Cai F, Melnicki M, Kerfeld CA. 2017. β-Carboxysome bioinformatics: identification and evolution of new bacterial microcompartment protein gene classes and core locus constraints. Journal of Experimental Botany **68**, 3841–3855.

Tcherkez GG, Farquhar GD, Andrews TJ. 2006. Despite slow catalysis and confused substrate specificity, all ribulose bisphosphate carboxylases may be nearly perfectly optimized. Proceedings of the National Academy of Sciences, USA 103, 7246–7251.

Tolleter D, Chochois V, Poiré R, Price GD, Badger MR. 2017. Measuring CO₂ and HCO₃⁻ permeabilities of isolated chloroplasts using a MIMS-¹⁸O approach. Journal of Experimental Botany **68**, 3915–3924.

Tsuji Y, Mahardika A, Matsuda Y. 2017. Evolutionarily distinct strategies for the acquisition of inorganic carbon from seawater in marine diatoms. Journal of Experimental Botany **68**, 3949–3958.

Tsuji Y, Nakajima K, Matsuda Y. 2017. Molecular aspects of the biophysical CO₂-concentrating mechanism and its regulation in marine diatoms. Journal of Experimental Botany **68**, 3763–3772.

Weiss RF. 1974. Carbon dioxide in water and seawater: the solubility of a non-ideal gas. Marine Chemistry 2, 203–215.

Young JN, Heureux AM, Sharwood RE, Rickaby RE, Morel FM, Whitney SM. 2016. Large variation in the Rubisco kinetics of diatoms reveals diversity among their carbon-concentrating mechanisms. Journal of Experimental Botany **67**, 3445–3456.

Young JN, Rickaby RE, Kapralov MV, Filatov DA. 2012. Adaptive signals in algal Rubisco reveal a history of ancient atmospheric carbon dioxide. Philosophical Transactions of the Royal Society of London. Series B, Biological Sciences **367**, 483–492.

Young JN, Hopkinson BM. 2017. The potential for co-evolution of CO₂-concentrating mechanisms and Rubisco in diatoms. Journal of Experimental Botany **68**, 3751–3762.