

Genetic susceptibility

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Synopsis

Why only 20% of smokers develop clinically relevant COPD was a puzzle for many years. Now, epidemiological studies point clearly towards a large heritable component. The combination of genome wide association studies and candidate gene analysis is helping to identify those genetic variants responsible for an individual's susceptibility to developing COPD. In this review, we will examine the current data implicating specific loci and genes in the pathogenesis of COPD.

Introduction

Although most smokers will die of a smoking-related disorder, only 20% suffer from significant COPD. Familial clustering suggests that heritable factors play an important role in the development of this disease ^{1,2}. In one large series, 18.6% of population-attributable risk for COPD could be accounted for by family history; patients with an affected parent having more severe disease, more frequent exacerbations and a worse quality of life ³. Similarly, in twin pairs the risk of developing COPD is higher for monozygotic than for dizygotic twins, with 60% of the individual susceptibility explained by genetic factors ⁴. Both the airway and emphysema components cluster independently suggesting that different genetic factors play a role in the development of these two components of the disease ⁵. Unsurprisingly, for a disorder with a substantial heritable component, race and ethnicity appear to impact upon the development of COPD. For example, in the COPDGene Study, 42% of affected African Americans were found to suffer severe early onset COPD (age < 55 years, FEV1 < 50% predicted) compared with 14% of non-Hispanic whites ⁶. In contrast, self-reported Hispanic ethnicity or Native American genetic ancestry have both been reported to be associated with significantly lower risks of developing COPD ⁷.

Efforts to identify the genetic determinants of COPD have evolved as the available technologies have changed. The analysis of candidate genes yielded some successes that will be discussed later, but that approach also led to numerous blind alleys, with initial excitement followed by disappointment as associations proved impossible to reproduce. The analysis of large cohorts of patients in genome-wide association studies (GWAS) using microarray technology to assay up

to a million single nucleotide polymorphisms (SNPs) in each case led to the unbiased identification of novel disease associated loci. This approach is hypothesis-free and so has the potential to open novel avenues of research.

Unbiased approaches

Until recently, only mutations of the *SERPINA1* gene that are responsible for α_1 -antitrypsin deficiency were unambiguously linked with the development of COPD. However, this disorder accounts for only 1-2% of cases of COPD and so other disease-associated alleles must exist. Recent large multinational GWAS have shed much light on this. In addition to validating the involvement of some candidate genes previously suspected of playing a role in the pathogenesis of COPD, these landmark studies have identified novel pathways that might plausibly lead to novel therapies for COPD.

SNPs in chromosome 15 at the α -nicotinic acetylcholine receptor *CHRNA3/5* locus (15q25.1; rs8034191 and rs1051730) were found to reach genome-wide significance and have subsequently been replicated in several independent studies⁸⁻¹³. This locus is significantly associated with pack-years of smoking, emphysema (by CT), and airflow obstruction^{9,14}. Notably, the C allele of the rs8034191 SNP was estimated to have a population attributable risk for COPD of 12.2% and has previously been identified in genome-wide association studies of lung cancer, being thought to be important in nicotine addiction¹⁵. Individuals who carry this SNP may require more cigarettes to satisfy nicotine addiction, may inhale more deeply and may find it more difficult to withdraw from cigarette smoking. Indeed, it has been reported that the association of the *CHRNA3/5* locus is substantially mediated by

smoking phenotype ¹², although this finding has been disputed ¹⁴. However, *IREB2* is a gene in tight linkage disequilibrium with *CHRNA3/5* and has also been identified as a potential determinant of COPD ^{9,16,17}. This gene encodes an iron regulatory protein localised in epithelia that may plausibly affect oxidative stress responses in smoked-exposed lungs. These candidate genes are not mutually exclusive and it remains possible that both genes within the haplotype contribute to the disease phenotype ¹².

GWAS have also identified a locus at 4q22.1 containing the gene *FAM13A* to be significantly associated with COPD and lung function in multiple cohorts ^{9,18}. While the function of *FAM13A* is unclear, another gene at 4q31 newly identified and replicated by GWAS as being associated with both COPD and lung function, encodes hedgehog interacting protein (HHIP), which appears to play a role in signalling that modulates lung development or remodelling ^{15,19-21}. HHIP is expressed in pulmonary tissues but at lower levels in COPD-affected lungs, and disease-associated SNPs have been identified within the gene's promoter (rs6537296A and rs1542725C) that appear to reduce its transcription ²². Other loci that have been identified using similar techniques include 2q35, 4q24, 5q33, 6p21, 15q23 and 19q13, although these require validation ^{20,23}. Other COPD-related phenotypes have also been linked with specific loci, for example low body mass index in COPD is significantly associated with SNP rs8050136 within the first intron of the fat mass and obesity-associated (*FTO*) gene ²⁴, while a SNP in *BICD1* (rs10844154 in 12p11.2) is associated with the presence and severity of emphysema on CT scan ²⁵.

Candidate gene approaches

The extra-cellular matrix

Alveolar tissue consists of epithelial cells, capillaries and extra-cellular matrix (ECM), the latter comprising a complex network of scaffolding proteins, principally elastin and collagen (Figure 1). The elastin filaments form from tropoelastin monomers that self assemble into aggregates and then fuse with microfilaments. Multiple covalent cross-links between the lysines in neighbouring filaments provide stability. Cutis laxa is a family of autosomal dominant (OMIM #123700), X-linked (OMIM #304150) and recessive (OMIM #219100, 219200) human diseases characterized by excessively slack connective tissues. Several families with the milder autosomal dominant form show early onset pulmonary pathology including emphysema²⁶, particularly if inherited with the Z allele of α_1 -antitrypsin²⁷. Two groups independently identified separate mutations within the ELN (elastin) gene that cause mild cutis laxa and early onset COPD^{28,29}. The ELN gene maps to 7q11.23 in man, but as chromosome 7 has not been identified in linkage analysis as a site associated with COPD, it is likely that ELN mutations are a rare cause of this disease.

Elastin fibres bind other proteins including fibulins, which in turn bind multiple ECM components and the basement membrane. The fibulins are a family of 6 proteins, at least 2 of which are mutated in severe autosomal recessive forms of cutis laxa and whose phenotype often includes early onset emphysema^{30,31}. A novel mutation in the fibulin-4 gene (FBLN4; 11q13) was recently identified in autosomal recessive cutis laxa with developmental emphysema³⁰. The mutation

caused an amino acid substitution in an epidermal growth factor (EGF) like domain of fibulin-4, leading to very low levels of extra-cellular protein. In a consanguineous Turkish family, a homozygous mutation in the related fibulin-5 gene (FBLN5; 14q32.1) was also found to cause cutis laxa and emphysema complicated by recurrent pulmonary infections³¹. Once again, the mutation was located within an EGF-like domain, suggesting these are critical for fibulins to maintain the integrity of the ECM within the lung. Interestingly, analogous mutations in fibrillin, which bares homology to the fibulins, cause Marfan's syndrome. Moreover, mutations of fibrillin (FBN1; 15q21.1) have been described in neonatal Marfan's with very early onset emphysema³²⁻³⁴.

Menkes disease (OMIM #309400) characterized by abnormal hair and dysmorphic features, is caused by mutations in an intracellular copper transporter (ATP7A; Xq13.3). The clinical features are due to defective connective tissue synthesis believed to be the result of dysfunction of lysyl oxidase. This copper-dependent enzyme is required for proper cross-linking of both collagen and elastin fibres. A recent case report described a child with Menkes disease and severe bilateral pan-lobular emphysema who died aged only 14 months³⁵. Gene sequencing revealed a splice-site mutation in ATP7A, suggesting that proper ECM cross-linking is vital for stability of the lung parenchyma.

In contrast to animal models of COPD, mutations in collagen have not been identified in humans. This does not appear to be due to an incompatibility of mutated collagen with survival, as numerous collagen mutations have been described that cause other human diseases. Instead, it may reflect a more important role for elastin integrity in emphysema in humans than in mice. However, aberrant

collagen synthesis has been implicated in COPD. The signaling molecule TGF β 1 enhances collagen synthesis in vivo, and polymorphisms in its gene (TGFB1; 19q13.1) have been associated with COPD ³⁶⁻⁴⁰, although a recent, large study found no association between TGFB1 polymorphisms and the rate of lung function decline in smokers ⁴¹. Intriguingly, the TGF β 1 gene maps to a locus on chromosome 19, which has high linkage (LOD 3.3) with FEV₁ in smokers ^{37,42}. However, as is frequently the case with polymorphism studies, the literature is unclear. For example, two TGFB1 single nucleotide polymorphisms (SNPs), rs1800469 and rs1982073, were found to be independently associated with COPD in two studies ^{36,39}, but in another, they were only significant when analysed as part of a haplotype (combination of alleles), while yet another SNP, rs6957, was significant in its own right ⁴⁰. Detailed analysis of the Boston Early-Onset COPD Study data revealed further complexity ³⁷. While some alleles of TGFB1 were associated with FEV₁ (rs2241712, rs2241718, rs6957), there was a separate but partially overlapping set of alleles associated with airflow obstruction (rs2241712, rs1800469, rs1982073). TGF β 1 protein is inactive when first secreted owing to the presence of an inhibitory N-terminal pro-peptide. It is secreted associated with latent TGF β 1 binding proteins (LTBP), which share structural features with fibrillins, and are assembled into the ECM. Mice with mutations in LTBP4 develop severe emphysema ⁴³. Intriguingly, the sole study that has addressed LTBP4 (19q13.1-q13.2) polymorphisms found an association with COPD in man ³⁹. More recently, genome-wide linkage analysis of pedigrees stratified by emphysema status (on CT scan) identified a region on chromosome 1p (LOD score = 2.99) ⁴⁴. An intronic SNP in TGFB-receptor-3 at this locus was found to be associated with COPD status, FEV₁ and CT emphysema.

Taken together, these studies provide strong evidence in support of a crucial role for the loss of ECM integrity, in particular the elastic components, in the development of COPD. It is therefore important to consider the enzymes implicated in degradation of the ECM.

Protease-antiprotease balance

The protease anti-protease theory has its roots in the observation that individuals with α_1 -antitrypsin are particularly susceptible to COPD and in experimental models of emphysema from the 1960s. This theory suggests that the pathogenesis of COPD and emphysema is the result of an imbalance between enzymes that degrade the ECM within the lung and proteins that oppose this proteolytic activity. Many proteases play important roles in remodeling or inflammation within the lung. It is essential that they be controlled by antiproteases to protect against uncontrolled degradation of the ECM (Figure 2).

The best-understood example of genetically induced emphysema results from mutations in the α_1 -antitrypsin gene (*SERPINA1*; 14q32.1). These increase the protein's propensity to form ordered polymers, which are incapable of inhibiting its target enzyme, neutrophil elastase. This abnormal behaviour leads to retention of the protein within hepatocytes as Periodic Acid Schiff positive inclusions and results in plasma deficiency of an important protease inhibitor (OMIM #107400). It is now increasingly recognised that mutant α_1 -antitrypsin can also form polymers within the interstitium and alveolar spaces of the lung. These polymers are chemotactic for neutrophils and so combine with the deficiency of α_1 -antitrypsin to focus and amplify the inflammatory response within the lung ⁴⁵. In most Northern European

populations the frequency of the most severe Z allele is about 1/2000. Classically, Z α_1 -antitrypsin homozygotes carry the Glu342Lys mutation and suffer from early onset emphysema when compared to normal MM α_1 -antitrypsin individuals. The onset and progression of emphysema is markedly accelerated by cigarette smoking. Moreover, it appears that even a single allele of Z α_1 -antitrypsin may increase the risk of COPD. In the longitudinal Copenhagen City Heart Study, the MZ α_1 -antitrypsin genotype increased the rate of decline of FEV₁ by 19% compared with those who were MM homozygotes, causing a 30% increased risk of obstructive lung function and a 50% increased risk of physician diagnosed COPD ⁴⁶. The authors found that the frequency of the MZ genotype in their Danish population was as high as 5% and so calculated that it would account for 2.4% of cases of COPD. This is in contrast to the ZZ genotype, which was causal in only 0.8% of cases. In meta-analysis, heterozygosity for the Z allele carried an odds ratio for COPD of 2.31 ⁴⁷. In one study, the MZ (but not MS; the S allele has the Glu264Val mutation) α_1 -antitrypsin genotype was associated with a rapid decline in FEV₁, which was even more marked if there was also a family history of COPD, suggesting an interaction with additional genetic factors ⁴⁸. A further meta-analysis combining 17 studies found a 3-fold increase in COPD in SZ α_1 -antitrypsin heterozygotes and a small increase in MS α_1 -antitrypsin heterozygotes. Other polymorphisms of the SERPINA1 gene do not appear to be associated with increased risk of developing COPD ⁴⁹.

Other pulmonary serine protease inhibitors may also be involved in the pathogenesis of COPD. Following earlier linkage studies demonstrating an association between chromosome 2q and COPD, expression profiling of genes

within that locus identified *SERPINE2* (2q33-q35) as being up regulated during murine lung development and in the lungs of individuals with COPD⁵⁰. The authors went on to demonstrate an association between SNPs in *SERPINE2* and COPD. *SERPINE2* SNPs were found to segregate with COPD in a large multi-centre family-based study and to be associated with COPD in a case-control analysis⁵¹. However another large study failed to replicate the association with COPD despite having adequate power⁵². The latter study included individuals with COPD with and without emphysema, while the studies by DeMeo and colleagues⁵⁰ included a preponderance of patients with emphysema assessed for lung volume reduction surgery. Nevertheless, while these differences may reflect different COPD phenotypes, they illustrate the need to replicate the findings of genetic association studies in multiple populations before drawing firm conclusions. A recent study of Finish construction workers found that three SNPs within *SERPINE2* (rs729631, rs975278, and rs6748795) were in tight linkage disequilibrium and so focused solely on one (rs729631)⁵³. This showed a significant association with panlobular emphysema, as seen with mutants of *SERPINA1*.

Since mutations of α_1 -antitrypsin so clearly lead to emphysema, one might infer that its target, neutrophil elastase, is central to the pathogenesis of disease. However, mutations in this protease have not been shown to be important, despite being studied extensively in other conditions. Instead, most evidence implicates matrix metalloproteases (MMPs) in the pathogenesis of COPD. These are zinc-dependent endopeptidases involved in the degradation of many ECM components. A SNP of MMP9 (20q11) was associated with COPD in Japanese⁵⁴ and Chinese⁵⁵ populations, however a further Japanese study found an association with

emphysema distribution rather than COPD *per se* ⁵⁶. Another large study failed to show MMP9 association with COPD, but instead MMP1 (11q22) and MMP12 (11q22) polymorphisms were identified ⁵⁷. Further support for a role for MMP9, but not MMP1 polymorphisms, has also been published ⁵⁸. Tissue Inhibitors of Metaloproteinases (TIMPs) inhibit the MMPs, but thus far, only one polymorphism in TIMP2 (17q25) has been associated with COPD ⁵⁹. When more than 8000 individuals were analysed, the minor allele of the promoter of MMP12 (rs2276109 [-82A-->G]) showed clear association with FEV₁ in a combined analysis of adult ever-smokers and children with asthma and with a reduced risk of the COPD ⁶⁰. In a separate study, a haplotype containing this SNP in MMP-12 (rs652438 and rs2276109) was found to be associated with severe COPD (GOLD Stages III and IV) (20078883). When expressed in cells *in vitro*, the COPD-associated A allele of rs652438 was 3-fold more proteolytically active than the G allele suggesting that it might mediate enhanced ECM degradation ⁶¹.

Reactive oxygen species

Cigarette smoke contains vast numbers of free radicals that impose an oxidative stress on the lung. Such stress is believed to induce damage through multiple mechanisms, including direct oxidation of cellular lipids and DNA, and through inactivation of key proteins such as α_1 -antitrypsin. For this reason, much work has gone in to assessing the role of endogenous antioxidant enzymes in protecting against smoke-induced lung damage.

Many toxins in cigarette smoke are subject to first pass metabolism in the liver. Amongst the many enzymes involved, microsomal epoxide hydrolase (EPHX1;

1q42.1) has been intensely studied in the context of COPD. Several EPHX1 SNPs have been described that affect its activity. One of these leads to a 40% loss of *in vitro* activity (rs1051740 Tyr113His, the “slow” allele), while another increases activity by 25% (rs2234922 His139Arg, the “fast” allele). In 1997, the “slow” variant of EPHX1 was found to increase the risk of emphysema by a staggering odds ratio of 5.0 and of COPD by an odds ratio of 4.1⁶². Since then, numerous studies have attempted to reproduce this effect with varying success^{39,63-74}. Recently, analysis of randomly selected white Danish individuals participating in the Copenhagen City Heart Study (n = 10,038) and the Copenhagen General Population Study (n = 37,022) for the rs1051740 and rs2234922 variants in the *EPHX1* gene combined with a meta-analysis of 19 previous studies indicate that genetically reduced EPHX1 activity is not a major risk factor for COPD or asthma in the Danish population.⁷⁵

Glutathione S-transferase (GST) comprises a large family of enzymes capable of catalyzing the conjugation of reduced glutathione to endogenous and xenobiotic electrophilic compounds. The GSTs are important in the detoxification of many compounds and are highly polymorphic. These polymorphisms have been linked to susceptibility to toxins and carcinogens. SNPs in GSTP1 have been associated with COPD⁷⁶, the distribution of emphysema⁷⁷ and more rapid decline in lung function⁷⁸. However, the data should be interpreted with caution as the third of the cohorts⁷⁸ has been used in multiple analyses⁶⁹ and there was a lack of Hardy-Weinberg equilibrium for GSTP1 in their population suggesting either a systematic defect in genotyping or an unidentified bias in the selection of subjects. Moreover, no convincing association was found in other studies^{71,79}. The null mutation of

GSTM1 (1p13.1) has also been associated with COPD ⁶⁴, but others have failed to reproduce this finding ⁷².

Heme oxygenase catalyses the first step in heme degradation. Heme oxygenase 1 (HMOX1; 22q13.1) is the inducible isoform that can be upregulated by a wide range of stresses. Bile pigments generated by heme cleavage are believed to have antioxidant properties, thus HMOX1 induction is protective during cellular oxidant injury and over-expression of HMOX1 in lung tissue protects against hyperoxia. The HMOX1 gene 5'-flanking region contains stretches of GC repeats that are highly polymorphic in length. An early report found a higher proportion of long repeats in patients with COPD and also demonstrated that long repeats were associated with impaired promoter activity ⁸⁰. Attempts to reproduce this effect have had varied success ^{65,78,81,82}. While HMOX1 GC-repeat length has not convincingly been shown to be associated with developing COPD, there are some data to support an association between the long allele and increased severity of disease ^{82,83}, although a recent study of smokers in the NHLBI Lung Health Study found no association between five HMOX1 SNPs and the decline of lung function ⁸⁴. Moreover, that study failed to detect evidence that the promoter polymorphisms affected regulation of the HMOX1 gene.

Superoxide dismutase (SOD) is an important antioxidant enzyme that catalyses the conversion of superoxide to oxygen and hydrogen peroxide. The extra-cellular isoform (SOD3; 4p15) is abundant in lung parenchyma. In the cross-sectional Copenhagen Heart Study, the R213G allele that results in higher plasma levels was associated with significantly less COPD in smokers ⁸⁵. A second study found similar results for the SOD3 isoenzyme, but not for other forms of SOD ⁸⁶.

Whilst biologically very plausible, current genetic evidence fails to provide clear support for the involvement of detoxifying enzymes in the pathogenesis of COPD. Since the potential list of candidates to detoxify cigarette smoke remains long, it would be preferable if future studies were to take an unbiased approach to target identification rather than studying small numbers of candidate genes.

Inflammation

Tumour necrosis factor- α (TNF; 6p21) is a multifunctional cytokine whose levels are elevated in bronchoalveolar lavage, induced sputa and biopsies from patients with COPD. It is a plausible candidate gene for susceptibility to inflammatory disease, especially as well-studied promoter polymorphisms clearly alter expression levels. Consequently, considerable effort has been invested into determining whether the promoter polymorphism in TNF α also predisposes smokers to COPD. Much interest was generated when an early study revealed an association (with a staggering odds ratio of over 10) between allele 2 and 'bronchitis' in Taiwanese men ⁸⁷. This study is difficult to interpret as a third of the men were 'never smokers'. Despite some supportive evidence ⁸⁸, many subsequent studies appeared to find little evidence that TNF polymorphisms are associated with, or modify the progression of COPD ^{69,74,89-98}.

Group specific component (GC; 4q12), also known as vitamin D binding globulin, is a multifunctional protein that enhances the neutrophil and monocyte chemotactic activity of complement component 5a. It is a highly polymorphic protein with more than 124 forms, although three, Gc*1F, Gc*1S and Gc2, make up the majority. Kueppers and colleagues found Gc2 homozygotes to be protected from

COPD⁹⁹. Others have seen this protective effect^{100,101}, while Gc*1F homozygosity has been found to be associated with COPD^{102,103}. However, a much larger recent study has failed to reproduce these associations¹⁰⁴.

Conclusion

While environmental exposure to smoke remains the preeminent risk factor for developing COPD, the evidence that heredity plays a major role in an individual's risk is clear. The combination of GWAS and carefully conducted candidate gene approaches is helping to tease out those genetic variants responsible for the familial clustering of this disease, offering both the personalisation of individual risk stratification and, more excitingly, the hope for rational therapeutic interventions based on a better understanding of the underlying molecular pathology. The confusion surrounding many of the early (and some current studies) lies almost entirely with study power. Apart from the notable exception of SERPINA1, the contribution of individual genetic variants to risk of disease will prove to be small, for this reason large stratified cohorts of well-phenotyped individuals are likely to prove invaluable. A recent large systematic review of all case control candidate genetic studies in COPD prior to 2008 concluded that although the majority of such studies were underpowered to detect small genetic effects (OR 1.2-1.5), four genetic variants (or the 27 for which adequate data were available) remained significantly associated with COPD: the GSTM1 null variant (OR 1.45), rs1800470 in TGFB1 (0.73), rs1800629 in TNF α (OR 1.19) and rs1799896 in SOD3 (OR 1.97)¹⁰⁵. Such findings, combined with the hypothesis generating observations from GWAS will direct COPD research for the next decade.

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References

1. Silverman EK, Chapman HA, Drazen JM, et al. Genetic epidemiology of severe, early-onset chronic obstructive pulmonary disease. Risk to relatives for airflow obstruction and chronic bronchitis. *Am J Respir Crit Care Med*. Jun 1998;157(6 Pt 1):1770-1778.
2. McCloskey SC, Patel BD, Hinchliffe SJ, Reid ED, Wareham NJ, Lomas DA. Siblings of patients with severe chronic obstructive pulmonary disease have a significant risk of airflow obstruction. *Am J Respir Crit Care Med*. Oct 15 2001;164(8 Pt 1):1419-1424.
3. Hersh CP, Hokanson JE, Lynch DA, et al. Family history is a risk factor for COPD. *Chest*. Aug 2011;140(2):343-350.
4. Ingebrigtsen T, Thomsen SF, Vestbo J, et al. Genetic influences on Chronic Obstructive Pulmonary Disease - a twin study. *Respiratory Medicine*. Dec 2010;104(12):1890-1895.
5. Patel BD, Coxson HO, Pillai SG, et al. Airway wall thickening and emphysema show independent familial aggregation in chronic obstructive pulmonary disease. *Am J Respir Crit Care Med*. Sep 1 2008;178(5):500-505.
6. Foreman MG, Zhang L, Murphy J, et al. Early-onset chronic obstructive pulmonary disease is associated with female sex, maternal factors, and African American race in the COPD Gene Study. *Am J Respir Crit Care Med*. Aug 15 2011;184(4):414-420.
7. Bruse S, Sood A, Petersen H, et al. New Mexican Hispanic smokers have lower odds of chronic obstructive pulmonary disease and less decline in lung function than non-Hispanic whites. *Am J Respir Crit Care Med*. Dec 1 2011;184(11):1254-1260.
8. Pillai SG, Ge D, Zhu G, et al. A genome-wide association study in chronic obstructive pulmonary disease (COPD): identification of two major susceptibility loci. *PLoS Genetics*. Mar 2009;5(3):e1000421.
9. Pillai SG, Kong X, Edwards LD, et al. Loci identified by genome-wide association studies influence different disease-related phenotypes in chronic obstructive pulmonary disease. *Am J Respir Crit Care Med*. Dec 15 2010;182(12):1498-1505.
10. Kaur-Knudsen D, Nordestgaard BG, Bojesen SE. CHRNA3 genotype, nicotine dependence, lung function and disease in the general population. *Eur Respir J*. Dec 2012;40(6):1538-1544.

11. Hardin M, Zielinski J, Wan ES, et al. CHRNA3/5, IREB2, and ADCY2 are associated with severe chronic obstructive pulmonary disease in Poland. *Am J Respir Crit Care Med*. Aug 2012;47(2):203-208.
12. Siedlinski M, Tingley D, Lipman PJ, et al. Dissecting direct and indirect genetic effects on chronic obstructive pulmonary disease (COPD) susceptibility. *Human Genetics*. Jan 9 2013.
13. Wilk JB, Shrine NR, Loehr LR, et al. Genome-wide association studies identify CHRNA5/3 and HTR4 in the development of airflow obstruction. *Am J Respir Crit Care Med*. Oct 1 2012;186(7):622-632.
14. Lambrechts D, Buyschaert I, Zanen P, et al. The 15q24/25 susceptibility variant for lung cancer and chronic obstructive pulmonary disease is associated with emphysema. *Am J Respir Crit Care Med*. Mar 1 2010;181(5):486-493.
15. Pillai SG, Ge D, Zhu G, et al. A genome-wide association study in chronic obstructive pulmonary disease (COPD): identification of two major susceptibility loci. *PLoS Genet*. Mar 2009;5(3):e1000421.
16. DeMeo DL, Mariani T, Bhattacharya S, et al. Integration of genomic and genetic approaches implicates IREB2 as a COPD susceptibility gene. *Am J Hum Genet*. Oct 2009;85(4):493-502.
17. Chappell SL, Daly L, Lotya J, et al. The role of IREB2 and transforming growth factor beta-1 genetic variants in COPD: a replication case-control study. *BMC medical genetics*. 2011;12:24.
18. Cho MH, Boutaoui N, Klanderman BJ, et al. Variants in FAM13A are associated with chronic obstructive pulmonary disease. *Nature Genetics*. Mar 2010;42(3):200-202.
19. Wilk JB, Chen TH, Gottlieb DJ, et al. A genome-wide association study of pulmonary function measures in the Framingham Heart Study. *PLoS Genetics*. Mar 2009;5(3):e1000429.
20. Repapi E, Sayers I, Wain LV, et al. Genome-wide association study identifies five loci associated with lung function. *Nature Genetics*. Jan 2010;42(1):36-44.
21. Van Durme YM, Eijgelsheim M, Joos GF, et al. Hedgehog-interacting protein is a COPD susceptibility gene: the Rotterdam Study. *Eur Respir J*. Jul 2010;36(1):89-95.
22. Zhou X, Baron RM, Hardin M, et al. Identification of a chronic obstructive pulmonary disease genetic determinant that regulates HHIP. *Hum Mol Genet*. Mar 15 2012;21(6):1325-1335.

23. Cho MH, Castaldi PJ, Wan ES, et al. A genome-wide association study of COPD identifies a susceptibility locus on chromosome 19q13. *Hum Mol Genet.* Feb 15 2012;21(4):947-957.
24. Wan ES, Cho MH, Boutaoui N, et al. Genome-wide association analysis of body mass in chronic obstructive pulmonary disease. *Am J Respir Cell Mol Biol.* Aug 2011;45(2):304-310.
25. Kong X, Cho MH, Anderson W, et al. Genome-wide association study identifies BICD1 as a susceptibility gene for emphysema. *Am J Respir Crit Care Med.* Jan 1 2011;183(1):43-49.
26. Callewaert B, Renard M, Huchtagowder V, et al. New insights into the pathogenesis of autosomal-dominant cutis laxa with report of five ELN mutations. *Human Mutation.* Apr 2011;32(4):445-455.
27. Corbett E, Glaisyer H, Chan C, Madden B, Khaghani A, Yacoub M. Congenital cutis laxa with a dominant inheritance and early onset emphysema. *Thorax.* Aug 1994;49(8):836-837.
28. Urban Z, Gao J, Pope FM, Davis EC. Autosomal dominant cutis laxa with severe lung disease: synthesis and matrix deposition of mutant tropoelastin. *J Invest Dermatol.* Jun 2005;124(6):1193-1199.
29. Kelleher CM, Silverman EK, Broekelmann T, et al. A functional mutation in the terminal exon of elastin in severe, early-onset chronic obstructive pulmonary disease. *Am J Respir Cell Mol Biol.* Oct 2005;33(4):355-362.
30. Huchtagowder V, Sausgruber N, Kim KH, Angle B, Marmorstein LY, Urban Z. Fibulin-4: a novel gene for an autosomal recessive cutis laxa syndrome. *Am J Hum Genet.* Jun 2006;78(6):1075-1080.
31. Loeys B, Van Maldergem L, Mortier G, et al. Homozygosity for a missense mutation in fibulin-5 (FBLN5) results in a severe form of cutis laxa. *Hum Mol Genet.* Sep 1 2002;11(18):2113-2118.
32. Revencu N, Quenum G, Demaille T, Verellen G, De Paepe A, Verellen-Dumoulin C. Congenital diaphragmatic eventration and bilateral uretero-hydronephrosis in a patient with neonatal Marfan syndrome caused by a mutation in exon 25 of the FBN1 gene and review of the literature. *Eur J Pediatr.* Jan 2004;163(1):33-37.
33. Shinawi M, Boileau C, Brik R, Mandel H, Bentur L. Splicing mutation in the fibrillin-1 gene associated with neonatal Marfan syndrome and severe pulmonary emphysema with tracheobronchomalacia. *Pediatr Pulmonol.* Apr 2005;39(4):374-378.

34. Tekin M, Cengiz FB, Ayberkin E, et al. Familial neonatal Marfan syndrome due to parental mosaicism of a missense mutation in the FBN1 gene. *Am J Med Genet A*. Apr 15 2007;143(8):875-880.
35. Grange DK, Kaler SG, Albers GM, Petterchak JA, Thorpe CM, DeMello DE. Severe bilateral panlobular emphysema and pulmonary arterial hypoplasia: unusual manifestations of Menkes disease. *Am J Med Genet A*. Dec 1 2005;139(2):151-155.
36. Wu L, Chau J, Young RP, et al. Transforming growth factor-beta1 genotype and susceptibility to chronic obstructive pulmonary disease. *Thorax*. Feb 2004;59(2):126-129.
37. Celedon JC, Lange C, Raby BA, et al. The transforming growth factor-beta1 (TGFB1) gene is associated with chronic obstructive pulmonary disease (COPD). *Hum Mol Genet*. Aug 1 2004;13(15):1649-1656.
38. Su ZG, Wen FQ, Feng YL, Xiao M, Wu XL. Transforming growth factor-beta1 gene polymorphisms associated with chronic obstructive pulmonary disease in Chinese population. *Acta Pharmacol Sin*. Jun 2005;26(6):714-720.
39. Hersh CP, Demeo DL, Lazarus R, et al. Genetic association analysis of functional impairment in chronic obstructive pulmonary disease. *Am J Respir Crit Care Med*. May 1 2006;173(9):977-984.
40. van Diemen CC, Postma DS, Vonk JM, Bruinenberg M, Nolte IM, Boezen HM. Decorin and TGF-beta1 polymorphisms and development of COPD in a general population. *Respir Res*. 2006;7:89.
41. Ogawa E, Ruan J, Connett JE, Anthonisen NR, Pare PD, Sandford AJ. Transforming growth factor-beta1 polymorphisms, airway responsiveness and lung function decline in smokers. *Respir Med*. May 2007;101(5):938-943.
42. Silverman EK, Palmer LJ, Mosley JD, et al. Genomewide linkage analysis of quantitative spirometric phenotypes in severe early-onset chronic obstructive pulmonary disease. *Am J Hum Genet*. May 2002;70(5):1229-1239.
43. Sterner-Kock A, Thorey IS, Koli K, et al. Disruption of the gene encoding the latent transforming growth factor-beta binding protein 4 (LTBP-4) causes abnormal lung development, cardiomyopathy, and colorectal cancer. *Genes Dev*. Sep 1 2002;16(17):2264-2273.
44. Hersh CP, Hansel NN, Barnes KC, et al. Transforming growth factor-beta receptor-3 is associated with pulmonary emphysema. *Am J Respir Cell Mol Biol*. Sep 2009;41(3):324-331.
45. Gooptu B, Lomas DA. Polymers and inflammation: disease mechanisms of the serpinopathies. *J Exp Med*. Jul 7 2008;205(7):1529-1534.

46. Dahl M, Tybjaerg-Hansen A, Lange P, Vestbo J, Nordestgaard BG. Change in lung function and morbidity from chronic obstructive pulmonary disease in alpha1-antitrypsin MZ heterozygotes: A longitudinal study of the general population. *Ann Intern Med*. Feb 19 2002;136(4):270-279.
47. Hersh CP, Dahl M, Ly NP, Berkey CS, Nordestgaard BG, Silverman EK. Chronic obstructive pulmonary disease in alpha1-antitrypsin PI MZ heterozygotes: a meta-analysis. *Thorax*. Oct 2004;59(10):843-849.
48. Sandford AJ, Weir TD, Spinelli JJ, Pare PD. Z and S mutations of the alpha1-antitrypsin gene and the risk of chronic obstructive pulmonary disease. *Am J Respir Cell Mol Biol*. Feb 1999;20(2):287-291.
49. Quint JK, Donaldson GC, Kumari M, Talmud PJ, Hurst JR. SERPINA1 11478G-->A variant, serum alpha1-antitrypsin, exacerbation frequency and FEV1 decline in COPD. *Thorax*. May 2011;66(5):418-424.
50. Demeo DL, Mariani TJ, Lange C, et al. The SERPINE2 gene is associated with chronic obstructive pulmonary disease. *Am J Hum Genet*. Feb 2006;78(2):253-264.
51. Zhu G, Warren L, Aponte J, et al. The SERPINE2 gene is associated with chronic obstructive pulmonary disease in two large populations. *Am J Respir Crit Care Med*. Jul 15 2007;176(2):167-173.
52. Chappell S, Daly L, Morgan K, et al. The SERPINE2 gene and chronic obstructive pulmonary disease. *Am J Hum Genet*. Jul 2006;79(1):184-186
53. Kukkonen MK, Tiili E, Hamalainen S, et al. SERPINE2 haplotype as a risk factor for panlobular type of emphysema. *BMC Medical Genetics*. 2011;12:157.
54. Minematsu N, Nakamura H, Tateno H, Nakajima T, Yamaguchi K. Genetic polymorphism in matrix metalloproteinase-9 and pulmonary emphysema. *Biochem Biophys Res Commun*. Nov 23 2001;289(1):116-119.
55. Zhou M, Huang SG, Wan HY, Li B, Deng WW, Li M. Genetic polymorphism in matrix metalloproteinase-9 and the susceptibility to chronic obstructive pulmonary disease in Han population of south China. *Chin Med J (Engl)*. Oct 2004;117(10):1481-1484.
56. Ito I, Nagai S, Handa T, et al. Matrix metalloproteinase-9 promoter polymorphism associated with upper lung dominant emphysema. *Am J Respir Crit Care Med*. Dec 1 2005;172(11):1378-1382.
57. Joos L, He JQ, Shepherdson MB, et al. The role of matrix metalloproteinase polymorphisms in the rate of decline in lung function. *Hum Mol Genet*. Mar 1 2002;11(5):569-576.

58. Tesfaigzi Y, Myers OB, Stidley CA, et al. Genotypes in matrix metalloproteinase 9 are a risk factor for COPD. *Int J Chron Obstruct Pulmon Dis*. 2006;1(3):267-278.
59. Hirano K, Sakamoto T, Uchida Y, et al. Tissue inhibitor of metalloproteinases-2 gene polymorphisms in chronic obstructive pulmonary disease. *Eur Respir J*. Nov 2001;18(5):748-752.
60. Hunninghake GM, Cho MH, Tesfaigzi Y, et al. MMP12, lung function, and COPD in high-risk populations. *N Engl J Med*. Dec 31 2009;361(27):2599-2608.
61. Haq I, Lowrey GE, Kalsheker N, Johnson SR. Matrix metalloproteinase-12 (MMP-12) SNP affects MMP activity, lung macrophage infiltration and protects against emphysema in COPD. *Thorax*. Nov 2011;66(11):970-976.
62. Smith CA, Harrison DJ. Association between polymorphism in gene for microsomal epoxide hydrolase and susceptibility to emphysema. *Lancet*. Aug 30 1997;350(9078):630-633.
63. Korytina GF, Ianbaeva DG, Viktorova TV. [Role of polymorphic variants of cytochrome P450 genes (CYP1A1, CYP2E1) and microsomal epoxide hydrolase (mEPHX) in pathogenesis of cystic fibrosis and chronic respiratory tract diseases]. *Mol Biol (Mosk)*. Sep-Oct 2003;37(5):784-792.
64. Cheng SL, Yu CJ, Chen CJ, Yang PC. Genetic polymorphism of epoxide hydrolase and glutathione S-transferase in COPD. *Eur Respir J*. Jun 2004;23(6):818-824.
65. Fu WP, Sun C, Dai LM, Yang LF, Zhang YP. Relationship between COPD and polymorphisms of HOX-1 and mEPH in a Chinese population. *Oncol Rep*. Feb 2007;17(2):483-488.
66. Hersh CP, Demeo DL, Lange C, et al. Attempted replication of reported chronic obstructive pulmonary disease candidate gene associations. *Am J Respir Cell Mol Biol*. Jul 2005;33(1):71-78.
67. Matheson MC, Raven J, Walters EH, Abramson MJ, Ellis JA. Microsomal epoxide hydrolase is not associated with COPD in a community-based sample. *Hum Biol*. Dec 2006;78(6):705-717.
68. Park JY, Chen L, Wadhwa N, Tockman MS. Polymorphisms for microsomal epoxide hydrolase and genetic susceptibility to COPD. *Int J Mol Med*. Mar 2005;15(3):443-448.
69. Sandford AJ, Chagani T, Weir TD, Connett JE, Anthonisen NR, Pare PD. Susceptibility genes for rapid decline of lung function in the lung health study. *Am J Respir Crit Care Med*. Feb 2001;163(2):469-473.

70. Takeyabu K, Yamaguchi E, Suzuki I, Nishimura M, Hizawa N, Kamakami Y. Gene polymorphism for microsomal epoxide hydrolase and susceptibility to emphysema in a Japanese population. *Eur Respir J*. May 2000;15(5):891-894.
71. Xiao D, Wang C, Du MJ, et al. Relationship between polymorphisms of genes encoding microsomal epoxide hydrolase and glutathione S-transferase P1 and chronic obstructive pulmonary disease. *Chin Med J (Engl)*. May 2004;117(5):661-667.
72. Yim JJ, Park GY, Lee CT, et al. Genetic susceptibility to chronic obstructive pulmonary disease in Koreans: combined analysis of polymorphic genotypes for microsomal epoxide hydrolase and glutathione S-transferase M1 and T1. *Thorax*. Feb 2000;55(2):121-125.
73. Yoshikawa M, Hiyama K, Ishioka S, Maeda H, Maeda A, Yamakido M. Microsomal epoxide hydrolase genotypes and chronic obstructive pulmonary disease in Japanese. *Int J Mol Med*. Jan 2000;5(1):49-53.
74. Brogger J, Steen VM, Eiken HG, Gulsvik A, Bakke P. Genetic association between COPD and polymorphisms in TNF, ADRB2 and EPHX1. *Eur Respir J*. Apr 2006;27(4):682-688.
75. Lee J, Nordestgaard BG, Dahl M. EPHX1 polymorphisms, COPD and asthma in 47,000 individuals and in meta-analysis. *Eur Respir J*. Jan 2011;37(1):18-25.
76. Ishii T, Matsuse T, Teramoto S, et al. Glutathione S-transferase P1 (GSTP1) polymorphism in patients with chronic obstructive pulmonary disease. *Thorax*. Aug 1999;54(8):693-696.
77. DeMeo DL, Hersh CP, Hoffman EA, et al. Genetic determinants of emphysema distribution in the national emphysema treatment trial. *Am J Respir Crit Care Med*. Jul 1 2007;176(1):42-48.
78. He JQ, Ruan J, Connett JE, Anthonisen NR, Pare PD, Sandford AJ. Antioxidant gene polymorphisms and susceptibility to a rapid decline in lung function in smokers. *Am J Respir Crit Care Med*. Aug 1 2002;166(3):323-328.
79. Rodriguez F, de la Roza C, Jardi R, Schaper M, Vidal R, Miravittles M. Glutathione S-transferase P1 and lung function in patients with alpha1-antitrypsin deficiency and COPD. *Chest*. May 2005;127(5):1537-1543.
80. Yamada N, Yamaya M, Okinaga S, et al. Microsatellite polymorphism in the heme oxygenase-1 gene promoter is associated with susceptibility to emphysema. *Am J Hum Genet*. Jan 2000;66(1):187-195.

81. Nakayama K, Kikuchi A, Yasuda H, et al. Heme oxygenase-1 gene promoter polymorphism and decline in lung function in Japanese men. *Thorax*. Oct 2006;61(10):921.
82. Budhi A, Hiyama K, Isobe T, et al. Genetic susceptibility for emphysematous changes of the lung in Japanese. *Int J Mol Med*. Mar 2003;11(3):321-329.
83. Fu WP, Zhao ZH, Fang LZ, et al. Heme oxygenase-1 polymorphism associated with severity of chronic obstructive pulmonary disease. *Chin Med J (Engl)*. Jan 5 2007;120(1):12-16.
84. Tanaka G, Aminuddin F, Akhabir L, et al. Effect of heme oxygenase-1 polymorphisms on lung function and gene expression. *BMC medical genetics*. 2011;12:117.
85. Juul K, Tybjaerg-Hansen A, Marklund S, Lange P, Nordestgaard BG. Genetically increased antioxidative protection and decreased chronic obstructive pulmonary disease. *Am J Respir Crit Care Med*. Apr 15 2006;173(8):858-864.
86. Young RP, Hopkins R, Black PN, et al. Functional variants of antioxidant genes in smokers with COPD and in those with normal lung function. *Thorax*. May 2006;61(5):394-399.
87. Huang SL, Su CH, Chang SC. Tumor necrosis factor-alpha gene polymorphism in chronic bronchitis. *Am J Respir Crit Care Med*. Nov 1997;156(5):1436-1439.
88. Cordoba-Lanus E, Baz-Davila R, de-Torres JP, et al. TNFA-863 polymorphism is associated with a reduced risk of chronic obstructive pulmonary disease: a replication study. *BMC Medical Genetics*. 2011;12:132.
89. Higham MA, Pride NB, Alikhan A, Morrell NW. Tumour necrosis factor-alpha gene promoter polymorphism in chronic obstructive pulmonary disease. *Eur Respir J*. Feb 2000;15(2):281-284.
90. Ishii T, Matsuse T, Teramoto S, et al. Neither IL-1beta, IL-1 receptor antagonist, nor TNF-alpha polymorphisms are associated with susceptibility to COPD. *Respir Med*. Sep 2000;94(9):847-851.
91. Ferrarotti I, Zorzetto M, Beccaria M, et al. Tumour necrosis factor family genes in a phenotype of COPD associated with emphysema. *Eur Respir J*. Mar 2003;21(3):444-449.
92. Patuzzo C, Gile LS, Zorzetto M, et al. Tumor necrosis factor gene complex in COPD and disseminated bronchiectasis. *Chest*. May 2000;117(5):1353-1358.

93. Seifart C, Plagens A, Dempfle A, et al. TNF-alpha, TNF-beta, IL-6, and IL-10 polymorphisms in patients with lung cancer. *Dis Markers*. 2005;21(3):157-165.
94. Chierakul N, Wongwisutikul P, Vejbaesya S, Chotvilaiwan K. Tumor necrosis factor-alpha gene promoter polymorphism is not associated with smoking-related COPD in Thailand. *Respirology*. Jan 2005;10(1):36-39.
95. Hegab AE, Sakamoto T, Saitoh W, et al. Polymorphisms of TNFalpha, IL1beta, and IL1RN genes in chronic obstructive pulmonary disease. *Biochem Biophys Res Commun*. Apr 22 2005;329(4):1246-1252.
96. Tanaka G, Sandford AJ, Burkett K, et al. Tumour necrosis factor and lymphotoxin A polymorphisms and lung function in smokers. *Eur Respir J*. Jan 2007;29(1):34-41.
97. Ruse CE, Hill MC, Tobin M, et al. Tumour necrosis factor gene complex polymorphisms in chronic obstructive pulmonary disease. *Respir Med*. Feb 2007;101(2):340-344.
98. Papatheodorou A, Latsi P, Vrettou C, et al. Development of a novel microarray methodology for the study of SNPs in the promoter region of the TNF-alpha gene: their association with obstructive pulmonary disease in Greek patients. *Clin Biochem*. Aug 2007;40(12):843-850.
99. Kueppers F, Miller RD, Gordon H, Hepper NG, Offord K. Familial prevalence of chronic obstructive pulmonary disease in a matched pair study. *Am J Med*. Sep 1977;63(3):336-342.
100. Horne SL, Cockcroft DW, Dosman JA. Possible protective effect against chronic obstructive airways disease by the GC2 allele. *Hum Hered*. 1990;40(3):173-176.
101. Schellenberg D, Pare PD, Weir TD, Spinelli JJ, Walker BA, Sandford AJ. Vitamin D binding protein variants and the risk of COPD. *Am J Respir Crit Care Med*. Mar 1998;157(3 Pt 1):957-961.
102. Ishii T, Keicho N, Teramoto S, et al. Association of Gc-globulin variation with susceptibility to COPD and diffuse panbronchiolitis. *Eur Respir J*. Nov 2001;18(5):753-757.
103. Ito I, Nagai S, Hoshino Y, et al. Risk and severity of COPD is associated with the group-specific component of serum globulin 1F allele. *Chest*. Jan 2004;125(1):63-70.
104. Kasuga I, Pare PD, Ruan J, Connett JE, Anthonisen NR, Sandford AJ. Lack of association of group specific component haplotypes with lung function in smokers. *Thorax*. Sep 2003;58(9):790-793.

- 105.** Castaldi PJ, Cho MH, Cohn M, et al. The COPD genetic association compendium: a comprehensive online database of COPD genetic associations. *Hum Mol Genet.* Feb 1 2010;19(3):526-534.

Figure 1. **Extracellular matrix.** The extracellular matrix is comprised primarily of collagen and elastin fibrils. When mutated, many of the components of these fibrils and enzymes involved in their assembly leads to true connective tissue disorders that are associated with premature emphysema. Human disorders linked to mutation of specific components are labelled in bold; those for which murine models exist are then given in non-bold text.

Figure 2. **Candidate COPD genes at the alveolus.** Toxic compounds in smoke are inhaled to the alveolus where some are detoxified. Many candidate modifier genes of COPD encode enzymes mediating this detoxification. According to the current *protease – anti-protease* model of smoke-induced lung damage, toxins that escape these protective mechanisms inactivate anti-proteases enabling the degradation of extracellular matrix. Variants of genes encoding anti-proteases, proteases, components of the extra-cellular matrix and signalling pathways that regulate regeneration have all been implicated in COPD.