# **Electronic Supplementary Information**

## **Tuning Riboflavin Derivatives for Photodynamic Inactivation of Pathogens**

Leander B. Crocker, Ju-Hyun Lee, Suraj Mital, Gabrielle C. Mills, Sina Schack, Andrea Bistrovic-Popov, Christoph O. Franck, Ioanna Mela, Clemens F. Kaminski, Graham Christie, Ljiljana Fruk\*

\* Department of Chemical Engineering and Biotechnology, University of Cambridge, Philippa Fawcett Drive, Cambridge, CB3 0AS, UK

E-mail: If389@cam.ac.uk

## 1 Table of Contents

1	Tał	Table of Contents		
2	Light Source			
3	Syr	4		
	3.1	Materials and Methods		
	3.2	Characterisation techniques		
	3.3	Flavins F1 and F2	5	
	3.4	Flavins F3 and F4		
4	Fluorescence Quantum Yield Calculations			
5	Singlet Oxygen Quantum Yield Calculations			
6	Fla	vin Photostability		
7	Bad	cterial Culture and Phototoxicity Assay		
	7.1	E. coli Log <sub>10</sub> Reductions		
	7.2	SIM Imaging		
8	Vir	$rology$ and $TCID_{50}$ Assay		
	8.1	MHV-A59 Log <sub>10</sub> Reductions		
9	Via	ability and Phototoxicity Assay		
1(	0 F	References		

### 2 Light Source

The light source used for experiments was an EvoluChem 6200K white LED (18W, 400-700 nm,  $\lambda_{max}$  = 445 nm, Hepatochem, US) with emission spectrum as shown in **Figure S1**. Illuminance was measured using an RS PRO IM720 Light Meter. Irradiance of emission was measured using a Thor Labs PM100D with a S120VC sensor.



Figure S1: Emission spectrum of EvoluChem 6200K white LED light source.

#### 3 Synthesis and Characterisation of Flavins

#### 3.1 Materials and Methods

All materials were purchased from either Fisher Scientific (UK), Acros Organics (UK), Alfa Aeser (UK), Sigma-Aldrich (UK) or TCI Chemicals (BE) in the highest purity available and used without further purification. For column chromatography 40-63 µm silica gel (Merck) was used as stationary phase. Analytical thin layer chromatography (TLC) was performed on aluminum foil pre-coated with SiO<sub>2</sub>-60 F254 (Merck) and visualized with a UV-lamp (254 and 365 nm).

#### 3.2 Characterisation techniques

<sup>1</sup>H and <sup>13</sup>C NMR measurements were carried out using a 400 MHz QNP Cryoprobe Spectrometer (Bruker) by the NMR service of the Department of Chemistry, University of Cambridge. The resonance multiplicity is abbreviated as: s (singlet), d (doublet), t (triplet), q (quadruplet), quint (quintet), sext (sextet), sep (septet), m (multiplet) and br (broad). The chemical shift  $\delta$  is expressed in "ppm" and the coupling constant J is in "Hz". The chemical shifts are referenced to the residual solvent peak as the internal standard. HRMS was recorded on a ThermoFinnigan Orbitrap Classic (Fisher Scientific) by the MS service of the Department of Chemistry, University of Cambridge. UV-Vis absorption spectra were obtained with an Agilent Cary 300 Spectrophotometer. Fluorescence emission spectra were obtained using a Varian Cary Eclipse Fluorescence Spectrophotometer using excitation and emission slits of 5 nm.

#### 3.3 Flavins F1 and F2



Flavin **1** was prepared as previously reported by Nehme *et al.*<sup>1</sup>

Scheme S1: Synthesis of dimethyl flavins F1 and F2.

*tert*-Butyl (2-(7,8-dimethyl-3-octyl-2,4-dioxo-3,4-dihydrobenzo[g]pteridin-10(2H)-yl)ethyl)carbamate (2):



Flavin **1** (0.50 g, 1.30 mmol, 1.0 equiv.)<sup>1</sup>, anhydrous  $K_2CO_3$  (0.27 g, 1.95 mmol, 1.5 equiv.) and anhydrous KI (21.5 mg, 0.130 mmol, 0.1 equiv.) were suspended in anhydrous DMF (50 mL) under Ar atmosphere. The mixture was then heated to 40 °C and 1-bromooctane (1.26 g, 1.12 mL, 6.50 mmol, 5.0 equiv.) dissolved in anhydrous DMF (5 mL) was added dropwise. The mixture was then stirred for 18 h at 40 °C under Ar atmosphere in the dark. The solvent was then removed under reduced pressure and the resulting residue was suspended in EtOAc (20 mL) and washed with water (3 x 20 mL) and brine (20 mL). The organic phase was then dried over  $Na_2SO_4$ , filtered, and concentrated under reduced pressure. The resulting orange residue was purified by silica gel column chromatography (1:2 EtOAc/DCM to 1:1 EtOAc/DCM) to yield flavin **2** as an orange solid (0.544 g, 70%).

<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ = 7.96 (s, 1H, H8), 7.84 (s, 1H, H10), 5.16 (br t, 1H, -NH-), 4.81 (t, J = 6.7 Hz, 2H, H14),
4.03 (t, J = 7.6 Hz, 2H, H13), 3.62 (br m, 2H, H15), 2.53 (s, 3H, H23), 2.41 (s, 3H, H24), 1.67 (quint, J = 7.4 Hz, 2H, H20), 1.39 (s, 9H, H19), 1.36-1.22 (m, 10H, H16/17/18/21/22), 0.84 (t, J = 6.9 Hz, 3H, H25) ppm.

<sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>) δ = 159.7 (C1), 156.5 (C11), 155.6 (C2), 148.7 (C3), 148.1 (C4), 136.7 (C5), 135.4 (C6), 134.9 (C7), 132.3 (C8), 131.5 (C9), 115.8 (C10), 80.0 (C12), 44.1 (C13), 42.1 (C14), 37.8 (C15), 31.8 (C16), 29.3 (C17), 29.2 (C18), 28.3 (C19), 27.8 (C20), 27.0 (C21), 22.6 (C22), 21.7 (C23), 19.5 (C24), 14.1 (C25) ppm.

**HRMS (ESI)** m/z:  $[M + H]^+$  Calcd for C<sub>27</sub>H<sub>40</sub>O<sub>4</sub>N<sub>5</sub> 498.3080; Found 498.3074.





2-(7,8-Dimethyl-3-octyl-2,4-dioxo-3,4-dihydrobenzo[g]pteridin-10(2H)-yl)ethan-1-aminium trifluoroacetate salt (F1):



Flavin **2** (244 mg, 0.49 mmol) was dissolved in DCM (5 mL) and TFA (2 mL) was added dropwise. The resulting mixture was stirred at RT for 2 h (reaction completion judged by TLC) before the solvent was removed under reduced pressure. Excess TFA was removed using toluene (20 mL) and DCM (3 x 20 mL) co-evaporation to yield the TFA salt of flavin **F1** as a yellow solid (248 mg, 99%).

<sup>1</sup>H NMR (400 MHz, DMSO-d<sub>6</sub>) δ = 8.01 (br s, 2H, -NH<sub>2</sub>), 7.95 (s, 1H, H10), 7.82 (s, 1H, H8), 4.87 (t, J = 6.3 Hz, 2H, H12), 3.87 (t, J = 7.4 Hz, 2H, H11), 3.23 (br m, 2H, H13), 2.50 (s, 3H, H20), 2.40 (s, 3H, H21), 1.56 (quint, J = 7.3 Hz, 2H, H17), 1.32-1.23 (m, 10H, H14/15/16/18/19), 0.85 (t, J = 6.8 Hz, 3H, H22) ppm.

<sup>13</sup>C NMR (100 MHz, DMSO-d<sub>6</sub>) δ = 159.4 (C1), 154.9 (C2), 149.8 (C3), 147.2 (C4), 136.5 (C5), 136.3 (C6), 134.3 (C7), 131.4 (C8), 130.6 (C9), 115.8 (C10), 41.2 (C11), 41.0 (C12), 36.7 (C13), 31.4 (C14), 28.9 (C15), 28.8 (C16), 27.5 (C17), 26.6 (C18), 22.2 (C19), 20.7 (C20), 18.9 (C21), 14.1 (C22) ppm.

HRMS (ESI) m/z: [M + H]<sup>+</sup> Calcd for C<sub>22</sub>H<sub>32</sub>O<sub>2</sub>N<sub>5</sub> 398.2556; Found 398.2564.



1,3-Di-Boc-2-(2-(7,8-dimethyl-3-octyl-2,4-dioxo-3,4-dihydrobenzo[g]pteridin-10(2H)-yl)ethyl)guanidine (3):



Flavin **2** (0.244 g, 0.49 mmol) was dissolved in DCM (5 mL) and TFA (2 mL) was added dropwise. The resulting mixture was stirred at RT for 2 h (reaction completion judged by TLC) before the solvent was removed under reduced pressure. Excess TFA was removed using toluene (3 x 20 mL) and DCM (3 x 20 mL) co-evaporation to yield flavin **1** which was then dissolved in anhydrous DMF (5 mL) under Ar atmosphere. DIPEA (0.17 mL, 0.98 mmol, 2.0 equiv.) was then added, followed by *N*,*N*'-di-Boc-1*H*-pyrazole-1-carboxamidine (152 mg, 0.49 mmol, 1.0 equiv). The mixture was then stirred for 18 h at RT under Ar atmosphere in the dark. The solvent was then removed under reduced pressure and the resulting residue was suspended in EtOAc (20 mL) and washed with water (3 x 20 mL) and brine (20 mL). The organic phase was then dried over Na<sub>2</sub>SO<sub>4</sub>, filtered, and concentrated under reduced pressure. The resulting yellow residue was purified by silica gel column chromatography (10% EtOAc/DCM) to yield flavin **3** as a yellow solid (282 mg, 90% over two steps).

<sup>1</sup>**H NMR** (400 MHz, CDCl<sub>3</sub>) δ = 11.42 (br s, 1H, -**NH**-), 8.72 (br s, 1H, -**NH**-), 8.00 (s, 1H, **H10**), 7.25 (s, 1H, **H8**), 4.93 (t, *J* = 7.0 Hz, 2H, **H12**), 4.07 (t, *J* = 7.5 Hz, 2H, **H11**), 3.80 (br m, 2H, **H13**), 2.58 (s, 3H, **H20**), 2.41 (s, 3H, **H21**), 1.70 (quint, *J* = 7.42 Hz, 2H, **H17**), 1.51 (s, 9H, **H28**), 1.45 (s, 9H, **H29**), 1.40-1.19 (m, 10H, **H14/15/16/18/19**), 0.85 (t, *J* = 6.9 Hz, 3H, **H22**) ppm.

<sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>) δ = 163.4 (C23), 159.8 (C1), 156.9 (C24), 155.7 (C2), 152.8 (C25), 148.7 (C3), 148.2 (C4), 136.6 (C5), 135.6 (C6), 134.8 (C7), 132.4 (C8), 131.3 (C9), 116.1 (C10), 83.7 (C26), 79.9 (C27), 42.2 (C11), 42.1 (C12), 37.2 (C13), 31.8 (C14), 29.4 (C15), 29.2 (C16), 28.4 (C28), 28.0 (C29), 27.8 (C17), 27.0 (C18), 22.7 (C19), 21.9 (C20), 19.5 (C21), 14.1 (C22) ppm.

**HRMS (ESI)** m/z:  $[M + H]^+$  Calcd for  $C_{33}H_{50}O_6N_7$  640.3823; Found 640.3824.



1-(2-(7,8-Dimethyl-3-octyl-2,4-dioxo-3,4-dihydrobenzo[g]pteridin-10(2H)-yl)ethyl)guanidinium trifluoroacetate salt (F2):



Flavin **3** (272 mg, 0.43 mmol) was dissolved in DCM (4 mL) and TFA (1.5 mL) was added dropwise. The resulting mixture was stirred at RT for 2 h (reaction completion judged by TLC) before the solvent was removed under reduced pressure. Excess TFA was removed using toluene (20 mL) and DCM (3 x 20 mL) co-evaporation to yield the TFA salt of flavin **F2** as a yellow solid (235 mg, 99%).

<sup>1</sup>**H NMR** (400 MHz, DMSO-d<sub>6</sub>) δ = 7.96 (s, 1H, **H8**), 7.81 (s, 1H, **H10**), 7.61 (br t, *J* = 6.2 Hz, 1H, -**NH**-) 4.71 (t, *J* = 6.3 Hz, 2H, **H12**), 4.48 (br s, 3H, =**NH**, -**NH**<sub>2</sub>), 3.88 (t, *J* = 7.4 Hz, 2H, **H11**), 3.59 (br m, 2H, **H13**), 2.50 (s, 3H, **H20**), 2.41 (s, 3H, **H21**), 1.58 (br m, *J* = 7.42 Hz, 2H, **H17**), 1.33-1.23 (m, 10H, **H14/15/16/18/19**), 0.86 (t, *J* = 6.8 Hz, 3H, **H22**) ppm.

<sup>13</sup>C NMR (100 MHz, DMSO-d<sub>6</sub>) δ = 159.4 (C1), 157.4 (C23), 154.7 (C2), 148.9 (C3), 147.0 (C4), 136.2 (C5, C6), 134.3 (C7), 131.2 (C8), 130.9 (C9), 115.9 (C10), 43.0 (C11), 41.0 (C12), 37.6 (C13), 31.3 (C14), 28.9 (C15), 28.7 (C16), 27.4 (C17), 26.5 (C18), 22.2 (C19), 20.7 (C20), 18.9 (C21), 14.0 (C22) ppm.

**HRMS (ESI)** m/z:  $[M + H]^+$  Calcd for C<sub>23</sub>H<sub>34</sub>O<sub>2</sub>N<sub>7</sub>440.2774; Found 440.2783.



#### 3.4 Flavins F3 and F4



1,2,4-Tribromo-5-nitrobenzene (4) was prepared as previously reported by Legrand  $et al.^2$ 

Scheme S2: Synthesis of flavins F3 and F4.

tert-Butyl (2-((4,5-dibromo-2-nitrophenyl)amino)ethyl)carbamate (5):



1,2,4-Tribromo-5-nitrobenzene (**4**) (1.00 g, 2.78 mmol, 1.0 equiv.) and anhydrous K<sub>2</sub>CO<sub>3</sub> (576 mg, 4.17 mmol, 1.5 equiv.) were suspended in anhydrous DMF (25 mL) before adding a solution of *tert*-butyl (2-aminoethyl)carbamate (1.70 g, 10.61 mmol, 5.0 equiv.) in anhydrous DMF (5 mL) dropwise at RT under Ar atmosphere. The resulting mixture was then stirred at 50 °C for 18 h. The solvent was then removed under reduced pressure and the resulting residue was suspended in EtOAc (20 mL) and washed with water (3 x 20 mL) and brine (20 mL). The organic phase was then dried over Na<sub>2</sub>SO<sub>4</sub>, filtered, and concentrated under reduced pressure. The resulting residue was purified by silica gel column chromatography (5-10% EtOAc/Cyclohexane) to yield **5** as an orange solid (785 mg, 64%).

<sup>1</sup>**H NMR** (400 MHz, CDCl<sub>3</sub>) δ = 8.37 (s, 1H, **H5**), 8.08 (br s, 1H, -**NH**-), 7.21 (s, 1H, H6), 4.83 (br s, 1H, Ar-**NH**-), 3.45-3.40 (br m, 4H, **H9**, **H10**), 1.45 (s, 9H, **H11**) ppm.

<sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>) δ = 156.2 (C1), 144.3 (C23), 134.1 (C2), 131.7 (C3), 130.6 (C4), 118.3 (C5), 109.4 (C6), 80.1 (C7), 43.5 (C8), 39.5 (C9), 28.3 (C10) ppm.

**HRMS (ESI)** m/z: [M + H]<sup>+</sup> Calcd for C<sub>13</sub>H<sub>17</sub>O<sub>4</sub>N<sub>3</sub>Br<sub>2</sub>Na 459.9484; Found 459.9470.



tert-Butyl (2-(7,8-dibromo-2,4-dioxo-3,4-dihydrobenzo[g]pteridin-10(2H)-yl)ethyl)carbamate (6):



*tert*-Butyl (2-((4,5-dibromo-2-nitrophenyl)amino)ethyl)carbamate (**5**) (785 mg, 1.78 mmol, 1.0 equiv.) was dissolved in MeOH/EtOAc (1:1 v/v, 20 mL) and system purged with Ar. Pd/C (50 mg) was then added and the atmosphere was replaced with H<sub>2</sub> gas (2 x balloons). The resulting mixture was stirred for 4 h (reaction completion judged by TLC) at RT before being filtered through Celite and concentrated under reduced pressure. The resulting red residue was then dissolved in glacial acetic acid (20 mL) under Ar atmosphere. Subsequently, B<sub>2</sub>O<sub>3</sub> (248 mg, 3.56 mmol, 2.0 equiv.) was added, followed by alloxan monohydrate (285 mg, 1.78 mmol, 1.0 equiv) and the resulting reaction mixture was stirred for 18 h under Ar atmosphere in the dark. The mixture was then partitioned between EtOAc (20 mL) and water (20 mL), and the organic phase was washed with brine (3 x 20 mL), dried over Na<sub>2</sub>SO<sub>4</sub>, filtered and concentrated under reduced pressure. The orange residue was then purified by silica gel column chromatography (1% EtOH/EtOAc) to yield flavin **6** as an orange solid (313 mg, 34% over two steps).

<sup>1</sup>**H NMR** (400 MHz, DMSO-d<sub>6</sub>) δ = 11.57 (s, 1H, Ar-NH), 8.52 (s, 1H, H7), 8.44 (s, 1H, H9), 7.00 (br t, *J* = 6.0 Hz, 1H, -NH-), 4.59 (t, *J* = 5.2 Hz, 2H, H11), 3.36 (br m, 2H, H12), 1.22 (s, 9H, H15) ppm.

<sup>13</sup>C NMR (100 MHz, DMSO-d<sub>6</sub>) δ = 159.4 (C1), 155.9 (C13), 155.4 (C2), 150.8 (C3), 139.7 (C4), 135.1 (C5), 134.6 (C6), 133.2 (C7), 130.9 (C8), 121.2 (C9), 120.6 (C10), 78.1 (C14), 44.6 (C11), 36.8 (C12), 28.0 (C15) ppm.

**HRMS (ESI)** m/z:  $[M + H]^+$  Calcd for  $C_{17}H_{18}O_4N_5Br_2$  513.9726; Found 513.9739.





tert-Butyl (2-(7,8-dibromo-3-octyl-2,4-dioxo-3,4-dihydrobenzo[g]pteridin-10(2H)-yl)ethyl)carbamate (7):



Flavin **6** (504 mg, 0.98 mmol, 1.0 equiv.) and 1-bromooctane (944 mg, 0.86 mL, 4.89 mmol, 5.0 equiv.) were dissolved in anhydrous DMF (20 mL) under Ar atmosphere. Cs<sub>2</sub>CO<sub>3</sub> (478 mg, 1.47 mmol, 1.5 equiv.) was then added and the resulting mixture was stirred for 18 h at RT under Ar in the dark. The solvent was then removed under reduced pressure and the resulting residue was suspended in DCM (20 mL) and washed with water (3 x 20 mL) and brine (20 mL). The organic phase was then dried over Na<sub>2</sub>SO<sub>4</sub>, filtered, and concentrated under reduced pressure. The resulting residue was purified by silica gel column chromatography (10-20% EtOAc/DCM) to yield flavin **7** as an orange solid (444 mg, 72%).

<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ = 8.35 (s, 2H, H7, H9), 5.27 (br t, J = 5.7 Hz, 1H, -NH-), 4.72 (t, J = 6.2 Hz, 2H, H12),
4.01 (t, J = 7.6 Hz, 2H, H11), 3.67 (q, J = 6.0 Hz, 2H, H13), 1.65 (quint, J = 7.4 Hz, 2H, H17), 1.40 (s, 9H, H23), 1.361.24 (m, 10H, H14/15/16/18/19), 0.85 (t, J = 6.8 Hz, 3H, H20) ppm.

<sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>) δ = 158.7 (C1), 156.6 (C21), 154.9 (C2), 148.6 (C3), 137.5 (C4), 135.8 (C5), 134.9 (C6), 133.7 (C7), 132.8 (C8), 122.8 (C9), 120.5 (C10), 80.5 (C22), 44.7 (C11), 42.4 (C12), 38.3 (C13), 31.8 (C14), 29.3 (C15), 29.2 (C16), 28.4 (C23), 27.7 (C17), 27.0 (C18), 22.6 (C19), 14.1 (C20) ppm.

**HRMS (ESI)** m/z:  $[M + H]^+$  Calcd for  $C_{25}H_{34}O_4N_5Br_2$  626.0978; Found 626.0980.





2-(7,8-Dibromo-3-octyl-2,4-dioxo-3,4-dihydrobenzo[g]pteridin-10(2H)-yl)ethan-1-aminium trifluoroacetate salt (F3):



Flavin **7** (212 mg, 0.34 mmol) was dissolved in DCM (4 mL) and TFA (1.5 mL) was added dropwise. The resulting mixture was stirred at RT for 2 h (reaction completion judged by TLC) before the solvent was removed under reduced pressure. Excess TFA was removed using toluene (20 mL) and DCM (3 x 20 mL) co-evaporation to yield the TFA salt of flavin **F3** as a yellow solid (215 mg, 99%).

<sup>1</sup>H NMR (400 MHz, DMSO-d<sub>6</sub>) δ = 8.61 (s, 1H, H7), 8.35 (s, 2H, H9), 7.93 (br m, *J* = 5.7 Hz, 2H, -NH<sub>2</sub>), 4.82 (t, *J* = 6.6 Hz, 2H, H12), 3.88 (t, *J* = 7.4 Hz, 2H, H11), 3.67 (br m, 2H, H13), 1.57 (quint, *J* = 7.3 Hz, 2H, H17), 1.30-1.23 (m, 10H, H14/15/16/18/19), 0.86 (t, *J* = 6.9 Hz, 3H, H20) ppm.

<sup>13</sup>C NMR (100 MHz, DMSO-d<sub>6</sub>) δ = 158.9 (C1), 154.8 (C2), 150.1 (C3), 139.3 (C4), 135.4 (C5), 135.0 (C6), 132.3 (C7), 131.6 (C8), 121.1 (C9), 120.8 (C10), 41.4 (C11), 41.3 (C12), 36.4 (C13), 31.4 (C14), 28.9 (C15), 28.8 (C16), 27.5 (C17), 26.6 (C18), 22.3 (C19), 14.1 (C20) ppm.

**HRMS (ESI)** m/z: [M + H]<sup>+</sup> Calcd for C<sub>20</sub>H<sub>26</sub>O<sub>2</sub>N<sub>5</sub>Br<sub>2</sub> 526.0453; Found 526.0455.





1,3-Di-Boc-2-(2-(7,8-dibromo-3-octyl-2,4-dioxo-3,4-dihydrobenzo[g]pteridin-10(2H)-yl)ethyl)guanidine (8):



Flavin **7** (270 mg, 0.43 mmol) was dissolved in DCM (4 mL) and TFA (1.5 mL) was added dropwise. The resulting mixture was stirred at RT for 2 h (reaction completion judged by TLC) before the solvent was removed under reduced pressure. Excess TFA was removed using toluene (3 x 20 mL) and DCM (3 x 20 mL) co-evaporation to yield flavin **3** which was then dissolved in anhydrous DMF (5 mL) under Ar atmosphere. DIPEA (0.15 mL, 0.86 mmol, 2.0 equiv.) was then added, followed by *N*,*N*'-di-Boc-1*H*-pyrazole-1-carboxamidine (133 mg, 0.43 mmol, 1.0 equiv). The mixture was then stirred for 18 h at RT under Ar atmosphere in the dark. The solvent was then removed under reduced pressure and the resulting residue was suspended in DCM (20 mL) and washed with water (3 x 20 mL) and brine (20 mL). The organic phase was then dried over Na<sub>2</sub>SO<sub>4</sub>, filtered, and concentrated under reduced pressure. The resulting residue was purified by silica gel column chromatography (10% EtOAc/DCM) to yield flavin **8** as an orange solid (264 mg, 80% over two steps).

<sup>1</sup>**H NMR** (400 MHz, CDCl<sub>3</sub>) δ = 11.3 (br s, 1H, -NH-), 8.57 (br t, *J* = 5.8 Hz 1H, -NH-), 8.45 (s, 1H, H7), 8.24 (s, 1H, H9), 4.88 (t, *J* = 6.0 Hz, 2H, H12), 4.04 (t, *J* = 7.6 Hz, 2H, H11), 3.88 (q, *J* = 6.1 Hz, 2H, H13), 1.68 (quint, *J* = 7.5 Hz, 2H, H17), 1.46 (s, 9H, H26), 1.43 (s, 9H, H27), 1.40-1.21 (m, 10H, H14/15/16/18/19), 0.85 (t, *J* = 7.0 Hz, 3H, H22) ppm.

<sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>) δ = 162.9 (C21), 158.8 (C1), 156.8 (C23), 155.0 (C2), 152.8 (C22), 148.9 (C3), 137.9 (C4), 136.2 (C5), 134.9 (C6), 133.0 (C7), 132.7 (C8), 122.2 (C9), 120.4 (C10), 83.8 (C24), 79.9 (C25), 43.6 (C11), 42.3 (C12), 37.6 (C13), 31.8 (C14), 29.3 (C15), 29.2 (C16), 28.3 (C26), 28.0 (C27), 27.7 (C17), 26.9 (C18), 22.6 (C19), 14.1 (C20) ppm.

**HRMS (ESI)** m/z:  $[M + H]^+$  Calcd for  $C_{31}H_{44}O_6N_7Br_2$  768.1720; Found 768.1720.





1-(2-(7,8-Dibromo-3-octyl-2,4-dioxo-3,4-dihydrobenzo[g]pteridin-10(2H)-yl)ethyl)guanidinium trifluoroacetate (F4):



Flavin **8** (264 mg, 0.34 mmol) was dissolved in DCM (4 mL) and TFA (1.5 mL) was added dropwise. The resulting mixture was stirred at RT for 2 h (reaction completion judged by TLC) before the solvent was removed under reduced pressure. Excess TFA was removed using toluene (20 mL) and DCM (3 x 20 mL) co-evaporation to yield the TFA salt of flavin **F4** as a yellow solid (232 mg, 99%).

<sup>1</sup>**H NMR** (400 MHz, DMSO-d<sub>6</sub>) δ = 8.61 (s, 1H, **H7**), 8.42 (s, 1H, **H9**), 7.43 (br t, *J* = 6.3 Hz, 1H, -**NH**-) 4.71 (t, *J* = 6.0 Hz, 2H, **H12**), 3.87 (br s, 3H, =**NH**, -**NH**<sub>2</sub>), 3.57 (t, *J* = 6.1 Hz, 2H, **H11**), 3.59 (br m, 2H, **H13**), 2.50 (s, 3H, **H20**), 2.41 (s, 3H, **H21**), 1.58 (br m, *J* = 7.42 Hz, 2H, **H17**), 1.33-1.23 (m, 10H, **H14/15/16/18/19**), 0.86 (t, *J* = 6.7 Hz, 3H, **H22**) ppm.

<sup>13</sup>C NMR (100 MHz, DMSO-d<sub>6</sub>) δ = 158.9 (C1), 157.3 (C21), 154.6 (C2), 149.5 (C3), 139.1 (C4), 135.2 (C5), 134.9 (C6), 132.4 (C7), 131.3 (C8), 121.0 (C9), 120.9 (C10), 43.2 (C11), 41.1 (C12), 37.5 (C13), 31.3 (C14), 28.8 (C15), 28.7 (C16), 27.3 (C17), 26.5 (C18), 22.2 (C19), 14.0 (C20) ppm.

**HRMS (ESI)** m/z:  $[M + H]^+$  Calcd for  $C_{21}H_{28}O_2N_7Br_2$  568.0671; Found 568.0669.



#### 4 Fluorescence Quantum Yield Calculations

The fluorescence quantum yields ( $\Phi_F$ ) for **F1-4** were determined by preparing a series of dilutions in DMSO ranging in absorbance values from 0.02 to 0.10 and excited at their respective absorbance maxima (see **Table S1**).<sup>3</sup> The emission peak was then integrated and plotted against the absorbance value (see **Figure S1**). Riboflavin (from *Eremothecium ashbyii*, ≥98%, Sigma) was chosen as the reference  $\phi_F$  for calculations ( $\Phi_F$ = 0.226 ± 0.01 in DMSO)<sup>4</sup> according to equation (1):

$$\Phi_{F;Sample} = \Phi_{F;Ref} \left( \frac{Slope_{Sample}}{Slope_{Ref}} \right) \left( \frac{\eta_{Sample}^2}{\eta_{Ref}^2} \right) (1)$$

Where Slope = gradient from the plot of integrated emission peak vs. absorption value and  $\eta$  = refractive index of the solvent.

**Table S1:** Spectroscopic details of flavins **F1-4** and reference compound riboflavin used for  $\phi_F$  calculations in DMSO.

Flavin	ε (M <sup>-1</sup> cm <sup>-1</sup> )	λ <sub>ex</sub> (nm)	λ <sub>em</sub> (nm)
Riboflavin	11718.41	447	513
F1	10368.79	444	507
F2	9838.05	446	512
F3	9541.26	443	508
F4	10208.50	444	513



**Figure S2:** Plot of integrated emission peak against absorbance value of flavins **F1-4** and reference compound riboflavin in DMSO.

#### 5 Singlet Oxygen Quantum Yield Calculations

The singlet oxygen quantum yield ( $\Phi_{\Delta}$ ) of flavins **F1-4** were calculated using the singlet oxygen probe, 1,3diphenylisobenzofuran (DPBF, 97%, Acros Organics, UK) according to the guidelines published by Bresolí-Obach *et al.*<sup>5</sup> The light source used was a 450 nm LED (18 W, HepatoChem, US) with the light output attenuated using a 2" x 2" Absorptive Neutral Density Filter (OD 3.0, Thor Labs, UK) as to avoid considerable self-bleaching of DPBF (blank control). Ru(bpy)<sub>3</sub><sup>2+</sup> (dichloro hexahydrate, 99.95%, Sigma-Aldrich, UK) was chosen as the reference  $\phi_{\Delta}$  for calculations ( $\Phi_{\Delta} = 0.57 \pm 0.06$  in MeCN)<sup>6</sup> according to equation (2):

$$\Phi_{\Delta;\text{Sample}} = \Phi_{\Delta;\text{Ref}} \frac{(\text{Slope}_{\text{Sample}} - \text{Slope}_{\text{Control}}) \cdot (1 - 10^{-A_{\text{Ref}}})}{(\text{Slope}_{\text{Ref}} - \text{Slope}_{\text{Control}}) \cdot (1 - 10^{-A_{\text{Sample}}})} \left(\frac{\eta_{\text{Sample}}^2}{\eta_{\text{Ref}}^2}\right) (2)$$

Where Slope = rate of DPBF photobleaching, Sample = flavin **F1-4**, Control = DPBF alone, Ref = Ru(bpy)<sub>3</sub><sup>2+</sup>, A = absorbance value at irradiation wavelength (450 nm) and  $\eta$  = refractive index of the solvent.

**General procedure:** DPBF (50  $\mu$ M) was irradiated with stirring at 450 nm in the presence of flavin (20  $\mu$ M) or Ru(bpy)<sub>3</sub><sup>2+</sup> (15  $\mu$ M) in MeCN (3 mL) at RT with the reaction vessel open to air. Aliquots of the reaction (100  $\mu$ L) were taken every 30 s for 3 min and measured by UV-Vis spectroscopy. The decrease of the  $\lambda_{max}$  of DBPF (410 nm) was then evaluated and plot according to A/A<sub>0</sub> vs. time as shown below (**Figure S2**). Experiments were carried out three separate times using fresh stock solutions of the reagents. The experimental error was estimated by combining the  $\Phi_{\Delta;Ref}$  error and the standard deviation of the average  $\Phi_{\Delta}$  for the flavin samples.



**Figure S3:** Plot of DPBF bleaching in the presence of flavins **F1-4** or reference compound  $Ru(bpy)_{3}^{2+}$  under 450 nm irradiation in MeCN. Blank = DPBF alone with no photosensitiser.



**Figure S4:** UV-Vis absorption spectra of DPBF bleaching in the presence of flavins **F1-4** under 450 nm irradiation in MeCN.



**Figure S4**: (A) Flavin photostability (100  $\mu$ M) in PBS (0.1% DMSO) irradiated with 1x10<sup>5</sup> lx (35 mW/cm<sup>2</sup>) where A = Abs at 450 nm and A<sub>0</sub> = Abs at 450 nm at 0 min. (B-F) Associated UV-Vis spectra of riboflavin and **F1-4** photostability. Experiments were performed using a quartz cuvette and measured using an Agilent Cary 300 Spectrophotometer.

#### 7 Bacterial Culture and Phototoxicity Assay



**Figure S5:** Photodynamic inactivation of *E. coli* incubated with 100  $\mu$ M of flavin compound irradiated with white LED light (1x10<sup>5</sup> lx, 35 mW/cm<sup>2</sup>) or incubated in the dark for 15 min in PBS (0.5% DMSO). Data are expressed as the mean ± SD of three biological replicates. Significance levels are defined as the following: ns for p>0.05, \* for p≤0.05, \*\* for p<0.01, \*\*\* for p<0.001, and \*\*\*\* for p<0.001.

#### E. coli Log<sub>10</sub> Reductions 7.1

Flavin	Irradiation time (min)	Concentration (µM)	Log <sub>10</sub> reduction <sup>[a]</sup>
	15	1	0.2
	15	10	0.2
Riboflavin	5	100	0.0
	10	100	0.0
	15	100	0.0
	15	1	0.2
	15	10	0.2
F1	5	100	0.1
	10	100	0.1
	15	100	0.1
	15	1	0.2
	15	10	0.8
F2	5	100	3.4
	10	100	5.9
	15	100	7.0
	15	1	0.2
	15	10	0.4
F3	5	100	1.3
	10	100	1.7
	15	100	1.8
	15	1	0.2
	15	10	2.8
	1	100	4.1 <sup>b</sup>
	2	100	5.8 <sup>b</sup>
F4	3	100	6.4 <sup>b</sup>
	4	100	7.6 <sup>b</sup>
	5	100	8.4
	10	100	8.7
	15	100	9.1

Table S2: Table of *E. coli* log<sub>10</sub> reductions in the presence of flavin under irradiation.

<sup>[a]</sup> calculated by  $\log_{10}(\text{control}) - \log_{10}(\text{sample})$  at respective irradiation time  $^{[b]}$  calculated by  $\log_{10}(t_0) - \log_{10}(t_{sample})$  where  $t_0 = 7.9$ 

#### 7.2 SIM Imaging

Structured illumination microscopy (SIM) of *E. coli* BL21(DE3) was performed as described previously by Mela *et* al.<sup>7</sup> In detail, a culture of *E. coli* BL21(DE3) was grown overnight at an OD<sub>600</sub> of ~1. Prior to SIM imaging the cultures were centrifuged at 3000 rpm for 5 minutes and resuspended in PBS (1x. pH 7.4) to an OD<sub>600</sub> of around 0.5. The *E. coli* suspension was incubated for 1 hour with 100  $\mu$ M of each one of the flavins (riboflavin, **F1** or **F2** in PBS). The bacteria were then centrifuged at the same settings and the bacterial pellet resuspended in PBS (1x. pH 7.4) to remove excess flavin. 2  $\mu$ l of the sample were deposited on a glass coverslip and an agarose pad was positioned over the sample to prevent the bacteria from moving during imaging. Another coverslip was positioned on top to minimise drying of the agarose pads.

Images of the sample were collected using 3-color SIM for optical sectioning. A ×60/1.2 NA water immersion lens (UPLSAPO 60XW, Olympus) focused the structured illumination pattern onto the sample, and the same lens was also used to capture the fluorescence emission light before imaging onto an sCMOS camera (C11440, Hamamatsu). The wavelength used for excitation was 488 nm (iBEAM-SMART-488, Toptica). Images were acquired using custom SIM software described previously.<sup>8</sup>



**Figure S6:** Large field of view (42.8 x 42.8  $\mu$ m) of *E. coli* after incubation with **F2** (100  $\mu$ M).



Figure S7: Large field of view (42.8 x 42.8  $\mu$ m) of *E. coli* after incubation with F1 (100  $\mu$ M).



**Figure S8:** Large field of view (42.8 x 42.8 μm) of *E. coli* after incubation with riboflavin (100 μM).

#### **Gel Electrophoresis**

Agarose gel electrophoresis of pDNA (pUC18) following incubation with 10  $\mu$ M flavin irradiated with white LED light (1x10<sup>5</sup> lx, 35 mW/cm<sup>2</sup>) for 15 min in PBS containing 0.1% DMSO. Rbf = Riboflavin.



Figure F9: Uncropped agarose gel demonstrating the DNA cleavage as shown in Fig.3d.

Virology and TCID<sub>50</sub> Assay

#### 7.3 MHV-A59 Log<sub>10</sub> Reductions

Flavin	Irradiation Concentration time (min) (µM)		Log <sub>10</sub> reduction <sup>[a]</sup>
	10	1	1.2
	10	5	3.2
Riboflavin	5	10	2.7
	10	10	3.4
	15	10	3.7
	10	1	0.0
	10	5	1.7
F1	5	10	1.2
	10	10	2.1
	15	10	2.2
	10	1	2.1
	10	5	3.9
F2	5	10	3.0
	10	10	3.8
	15	10	5.0
	10	1	2.1
	10	5	4.1
F3	5	10	3.9
	10	10	4.8
	15	10	5.4
	10	1	3.4
	10	5	4.6
F4	5	10	3.8
	10	10	5.9
	15	10	6.7

Table S3: Table of MHV-A59 log<sub>10</sub> reductions in the presence of flavin under irradiation.

 $^{\rm [a]}$  calculated by  $\log_{10}({\rm control})$  -  $\log_{10}({\rm sample})$  at respective irradiation time

#### Gel electrophoresis of ssRNA cleavage



Figure S10: Uncropped gel electrophoresis image of ssRNA cleavage shown in Fig. 4d.

### 8 Viability and Phototoxicity Assay



**Figure S11**. *In vitro* cytotoxicity effect of **F1–F4** and riboflavin on 17Cl-1 cells after 24 h incubation determined by MTS assay. Data are expressed as the mean ± SD of three biological replicates.

Table S4: IC<sub>50</sub> and cLogP values for flavins F1-4 and riboflavin.



<b>F</b> 1. 1.	x	R	IC₅₀ (μM)ª			
Flavin			WI-38	17Cl-1	cLogP <sup>b</sup>	
Riboflavin	Me	Ribityl chain	>100	>100	-2.07	
F1	Me	* NH <sub>3</sub>	96.1	70.2	-0.55	
F2	Me		>100	>100	2.59	
F3	Br	**************************************	30.9	3.66	0.22	
F4	Br	NH <sub>2</sub> NH <sub>2</sub> NH <sub>2</sub>	>100	50.9	3.36	

<sup>a</sup> determined after 24 h incubation by MTS assay

<sup>b</sup> determined using DataWarrior V5.5.0

### 9 References

- 1. Nehme, S. I., Crocker, L. & Fruk, L. Flavin-conjugated iron oxide nanoparticles as enzyme-inspired photocatalysts for Azo Dye degradation. *Catalysts* **10**, 324 (2020).
- Legrand, Y. M., Gray, M., Cooke, G. & Rotello, V. M. Model Systems for Flavoenzyme Activity: Relationships between Cofactor Structure, Binding and Redox Properties. *J. Am. Chem. Soc.* 125, 15789– 15795 (2003).
- Pokluda, A. *et al.* Robust Photocatalytic Method Using Ethylene-Bridged Flavinium Salts for the Aerobic
   Oxidation of Unactivated Benzylic Substrates. *Adv. Synth. Catal.* 363, 4371–4379 (2021).
- 4. Drössler, P., Holzer, W., Penzkofer, A. & Hegemann, P. Fluoresence quenching of riboflavin in aqueous solution by methionin and cystein. *Chem. Phys.* **286**, 409–420 (2003).
- Bresolí-Obach, R., Torra, J., Zanocco, R. P., Zanocco, A. L. & Nonell, S. Singlet oxygen quantum yield determination using chemical acceptors. in *Methods in Molecular Biology* vol. 2202 165–188 (Humana Press Inc., 2021).
- Abdel-Shafi, A. A., Beer, P. D., Mortimer, R. J. & Wilkinson, F. Photosensitized generation of singlet oxygen from ruthenium(II)-substituted benzoaza-crown-bipyridine complexes. *Phys. Chem. Chem. Phys.* 2, 3137–3144 (2000).
- 7. Mela, I. *et al.* DNA Nanostructures for Targeted Antimicrobial Delivery. *Angew. Chemie* **132**, 12798–12802 (2020).
- 8. Young, L. J., Ströhl, F. & Kaminski, C. F. A guide to structured illumination TIRF microscopy at high speed with multiple colors. *J. Vis. Exp.* **2016**, e53988 (2016).