Invited Review: Climate change impacts in polar-regions: lessons

2 from Antarctic moss bank archives

- 3 Running head: Lessons from Antarctic moss banks archives
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- 11 **Keywords**: Stable isotopes; climate change; Antarctic moss; radiocarbon dating; peat
- accumulation; assimilation

13 Abstract:

- 14 Mosses are the dominant plants in polar and boreal regions, areas which are experiencing
- rapid impacts of regional warming. Long-term monitoring programmes provide some records
- of the rate of recent climate change, but moss peat banks contain an unrivalled temporal
- 17 record of past climate change in terrestrial plant Antarctic systems. We summarise the current
- 18 understanding of climatic proxies and determinants of moss growth for contrasting
- 19 continental and maritime Antarctic regions, as informed by ¹³C and ¹⁸O signals in organic
- 20 material. Rates of moss accumulation are more than three times higher in the maritime
- 21 Antarctic than continental Antarctica with growing season length being a critical determinant

of growth rate, and high carbon isotope discrimination values reflecting optimal hydration conditions. Correlation plots of ¹³C and ¹⁸O values show that species (*Chorisodontium aciphyllum / Polytrichum strictum*) and growth form (hummock / bank) are the major determinants of measured isotope ratios. The interplays between moss growth form, photosynthetic physiology, water status and isotope composition are compared with developments of secondary proxies, such as chlorophyll fluorescence. These approaches provide a framework to consider the potential impact of climate change on terrestrial Antarctic habitats as well as having implications for future studies of temperate, boreal and Arctic peatlands. There are many urgent ecological and environmental problems in the Arctic related to mosses in a changing climate, but the geographical ranges of species and life-forms are difficult to track individually. Our goal was to translate what we have learned from the more simple systems in Antarctica, for application to Arctic habitats.

1. Introduction

Global climate is changing, with heterogeneous effects on the biological world including direct impacts on plant phenology. The resilience and future dynamics of ecosystems will depend on their responsiveness to gradual environmental change, as well as susceptibility to more frequent climatic extremes (e.g. Orsenigo *et al.*, 2014, Xu *et al.*, 2013). High latitude regions are experiencing particularly rapid climatic changes, and bryophytes, specifically mosses, are the dominant plants in both the Arctic and the Antarctic. Wetlands cover 70% of the Arctic, largely made up of *Sphagnum* dominated peatlands that are shaped by freeze-thaw processes and may be minerotrophic fens or ombrotrophic bogs (Minayeva & Sirin, 2010). In the Antarctic, over one hundred species of moss have been identified (Ochyra *et al.*, 2008), with two native vascular plant species. Most Antarctic mosses form occasional low growing

- carpets, mats, turfs and hummocks, whilst some ombrotrophic peat banks up to three metres
- deep have accumulated in more maritime areas.
- 48 Global peatlands contain approximately 600 Pg of stored carbon (Yu et al., 2010) but they
- 49 are under threat across high latitude regions (Gallego-Sala & Prentice, 2013), with
- 50 potentially significant impacts on the global carbon cycle (McGuire et al., 2009). Therefore,
- 51 incorporating moss dynamics into vegetation models is essential (Turetsky et al., 2012) and
- 52 understanding the bryophytes within the simple ecosystem context of Antarctica, in
- 53 combination with contemporary bryophyte physiology, can provide important lessons
- 54 applicable to more complex Arctic habitats.
- 55 This review firstly summarizes the climate, vegetation and growth dynamics of Antarctic
- 56 moss-peat accumulations before considering the current understanding of climatic proxies,
- 57 particularly focusing on the use of stable isotope analysis of preserved organic material, and
- contrasting responses between continental and maritime regions. The interplay between moss
- 59 photosynthetic physiology, water status and growth-form are compared with recent
- developments of secondary proxies (such as ¹³C and ¹⁸O in organic material). These
- approaches provide a framework to consider the potential impact of climate change on the
- 62 growth and distribution of Antarctic mosses, and the future of temperate, boreal and Arctic
- 63 peatlands. Finally, we explore the need for additional experimental validation of these
- proxies, and consider developments which could facilitate future monitoring of moss banks
- under a changing climate.

Poles apart: contrasting diversity and climatic impacts upon high latitude vegetation

- 67 Both polar-regions experience low mean annual temperatures, seasonal extremes in day
- length and often have low free water availability. At high northern latitudes recent
- 69 temperatures are unprecedented over at least the last 600 years (Tingley & Huybers, 2013),

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and the maritime Antarctic Peninsula (AP), the most vegetated area of Antarctica, has been one of the most rapidly warming regions on Earth (Turner et al., 2009). As winter temperatures increase more rapidly than summer temperatures, polar seasonality has decreased (Franzke, 2012, Xu et al., 2013), and permafrost distribution has altered (Bockheim et al., 2013, Guglielmin et al., 2014, Quinton et al., 2011, Turetsky et al., 2007) which is expected to have significant effects on the global carbon budget (Koven et al., 2011, Schuur et al., 2009). Earlier annual melting of both northern (Surdu et al., 2014) and southern (Quayle et al., 2002) polar lakes indicates a potential lengthening of the biological growing season. Despite the similar physical responses of the poles to warming, the contrasting relative geographical isolation of Arctic and Antarctic terrestrial ecosystems have generated substantial differences in past, present and likely future floral diversity. Antarctic vegetation is limited to Southern Ocean islands, areas of the AP, rare ice-free locations around the continent's perimeter and occasional interior nunataks. Seasonally snowand ice-free ground comprises only 0.34% of the total continental area (Convey et al., 2009). The isolation of Antarctica, separated from land masses by the Southern Ocean, Antarctic Circumpolar Current and Southern Westerly Wind belt (Barnes et al., 2006) is in contrast to the Arctic, where the continuity of Eurasia and North America has facilitated the development of more complex ecosystems. Along with substantial vascular plant diversity (1800 species), 600 moss species are found in the Arctic, which represents over 4% of global diversity (Callaghan et al., 2005). Bryophytes are poikilohydric plants with little capacity to buffer changes in water availability, so periods of metabolic activity are tightly coupled to microclimatic conditions (Longton, 1988, Royles et al., 2013b, Schlensog et al., 2013, Wasley et al., 2006). Consequently, preserved bryophyte tissue has the potential to provide an archive of local conditions at the time of active photosynthesis (Clarke et al., 2012, Royles et al., 2012).

Bryophytes as polar palaeoclimate recorders

Given the scarcity of plant monitoring programmes in Antarctica (Wall *et al.*, 2011), permafrost-preserved moss peat deposits contain an unrivalled temporal biological record. These ombrotrophic peat banks, comprised of *Chorisodontium aciphyllum* (Brotherus) and/or *Polytrichum strictum* (Bridwell), can date from over 5000 years ago (Björck *et al.*, 1991b, Fenton, 1980) and multi-proxy analyses provides information about the past environment along with insights for the likely responses to a changing climate. As moss growth is entirely dependent on enzymatic processes, tempered by liquid water availability, peat cores provide unequivocal evidence of periods during which environmental conditions were suitable for photosynthesis. Globally, mining the information stored in stratigraphic peat deposits has allowed significant advances in the understanding of past environments via reconstruction of vegetation history from pollen and higher plant remnants in combination with absolute accumulation rates derived from age-depth profiles (e.g. Chambers *et al.*, 2011, Charman *et al.*, 2013, Piotrowska *et al.*, 2011).

2. Distribution, past change and possible future dynamics of maritime

Antarctic peat banks

The majority of high latitude Southern Hemisphere peatlands are in Patagonia, however they comprise only 1% of the global peatland area (Yu *et al.*, 2010). Patagonian peatlands experience a climate with low precipitation and mild temperatures, and this minimal seasonality facilitates a long, potentially continuous, growing season (Loisel & Yu, 2013, Smith & Clymo, 1984). Peat addition rates are high, as conditions favour long periods of assimilatory carbon gain, whilst decomposition is limited by low summer temperatures (Loisel & Yu, 2013). Significant peat accumulations are also present on several sub-

Antarctic islands (Van der Putten *et al.*, 2012), with this region again being characterised by limited seasonality in many environmental variables (Convey, 2013).

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Further south, Antarctic peat banks are characterised by the absence of vascular plant material, the dominance of C. aciphyllum, or P. strictum in drier areas, and slow accumulation rates of approximately 1 mm yr⁻¹ (Fenton & Smith, 1982, Smith & Corner, 1973). The most significant banks are located within the Antarctic Peninsula and Scotia Arc, between Signy Island (60°S, South Orkney Islands) (Fenton, 1980, Smith, 1972) and Elephant Island (61°S, South Shetland Islands) (Björck et al., 1991b) in the north, and Alexander Island (Convey et al., 2011, Royles et al., 2013a) (69°S) in the south (Figure 1). In comparison with the peat deposits of the sub-Antarctic islands (Van der Putten et al., 2012) and Patagonia (Loisel & Yu, 2013), which have higher species diversity, there are both advantages and disadvantages to the use of cores from Antarctic moss peat banks as palaeoclimate archives. Mono- or bi-specific cores in which species can be readily identified facilitate chemical analysis without the need for subsequent separation of species effects from climate effects. However, the low diversity precludes direct interpretation of change to species composition as an indicator of environmental conditions. With the presence of permafrost below c. 30 cm depth (Fenton, 1980, Guglielmin et al., 2012) and little microbial activity (Block, 1984, Convey, 2001) within the active layer, peat compression and decomposition are minimal. Consequently, organic matter is extraordinarily well preserved in the Antarctic banks, to the extent that 1500 year old moss shoots have recently been shown to regenerate spontaneously in the laboratory (Roads et al., 2014).

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The appropriate interpretation of palaeoclimate archives in the context of broader spatial and temporal changes requires accurate methods for dating material. The purity and cryo-

protection of moss shoots, the absence of roots and minimal translocation of substances makes moss peat ideal material for accurate radiocarbon dating (Björck *et al.*, 1991a). Dating of basal peat provides a minimum date at which conditions became suitable for net assimilation following the Last Glacial Maximum (LGM). The Antarctic moss banks vary in their age, accumulation rate and continuity, with the oldest known banks on Elephant Island (61°08'S 054°42'W) dating back over 5000 years (Björck *et al.*, 1991b). The new date for the base of the peat at Norsel Point, Anvers Island (64°45'S 064°05'W), 620 km SW of Elephant Island suggests accumulation for at least 1500 years, whilst at the southernmost site, a further 600 km SSW on Alexander Island (69°22'S 71°51'W) the basal date shows that net accumulation began within the last two centuries (Figure 1b).

The restricted distribution of moss peat banks demonstrates that, whilst permanently ice free ground is essential for their establishment, it is by no means sufficient. Fenton and Smith (1982) identified eight factors (i-viii, below) that determine the extent of an Antarctic moss bank. Thirty years on, we re-assess each of these factors in terms of establishment, maintenance and accumulation.

i) Length of time the ground is snow-free in summer

Moss requires snow and ice-free ground on which to establish, and, although limited photosynthesis can occur through some snow cover as long as the tissue has thawed (Longton, 1988), the majority of net assimilation will occur following the spring snow melt. The length of time that the ground is snow-free in summer has generally increased. On Signy Island, ice cover reduced by 45% between 1950 and 1990 (Smith, 1990), and freshwater lakes are annually ice-free for an additional two months (Quayle *et al.*, 2002), both of which are proxies for an increase in the length of the biologically active season (Block & Convey, 2001). This has been reflected in the growth rates of *C. aciphyllum* moss banks on Signy

Island which increased from a maximum of 2.4 mm y⁻¹ in 1976 (Fenton, 1980) to 3.9 mm y⁻¹ in 2009 despite windier and wetter summers reducing the instantaneous photosynthetic assimilation rate (Royles *et al.*, 2012).

ii) Length of time the ground has been free from permanent snow and ice cover

The AP and surrounding islands were ice covered during the LGM. Currently approximately 3.6% (4300 km²) of the land area of the AP, South Orkney and South Shetland Islands is ice-free (Peat et al., 2007), an essential requirement for the establishment of moss. The retreat of 87% of AP glacier termini and reduced permanent snow cover (Cook et al., 2005, Strauss et al., 2009) provides land on which new moss banks could establish, and may re-expose moss tissue that can spontaneously regenerate new growth (LaFarge et al., 2013; Roads et al., 2014). Rapid glacier retreat on Ellesmere Island, Canada revealed viable bryophyte assemblages that had been buried under ice for up to 400 years (La Farge et al., 2013). Laboratory experiments show that Antarctic C. aciphyllum has the capacity for regrowth from shoots at least 1500 years old (Roads et al., 2014). Although photosynthesis can occur through a limited snow covering, changes in snow cover patterns over time could explain periods of very slow / negligible peat accumulation that are evident in some age-depth models (e.g. Björck et al., 1991b, Royles et al., 2012), but the capacity for re-establishment of growth following subsequent alterations to snow distribution is also becoming clear.

iii) Stability of the substratum

Moss banks require stable rock or soil in order to establish and accumulate (Fenton & Smith, 1982) so volcanic soils, areas undergoing substantial frost heave and beaches are unsuitable. Changes in the extent and depth of permafrost will impact the stability of the substratum more rapidly than substantial geological changes. On Signy Island, the active layer thickness is projected to increase at 10 mm yr⁻¹ (Cannone *et al.*, 2006). In 1976 the depth of seasonal *C*.

aciphyllum bank thaw was 210±10 mm (Fenton, 1980), by 2009 the thaw in the same banks was 300±10 mm (Royles *et al.*, 2012). This 40% increase in seasonally unfrozen biomass was coincident with an increase in mean annual temperature (MAT) of 0.9°C (Quayle *et al.*, 2002); however, there can be substantial inter-annual variation in permafrost depth (Guglielmin *et al.*, 2014). Further south, in Victoria Land, continental Antarctica the active layer depth has increased at approx. 0.3 cm yr⁻¹ since 2000, which has altered soil stability and decreased water availability to mosses (Guglielmin *et al.*, 2014).

iv) Biotic disturbance

Growing slowly without strong attachment to the substratum, moss will not successfully establish on frequently disturbed ground. Established moss banks are vulnerable to erosive damage by both abiotic and biotic agents and are increasingly at risk as the depth of the active layer increases (see above). Fur seals (*Arctocephalus gazella*) cause physical damage to terrestrial ecosystems (Smith, 1988a), which is evident on Signy Island where 73% land is covered with bryophytes in areas without seal pressure compared with 1.7% of land in areas with high seal pressure (Favero-Longo *et al.*, 2011). Thus, whilst potentially increasing nutrient availability, a recent eight-fold increase in the fur seal population (1600 in 1978, 12600 in 2008) (Waluda *et al.*, 2010) has negative consequences for the establishment and maintenance of moss banks.

v) Harshness of the environment – i.e. exposure

Antarctica is windy: on the South Shetland Islands the mean wind speed is 26 km hr⁻¹, with gusts frequently over 100 km hr⁻¹ (Bañón *et al.*, 2013). Furthermore, summer and autumn wind speeds significantly increased (by 7-27% in summer) between 1980 and 2002 in the latitude band from 50 to 65°S (Korhonen *et al.*, 2010). With increased wind speed the rate of erosion is likely to increase (especially with the increased depth of the active layer), and the

rate of evaporation will increase from the surface of soils and mosses. Although the mass of a bank largely buffers the overall water content (Gimingham & Smith, 1971), if the surface water film evaporates, the growing apices may become increasingly water limited and desiccated.

vi) Nutrient status

Animal colonies provide important nutrient inputs to terrestrial ecosystems, both directly, and indirectly from wet and dry atmospheric deposition (Bokhorst *et al.*, 2007). There have been substantial changes in animal populations and distributions over the past 30 years in the AP region. Populations of chinstrap (*Pygoscelis antarctica*) and Adélie (*Pygoscelis adeliae*) penguins have decreased, whilst gentoo (*Pygoscelis papua*) penguin and fur seal populations have increased (Lynch *et al.*, 2012, Waluda *et al.*, 2010) which may alter local nutrient distribution. In addition to changes in nutrient availability, there is potentially increased competition for nutrient uptake as the vascular plant *Deschampsia antarctica* is able to take up amino acids directly through its roots and thus acquire nitrogen over 160 times more quickly than Antarctic mosses (Hill *et al.*, 2011) and this efficient uptake and usage may be a mechanism by which the Antarctic vascular plants out-compete bryophytes in the long term.

vii) Water supply

The form and timing of precipitation are critical to the establishment, growth and development of plants. Indeed, water supply is the most important physical driver of Antarctic terrestrial communities and limits floral distribution across the continent (Convey *et al.*, 2014). Across the maritime Antarctic precipitation is frequent, with an increasing proportion falling as rain rather than snow; however, water availability can vary very locally which has a significant impact on the metabolic activity of plants (See sections 4&5) (Schlensog *et al.*, 2013, Wasley *et al.*, 2012). The capacity for capillarity may limit the depth

across Antarctica.

to which poikilohydric moss tissue can accumulate as exposed growing tips lose water through evaporation (Melick & Seppelt, 1997).

viii) Slope

With the depth of the active layer deepening, the wind strengthening and increased summer rain, moss banks are more vulnerable to erosion, slumping and collapse. After loss of the top section, the newly-exposed surface moss may resume growth, or be invaded by epilithic lichens, other mosses, or provide an environment for vascular plant growth (Fabiszewski & Wojtun, 1997).

In conclusion, the preceding analyses suggest that, whilst some edaphic factors may limit colonisation and moss bank formation/degradation, other factors associated with climate change, such as an extended growing season and altered precipitation patterns may help to enhance carbon accumulation overall. We now consider the implications for moss bank growth, and specifically compare the drivers for the contrasting extent of peat accumulation

3. Moss accumulation rates across Antarctica

Fewer than 30 species of the known moss flora of Antarctica have been identified south of 67°S, and none are currently thought to be endemic to the very cold and dry continental region (Ochyra *et al.*, 2008, Peat *et al.*, 2007). Whilst across the AP diversity decreases at higher latitudes (Peat *et al.*, 2007), the continental distribution of plants suggests that it is local microclimate conditions, particularly water availability, that are critical in determining floral diversity (Cannone *et al.*, 2013, Colesie *et al.*, 2014, Convey *et al.*, 2014, Green *et al.*, 2011). Vegetation surveys across Antarctica have been very limited spatially and temporally,

with basic species presence and proportion cover records mainly focussed around research stations (Chown & Convey, 2007). It is rare for the depth of moss growth to be recorded; *in situ* experimental systems, to measure rates of moss growth, are difficult to establish and repeat visits to assess changes are rare. Consequently, there is limited baseline information available on Antarctic bryophytes from which any past or future effects of climate change can be assessed.

The best studied terrestrial vegetation in continental Antarctica is from the Windmill Islands region in East Antarctica, where three moss species (*Schistidium antarctici* (Cardot), *Bryum pseudotriquetrum* (Hedwig) and *Ceratodon purpureus* (Hedwig)) are found (Melick & Seppelt, 1997, Robinson *et al.*, 2000, Wasley *et al.*, 2012). Areas of moribund moss suggest that vegetation has changed, but it is difficult to determine the timing of moss establishment due to fungal contamination compromising radiocarbon dating. Direct measurement of moss growth using tags is problematic due to the low growth rates (<7 mm in four growing seasons) and loss of tags (Melick & Seppelt, 1997).

The uneven species' distribution across Antarctica complicates the comparison between moss growth in the eastern and western regions, but the responses do provide contrasting markers for different life forms and climatic extremes. In the absence of detailed laboratory comparisons of physiology and growth under controlled environment conditions, we feel this broad comparison is informative. Whilst the moss peat accumulations in the maritime Antarctic region reach a maximum depth of 3 m on Elephant Island (Björck *et al.*, 1991b), the continental Antarctic moss associations rarely reach 10 cm (Melick & Seppelt, 1997). Combining published age-depth profiles from moss accumulations from the Windmill Islands (Clarke *et al.*, 2012), with those from Lazarev Bay (Convey *et al.*, 2011, Royles *et al.*,

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2013a) and Signy Island (Royles *et al.*, 2012) since 1960 it is apparent that the rate of moss organic matter accumulation in the maritime Antarctic is at least three times higher than that in continental Antarctica (Figure 3).

Within the continental Antarctic mosses, the accumulation of C. purpureus in the Windmill Islands, which prefers drier habitats (Robinson et al., 2000), is slower than that of the more mesophytic B. pseudotriquetrum and Bryoerythrophyllum recurvirostre (Hedwig) from the Vestfold Hills (Clarke et al., 2012). Compression and decomposition are not taken into account, but as the measurements are made on individual shoots up to only 50 mm long, the effects of both processes are minimal. In the maritime Antarctic region, the similarity in accumulation rate between the Lazarev Bay and Signy Island banks since 1960 is interesting, as they lie at opposite extremes of the distribution range, approximately 1500 km apart, and are comprised of P. strictum and C. aciphyllum respectively. At both locations the top 100 mm of peat, within which zone compression will be minimal, has accumulated since 1988, at approximately 4 mm vr⁻¹. Using bulk density measurements, the peat accumulation rate at Signy Island can be compared with that previously published at Lazarev Bay (Figure 4; (Royles et al., 2013a)). Whilst accumulation reached a peak at Lazarev Bay in the 1970s (briefly over 0.1 g DM cm⁻² yr⁻¹, when smoothed over three successive time-points), the rate at Signy Island reached a peak in the 1990s, at 0.06 g DM cm⁻² yr⁻¹. Both start to increase from around 1950 and the most recent measurements are similar at around 0.04 g DM cm⁻² yr⁻¹. Similarly in continental Antarctica, where moss growth rate was inversely proportional to summer wind-speed, and proportional to the number of days above 0°C and to summer temperature, the Windmill Island mosses had maximum growth rates in the 1950s-1980s that subsequently fell in the 90s and 00s. However, at the Vesfold Hills the most recent moss growth rates have been highest (Clarke et al., 2012). The length of the growing season is a critical determinant of moss growth rate in both continental (Clarke et al., 2012) and AP

locations (Royles *et al.*, 2012). The length of past growing seasons cannot be determined directly from moss-cores, but estimates have been made by combining accumulation rates derived from radiocarbon dating with stable isotope proxies for assimilation rate (Royles *et al.*, 2012).

4. Isotope proxies preserved in Antarctic moss over space and time

Stable isotopes provide natural markers for the environmental control of metabolic reactions (Dawson et al., 2002, Farquhar et al., 1989). Analyses of ¹³C or ¹⁸O, relative to the more common isotopomers (¹²C or ¹⁶O), when preserved in tree rings and peat cores, are widely used as environmental tracers that provide information about past growth conditions, with work published from Canada (e.g. Daley et al., 2011, Kaislahti Tillman et al., 2010) and northern Europe (e.g. Daley et al., 2010, Loader et al., 2008, Ménot-Combes et al., 2002, Ménot & Burns, 2001). More recently the ¹³C, ¹⁸O and ²H composition of cellulose, organic matter and source water have been used to quantify responses of Antarctic moss to climate change (Clarke et al., 2012, Royles et al., 2013a, Royles et al., 2012, Royles et al., 2013c). Mass-dependent "kinetic" fractionation of isotopes occurs either between the reactants and products of a unidirectional reaction or during diffusion, whilst "equilibrium" fractionation is the partial separation of isotopes between substances or phases in chemical equilibrium. Heavy isotopic species (e.g. ¹³CO₂ or H₂ ¹⁸O) tend to react more slowly and usually become enriched in a denser equilibrium phase, or depleted during a kinetic transformation, and thus provide important biomarkers (Farquhar et al., 1989). The ratio (R) of heavy to light isotopic species (e.g. $R_{SAMPLE} = {}^{13}CO_2/{}^{12}CO_2$) is usually measured via high-precision mass spectrometry, compared to a known standard ($R_{STANDARD}$) and presented as a differential (δ) notation, such as a δ^{13} C value:

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Source independent photosynthetic carbon isotope discrimination ($\Delta^{13}C$) can then be derived, if $\delta^{13}C$ values are known for both the plant ($\delta^{13}C_p$) material and the source CO_2 , usually atmospheric CO_2 ($\delta^{13}C_a$):

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$$\Delta^{13}C = \delta^{13}C_a - \delta^{13}C_p / 1 + \delta^{13}C_p \text{ (Farquhar et al., 1989)}.$$

 Δ^{13} C is dominated by the biochemical fractionation of the carboxylase enzyme RuBisCO, having a value of around 29% in C₃ plants (O'Leary, 1988). This maximal fractionation is proportionally inhibited by resistance to CO₂ diffusion, leaving assimilates (and subsequently plant tissue) ¹³C-depleted compared to atmospheric, source CO₂ inputs. In astomatous bryophytes, the external water layer is a critical determinant of diffusion resistance and, consequently, the extent of discrimination against ¹³CO₂. Real-time measurements on liverworts, Sphagnum moss and Syntrichia ruralis show that a reduction in the external water layer is associated with an increase in instantaneous discrimination against ¹³CO₂ and in assimilation rate (Meyer et al., 2008, Rice & Giles, 1996, Royles et al., 2013b, Williams & Flanagan, 1996). As a proportion of the assimilated carbon is used to synthesise the structural carbohydrate cellulose, a major degradation-resistant component of bryophyte organic matter, the carbon isotope ratio of moss cellulose ($\delta^{13}C_C$) is a good proxy of the assimilation conditions during photosynthesis (Royles et al., 2012). Antarctic moss tissue δ^{13} C values are dependent on wind speed, temperature (both of which influence water availability) and ozone depth (which is linked to the phase of the Southern Annular Mode, and, consequently, windspeed) (Clarke et al., 2012, Robinson & Erickson III, in press). Measurements of the ¹³C composition of Antarctic moss have been carried out on different species (including P. strictum, C. aciphyllum and C. purpureus), with different growth habits (large banks, hummocks and small cushions), with concurrent measurements back to the 1960s (Clarke et al., 2012, Royles et al., 2013a, Royles et al., 2012). Whilst δ^{13} C was

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measured in whole organic matter for C. purpureus, most measurements on C. aciphyllum and P. strictum were made on cellulose; however, comparative measurements of organic matter and cellulose had a highly significant linear relationship (Royles, 2012) so $\delta^{13}C_{OM}$ measurements could be derived. Over the 60 y period under consideration in these environments in which the rate of degradation is low, the compound specific diagenetic effects that are an important factor to consider over long time periods should not be substantial. There is a significant depletion in the composition of all the moss tissue over time due to the assimilation of atmospheric CO₂ which is globally becoming more depleted following the combustion of fossil fuels (Figure 5a) (Friedli et al., 1986). Source-independent Δ^{13} C was calculated for each measurement (Farquhar et al., 1989), dependent upon the isotopic composition of atmospheric CO₂ (Rubino et al., 2013) at the time of synthesis (as derived from ¹⁴C measurements and estimated from age-depth model; (Figure 5b)). Any contribution to assimilated carbon from sources of respiratory CO₂, such as those associated with microbes or moss decay, were not included in the calculation of Δ^{13} C. Overall rates of respiration and microbial metabolism are low (Royles et al., 2013a) and the exposed banks are generally well-coupled to the atmosphere in the prevailing windy conditions. Were any respiratory CO₂, with a source isotope composition close to that of bulk material, to be refixed, it would tend to slightly increase the Δ^{13} C values to a similar extent at all water contents. This response is not consistent with the declining Δ^{13} C seen in Signy Island moss banks (Royles et al., 2012), associated with warmer and wetter growth (and presumably higher respiration rates) over the past decade. Despite the species variation and geographic separation of up to 5000 km between sites, the measured Δ^{13} C values are very similar, covering a 4% range from 17 to 21%. The Lazarev Bay moss had the highest source independent discrimination with the exception of 1970 when one C. purpureus value was higher. The lowest discrimination values of 17‰ were

measured on Signy Island, with the three *C. purpureus* and three *C. aciphyllum* cores largely overlapping in values between 17% and 20%. When the Signy Island and Lazarev Bay $\Delta^{13}C_C$ values are considered back to 1850, the Lazarev Bay values show higher discrimination, which could be due to the species effect i.e. the Signy core is comprised of *C. aciphyllum*, whilst the Lazarev core is *P. strictum*, two species which have different anatomies and external water layers, along with potentially different metabolic responses to environmental conditions. In all the cores, the source independent discrimination was higher in 2000 than at the start of the record, with one of the Signy Island cores showing a 3% increase, with a 2% range at Lazarev and 1.5-2% elsewhere on Signy. On Signy Island the majority of the increase in $\Delta^{13}C$ occurred prior to 1960, compared with the substantial increase being measured from 1960 onwards at Lazarev Bay.

Higher discrimination values generally suggest more optimal conditions of hydration, when photosynthetic carbon is fixed during periods of minimal diffusion limitation (i.e. neither too wet nor too dry). At both Signy Island and Lazarev Bay there has been a tendency for a reduction in the measured values most recently, with highest discrimination in the mid 1990s. The increase in *C. purpureus* Δ^{13} C reflects drying conditions over time (Clarke *et al.*, 2012), an effect also measured spatially across the bryophyte-dominated community of the Windmill Islands. Here, moss gametophyte organic matter δ^{13} C values were significantly less negative than in the dry, lichen-dominated community, which was suggested to indicate more frequent submergence and consequently more significant diffusion limitation (Wasley *et al.*, 2012).

Without roots and with limited conduction systems, mosses are dependent on precipitation, dewfall or ground water for hydration and the isotopic composition of this source water ($\delta^{18}O_{SW}$) is an important determinant of cellulose isotope composition ($\delta^{18}O_{C}$). Globally, the composition of source water depends on various factors, the most relevant for

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Antarctica being the form and amount of precipitation. Following initial evaporation from seawater (generating water vapour isotopically depleted in ¹⁸O) successive precipitation events (which favour the remaining heavy isotopes) become progressively depleted due to a Rayleigh distillation (Gat, 2000). Snow (and subsequent snow melt water) is more isotopically depleted than rain, and precipitation is also more depleted at high latitudes. However, even if the water source is known, $\delta^{18}O_{SW}$ is often unequal to the isotopic composition of leaf water ($\delta^{18}O_L$) at the site of cellulose synthesis due to preferential evaporation of lighter water isotopologues (H₂¹⁶O). Additionally, under high humidity conditions the rate of diffusive vapour exchange may be sufficient to imprint $\delta^{18}O_{I}$ with the atmospheric vapour isotope signal rather in place of $\delta^{18}O_{SW}$ (Helliker & Griffiths, 2007). Furthermore, there is an approximately 27‰ biochemical fractionation during the synthesis of organic matter, either following direct transfer of the $\delta^{18}O_L$ signal to sugars formed during photosynthesis and thence to organic material, or after re-equilibration of carbonyl groups that exchange with oxygen atoms in the surrounding water during the metabolic pathway of incorporation into storage polysaccharides (Barbour, 2007, Da Silveira et al., 1989, DeNiro & Epstein, 1979, Sternberg et al., 2006). The biochemical fractionation factor may be temperature dependent, with particular sensitivity at low temperatures (5-15°C) (Sternberg & Ellsworth, 2011) which would be highly relevant to Antarctic plants. Thus, there are multiple factors that influence the measured oxygen isotope composition of bryophyte material at any particular time or place, with further complexity introduced from morphological and physiological differences between species. Factors which affect desiccation tolerance, the timing of cellulose synthesis relative to the extent of saturation, and microclimate and micro-topographical effects on relative humidity around the leaf could all affect the extent of evaporative enrichment. On Signy Island, there was a consistent offset in the oxygen isotope composition of cellulose between material obtained from low lying

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hummocks, and more extensive moss banks (Royles *et al.*, 2013c). This was attributed to the dominant water source in the former being more depleted snow melt water as compared to summer precipitation.

The extent of capillary water is an important determinant of both the carbon and oxygen isotope compositions, but $\delta^{18}O_C$ is less dependent upon the photosynthetic rate than δ¹³C_C where, in combination with the capillary water, photosynthetic rate largely determines the diffusive supply of CO₂ from the atmosphere to the chloroplast (Rice & Giles, 1996, Royles et al., 2013b, Williams & Flanagan, 1996). The oxygen signal is largely dependent upon the difference in relative humidity between the moss tissue and atmosphere. The two isotope signals also represent subtly different time points. Whilst the $\delta^{13}C_C$ signal represents atmospheric CO₂ the external water layer thickness and internal conductance during carbon assimilation, $\delta^{18}O_C$ reflects the time of cellulose synthesis, when most oxygen atoms will exchange and re-equilibrate with contemporary tissue water. Experimental evidence from the desiccation tolerant moss S. ruralis suggests that this temporal separation may be important, with $\delta^{13}C_C$ reflecting the time of maximum assimilation, just prior to metabolic limitation due to desiccation, whilst cellulose synthesis occurs following re-saturation with source water after rain / dewfall (Royles et al., 2013b) as turgor pressure is required for cell wall expansion and growth (S. A. Robinson pers. comm.). For moss banks and hummocks that experience lower daily fluctuations in water content than Syntrichia ruralis, the separation between periods of maximum assimilation and maximum cellulose synthesis are likely to be less distinct. Further experimentation under controlled environmental conditions is required to define these responses more generally.

With multiple factors influencing the measured $\delta^{13}C$ and $\delta^{18}O$ values, here we consider the values in tandem, in an attempt to identify biologically relevant clusters (Figure 7). Measurements of $\delta^{13}C_C$ and $\delta^{18}O_C$ have been made on *C. aciphyllum* and *P. strictum* from

extensive moss banks and hummocks on Signy Island, and from a hummock at Lazarev Bay on Alexander Island (Figure 7) (Royles *et al.*, 2013a, Royles *et al.*, 2012, Royles *et al.*, 2013c). $\Delta^{13}C_C$ values were used to remove the impact of the systematic isotopic depletion in atmospheric CO_2 over the industrialised period; however, a source effect was not removed from $\delta^{18}O_C$, as there was no independent record of source water composition. When both $\delta^{13}C$ and $\delta^{18}O$ measurements are considered the data points separate into three clusters:

- 1) C. aciphyllum from extensive Signy Island banks (Δ^{13} C= 14-17‰, δ^{18} O = 26-31‰)
- 2) *C. aciphyllum* from small Signy Island hummocks (Δ^{13} C = 15-17‰, δ^{18} O= 22-27‰)
- 468 3) *P. strictum* from Signy Island and Lazarev Bay (Δ^{13} C 17-19.5‰, δ^{18} O= 23-27‰)
- This can also be characterised as:

- 470 1) *C. aciphyllum*: Δ^{13} C< 17‰;
- *P. strictum*: Δ^{13} C >17‰
- 472 2) Hummock (e.g. Fig. 2f): δ^{18} O < 26%;
- 473 Bank (e.g. Fig. 2a): $\delta^{18}O > 27\%$

Several factors underlie these groupings. Species has a dominant effect on Δ^{13} C. Lamellae on the leaves of *P. strictum* facilitate relatively faster diffusion of CO_2 into the leaf as compared to *C. aciphyllum* under the same moisture conditions, hence the higher discrimination. *Chorisodontium aciphyllum* is dominant in wetter areas whilst *P. strictum* is associated with drier conditions, (Fenton & Smith, 1982), as, being endohydric, *P. strictum* has some capacity for internal water transport which enables tissues to maintain hydration for longer in dry conditions. It is surprising not to see a latitude effect in *P. strictum* δ^{18} O values, as being 9° further south than Signy Island, the isotopic composition of precipitation at Lazarev Bay would be expected to be more depleted in 18 O values. In contrast, whilst the carbon isotope composition of all the *C. aciphyllum* samples falls within the 14-17% range, the oxygen isotope composition of cellulose from bank-forming *C. aciphyllum* is significantly higher

than that from hummock forming *C. aciphyllum*. This offset can be attributed to the inputs of isotopically lighter summer precipitation, as compared to more depleted melt water, and also influenced by the extent of evaporative enrichment (Royles *et al.*, 2013c).

Variation in δ^{13} C and δ^{18} O measurements of 3‰ and 4‰ respectively remains within the clusters, and indeed individual cores, representing variation measured over time that reflects local environmental changes. Photosynthetic conditions can vary locally, as cryo-perturbation makes the moss surface uneven at the centimetre scale, with the tops of ridges being driest and windiest (Lovelock & Robinson, 2002, Lucieer *et al.*, 2013). This can affect both Δ^{13} C and δ^{18} O measurements, with higher Δ^{13} C values associated with drier conditions and higher δ^{18} O with a greater degree of evaporative enrichment. On the millennial scale that the *C. aciphyllum* core data represents, differential rates of growth and the disturbing effects of winter snow cover may alter the relative positioning of the growing moss within the local micro-topography. Thus, the stable isotopic composition of mosses over time is dependent upon species and can provide information about the growth form and microclimate conditions at the leaf surface, as well as reflecting to some extent the narrow range of environmental conditions which support growth.

Not all isotopic variation can currently be attributed to a particular environmental factor, hence more laboratory work is required to elucidate under controlled conditions some of the drivers of the variation (such as the influence of temperature on isotope fractionation factors during cellulose biosynthesis (Sternberg & Ellsworth, 2011)) and also additional proxies could be analysed to provide more information about the past. For example, the analysis of $\delta^2 H_C$, alongside ¹⁸O isotopomers, could be used to compare the $\delta^2 H$ -temperature relationship along modern elevation and latitude gradients. In this way, *Sphagnum* $\delta^2 H_C$ from a Patagonian peat bog was used to reconstruct the past meteoric water composition and

revealed abrupt temperature changes during the late Pleistocene and early and middle Holocene (Pendall *et al.*, 2001). By combining δ^2 H and δ^{18} O values from organic compounds it is possible to estimate the meteoric water composition at the time of synthesis, and potentially estimate temperature and relative humidity conditions that could result in the measured isotope values (Edwards, 1993, Edwards *et al.*, 2000).

5. Contemporary moss physiology: integrating isotope signals, plant water relations, photosynthetic gas exchange and chlorophyll fluorescence

Identifying conditions under which mosses are photosynthetically active, and especially periods of net assimilation, is crucial to interpreting preserved proxy signals and predicting the responses of moss to climate changes. In addition to water availability, which is the most critical factor in determining Antarctic moss distribution (Cannone *et al.*, 2013, Colesie *et al.*, 2014, Convey *et al.*, 2014), diffusion limitation, life-form, length of growing season (Royles et al 2012) and light availability (Loisel et al. 2012; Charman et al. 2013) have also been implicated as major contributors to the dynamics of moss growth at the ecosystem level.

Photosynthetic light use can be estimated using chlorophyll fluorescence as a non-destructive measure of photosystem activity, allowing quantum yield (Ψ_{PSII}), electron transport rate (ETR) and energy dissipation (non-photochemical quenching, NPQ) to be determined for Antarctic mosses under laboratory (Stanton *et al.*, 2014) and field conditions (Robinson *et al.*, 2000, Schlensog *et al.*, 2013) to show, for example, the relative sensitivity of mosses to desiccation (Robinson *et al.*, 2000). The mesic mosses *Bryum argentum* and *B. pseudotriquetrum* had sufficient water for continuous metabolism with seasonally integrated

ETR (representing assimilation) largely dependent upon light and temperature (Schlensog *et al.*, 2013). In contrast, the most xeric mosses (*C. purpureus, Stereocaulon alpinum* and *Andreaea gainii*) were only active for 40-50% of time. With 60% activation *Polytrichum alpinum* was the most active xeric moss as being endohydric it has as some capacity for water transport and a cuticle to reduce the rate of water loss (Schlensog *et al.*, 2013).

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By following photosynthetic gas exchange and isotope discrimination during a drying curve stable isotope physiology and hydrology can be investigated (Rice & Giles, 1996, Royles et al., 2013b, Williams & Flanagan, 1996). Chlorophyll fluorescence and derived characteristics (ETR, NPQ) can also be used to track photosynthetic activity and the impact of dehydration (Figure 8, J Royles, H Jenkins and H Griffiths, unpublished data). During desiccation, S. ruralis ETR increased as photosynthesis became less diffusion limited, reaching a maximum at a relative water content (RWC) of 150%; below 100% RWC, ETR declined rapidly as moss photosynthesis was compromised by desiccation (Figure 8a). This profile tracked the real-time carbon isotope discrimination signal (Royles et al., 2013b) and was consistent with ETR as an additional proxy for carbon uptake. Successive light response curves throughout the drying curve (Figure 8b) also show the overall responsiveness of fluorescence, and potential for the development of ETR as a remotely-sensed proxy for photosynthetic activity as a function of moss hydration status. Additional measurements on this system showed that the isotope composition of soluble carbohydrates can also be used to integrate daily carbon gain. The 13 C signal (Δ^{13} C=20±0.5‰ (n=8)) was indicative of optimal gas exchange, whilst the ¹⁸O signal was more sensitive to evaporative enrichment during desiccation (increasing from 21.7±0.8‰ to 25.1±0.6‰ (n=4 each)) as it was probably reequilibrating with tissue water during growth in the light (J Royles, A Sturgess and H Griffiths, unpublished data). The rapid physiological responses of S. ruralis to RWC reflect the tight coupling between the moss and current environmental conditions. We would expect bank forming mosses, with less variable tissue water contents, to show somewhat buffered, or muted, coupling between changing environmental conditions, fluorescence and isotope signals. However, the prospects for resolving the variations within and between ¹³C and ¹⁸O signals in Antarctic mosses seem promising (Figure 7), as well as the use of chlorophyll fluorescence to investigate moss bank responses at ecosystem and regional scales.

6. Responses of polar mosses to climate change

There are many urgent ecological and environmental problems in the Arctic related to mosses in a changing climate, but the geographical range of species and life-forms are difficult to track individually. The goal was to translate what we have learned from more simple systems in Antarctica, and apply to Arctic habitats.

i) Direct and indirect responses to temperature

For Antarctic bryophytes, chlorophyll fluorescence measurements were used to infer that the extent of the hydrated (active) period was a more important influence on photosynthetic performance than temperature, with reliance on melt water or more temporary activation from precipitation being a key factor (Schlensog *et al.*, 2013). Increases in temperature will alter water availability to mosses through changes in the timing and extent of melt streams and increases in active layer depth (Guglielmin *et al.*, 2014). The speed with which plants reactivate photosynthesis following both prolonged winter freezing, rapid freeze-thaw cycles and periods of summer desiccation, is relevant to the seasonal assimilates preserved in organic matter. Mesic Antarctic mosses take 24 h to fully recover net photosynthesis following desiccation due to resaturation respiration (Schlensog *et al.*, 2004), a transient increase in respiration widely seen in anhydrobiotic organisms following rewetting, possibly due to metabolic costs of repair, or mitochondrial inefficiency (Nijp *et al.*, 2014).

Experimental warming was tentatively associated with a decrease in moss cover in the Arctic tundra, but the response varied substantially over space and time (Elmendorf *et al.*, 2012) and with limited samples it is difficult to account for the influence of ambient conditions and species-specific dynamics (Lang *et al.*, 2012, Street *et al.*, 2013) especially as moss distribution may be more sensitive to moisture availability than warming (Lang *et al.*, 2009). Other indirect effects likely to be associated with a warming climate for both polar regions include the extended growing season identified for maritime (Royles *et al.*, 2012) and continental Antarctic mosses (Clarke *et al.*, 2012), as well as altered precipitation patterns affecting wetting and light limitation (cloudiness). In conclusion, it is predicted that significant changes will occur to both polar plant growth and vegetation distribution in response to a changing climate (Chapin III, 2003, Chapin III *et al.*, 2005, Chapin III *et al.*, 2010), but a more detailed understating of how contrasting plant functional groups respond to these challenges (Turetsky *et al.*, 2012) is needed to test predictions of polar ecosystem resilience to climate change.

ii) Direct and indirect responses to light

Bryophyte photosynthetic activity is responsive to light intensity in moist habitats, but less so in more xeric habitats, where water limitation is the major determinant (Schlensog *et al.*, 2013). Changes to cloud cover will alter light availability for photosynthesis, at Vernadsky station (Fig 1a) cloud cover increased significantly between 1960 and 2005 (Kirchgaßner, 2010). The growth of *Sphagnum* was directly related to light intensity in wetter sites in Alaska, where increasing water table and acidification were associated with a decline in black spruce (Fenton & Bergeron, 2006). In two recent extensive meta-analyses of peat accumulation, light availability was related to *Sphagnum* growth, particularly at high latitudes (Charman *et al.*, 2013, Loisel *et al.*, 2012). Indeed, total carbon accumulation in northern

peatlands over the past 1000 y was linearly related to photosynthetically active radiation projected over the growing season (defined as the Growth Degree Days, for which air temperature is greater than 0°C, GDD0), and with a stronger correlation than with GDD0 alone (Charman *et al.*, 2013).

iii) Responses to water availability

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Mosses from xeric habitats will go through multiple desiccation cycles during a growing season and be tolerant of desiccation (c.f. "low-inertia" species (Proctor & Tuba, 2002)), whilst those in wet flushes are likely to remain damp and make more use of dehydration repair processes ("high-inertia species" (Proctor & Tuba, 2002)) (Proctor, 2000, Schlensog et al., 2004). Furthermore, carbon use efficiency differs between species that maintain a relatively constant RWC over time (e.g. Sphagnum spp.) and those which are metabolically active throughout drying cycles from saturation to desiccation (Street et al., 2013). The biphasic response of carbon accumulation to Precipitation/Evaporation (P/E) over the past 1000 y, identified by both (Charman et al., 2013, Loisel et al., 2012), is consistent with daily changes seen in individual mosses (Royles et al., 2013b) (Figure 8a). Thus, there is an optimal carbon accumulation under relatively low P/E values, with a rapid decline in growth under driest conditions (high evaporative demand), and a more gradual reduction under increasingly wet conditions (higher P/E conditions (see Figure 5c in (Charman et al., 2013)). Therefore, there is still a need to characterise the relationships between light availability and the impact of warmer and wetter, or warmer and drier, conditions for bryophyte photosynthetic activity over the course of a longer growing season. An important additional consideration for the water relations of mosses is plant life-form and functional type (filamentous, hummock/hollow (Turetsky et al., 2012)). There may be separation between the photosynthetic tissues, which are usually restricted to the top 5 mm of

shoots and thus exposed to wind and hence dry out quickly, and water storage in spongy tissue below (Stanton *et al.*, 2014). Rates at which individual shoots lose water to the atmosphere can be very different to that of whole clumps and capacity for internal water redistribution is a critical component of physiology. Similarly, the moss temperature can be much warmer than the air temperature (Longton, 1982, Smith, 1988b), and it is important to separate the effects of vapour pressure differences between tissue and atmosphere (VPD) and air temperature: VPD, which determines the rate of drying, varies non-linearly with temperature (Stanton *et al.*, 2014). Indeed, polar mosses can have high rates of photosynthesis over a range of temperatures (Davey & Rothery, 1997, Pannewitz *et al.*, 2005). The strong influence of VPD on moss photosynthesis in both *C. purpureus* and *S. antarctici* means that temperature, precipitation and VPD all need to be modelled accurately before conclusions can be drawn about the effects of climate change (Stanton *et al.*, 2014).

7. Scaling physiological proxies to monitor and model climate change

for the future

Mosses from across Antarctica are responding to climate change with changes in growth rates and stable isotope proxies as discussed above (Clarke *et al.*, 2012, as discussed above; Royles *et al.*, 2013a, Wasley *et al.*, 2012). However, whilst data are logistically difficult to obtain in such remote regions, interpretation might be relatively simple compared to complex seasonal vegetation-permafrost dynamics in Arctic ecosystems (Chapin III *et al.*, 2010, Sistla *et al.*, 2013, Tingley & Huybers, 2013, Turetsky *et al.*, 2012).

Firstly, one must distinguish between boreal forest (coniferous dominated, mainly north of 45°N) and tundra (8.2 M km², mostly north of 65°N) (Xu *et al.*, 2013), and the contrasting responses of bryophytes and their associated carbon storage capacity (Charman *et al.*, 2013, Sistla *et al.*, 2013, Yu *et al.*, 2010). Within this enormous range of habitats, three generalised

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responses might be anticipated: (i) a northerly shift in vegetation boundaries associated with rapid warming and restricted extent of permafrost (Baltzer et al., 2014, Turetsky et al., 2012, Turetsky et al., 2010, Xu et al., 2013); (ii) increasing frequency and intensity of drought, and associated fires in more continental areas, likely to reduce Sphagnum dominance and increase conifers capable of regenerating after fire (Chapin III, 2003, Chapin III et al., 2010, Turetsky et al., 2012, Turetsky et al., 2010); (iii) increased precipitation in more maritime regions of Alaska and western Canada, as well as in Fennoscandinavia and Western Siberia, leading to enhanced Sphagnum growth, elevated water tables and forest decline associated with paludification (Crawford, 2008, Crawford et al., 2003, Fenton & Bergeron, 2006, Turetsky et al., 2012). Key questions for the future relate to the development of methods and approaches needed to monitor these processes remotely, to allow modelling and predictions of climate change impacts to be validated (Chapin III et al., 2010, Crawford et al., 2003). In the Antarctic, we have recently seen the use of Landsat satellite images to project current vegetation distribution via Normalised Distribution Vegetation Index (NDVI: (Fretwell et al., 2011)) and spatial variations in moss bank surface temperature and spectral reflectance, estimated from an Unmanned Aerial Vehicle (UAV), mounted with sensors and cameras (Turner et al., 2014). Chlorophyll fluorescence has been used to ground-truth the responses of cryptogamic communities to water availability (Schlensog et al., 2013), and there is now the possibility of using the Laser Induced Fluorescence Technology (LIFT) as a remote-sensing system, across a range of spatial scales (Kolber et al., 2005, Pieruschka et al., 2012). Finally, the interpretation of solar-induced fluorescence from UAVs (Damm et al., 2014) or satellite systems (Zhang et al., 2014) has recently been provided with a quantitative framework to interpret components such as quantum yield, ETR and NPQ (Guanter et al., 2014). The opportunity for their use in distinguishing moss photosynthetic activity and carbon

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sequestration as a function of water availability, across a changing forest mosaic, and at regional scales for polar regions, is now a realistic prospect. There will be exciting opportunities for ground-truthing these remote proxies under challenging Arctic and Antarctic conditions.

8. Conclusion

In conclusion, recent key analytical and modelling advances in the use of stable isotopes of carbon have helped to provide a quantitative basis to moss photosynthesis, growth and seasonality across Antarctica, and with implications for arctic and boreal regions. Many of the eight factors identified by Fenton and Smith (1982) as being important for moss establishment in Antarctica, are sensitive to a changing climate, including length of growing season, reduction in permanent snow cover, substrate stability (extent of permafrost), wind speed, nutrient availability and water supply. Such conditions already partly explain the contrasting degree of moss colonisation between maritime and continental Antarctica, and the increased extent of moss bank formation in the Scotia Arc. Having reviewed the rates of moss bank growth in these habitats, we have also demonstrated the power of stable isotopes (¹³C, ¹⁸O) to integrate photosynthesis, growth and water sources. However, more work is needed to understand the biochemical and hydraulic determinants of oxygen and deuterium signals, and their relationship to climatic inputs. Photosystem II fluorescence was demonstrated to be one potential physiological proxy which integrates the effect of light intensity, precipitation/evaporation rates on photosynthetic activity and hence bryophyte growth and carbon sequestration. Once validated by ground truthing, solar-induced fluorescence, sensed remotely by UAVs or satellites, could be used to map bryophyte productivity in polar and boreal regions. For the arctic, it should be possible to capture the changing spatial landscape, as paludification in warming, maritime areas and increased intensity of fires in continental

- regions, are anticipated in the future. Additionally, the integration of surface proxies and a
 better understanding of stable isotope signals will also improve the historical reconstruction
 of climatic conditions for peat archives, as both temporal and spatial proxies will be needed
- to understand how polar-regions will respond to warming and other climate change effects.

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References

- Baltzer J. L., Veness T., Chasmer L. E., Sniderhan A. E., Quinton W. L. (2014) Forests on thawing permafrost: fragmentation, edge effects, and net forest loss. *Global Change Biology*, **20**, 824-834.
- Bañón M., Justel A., Velázquez D., Quesada A. (2013) Regional weather survey on Byers
 Peninsula, Livingston Island, South Shetland Islands, Antarctica. *Antarctic Science*,
 25, 146-156.
- Barbour M. M. (2007) Stable oxygen isotope composition of plant tissue: a review. *Functional Plant Biology*, **34**, 83-94.
- Barnes D. K. A., Hodgson D. A., Convey P., Allen C. S., Clarke A. (2006) Incursion and excursion of Antarctic biota: Past, present and future. *Global Ecology and Biogeography*, **15**, 121-142.
- Björck S., Hjort C., Ingolfsson O., Skog G. (1991a) Radiocarbon dates from the Antarctic Peninsula problems and potential. *Quaternary Proceedings*, **1**, 55-65.
- Björck S., Malmer N., Hjort C. *et al.* (1991b) Stratigraphic and Paleoclimatic Studies of a
 5500-Year-Old Moss Bank on Elephant Island, Antarctica. *Arctic and Alpine Research*, 23, 361-374.
- Block W. (1984) Terrestrial microbiology, invertebrates and ecosystems (Antarctic). In: *Antarctic Ecology*. (ed Laws RM) pp 163-236. London, Academic Press.
- Block W., Convey P. (2001) Seasonal and long-term variation in body-water content of an Antarctic springtail A response to climate change? *Polar Biology*, **24**, 764-770.
- Bockheim J., Vieira G., Ramos M. *et al.* (2013) Climate warming and permafrost dynamics in the Antarctic Peninsula region. *Global and Planetary Change*, **100**, 215-223.
- Bokhorst S., Huiskes A., Convey P., Aerts R. (2007) External nutrient inputs into terrestrial ecosystems of the Falkland Islands and the Maritime Antarctic region. *Polar Biology*, **30**, 1315-1321.

765

- Callaghan T. V., Björn L. O., Chapin III F. S. *et al.* (2005) Tundra and polar desert
 ecosystems. In: *Arctic Climate Impacts Assessment*. pp 243-352. Cambridge,
 Cambridge University Press.
- Cannone N., Convey P., Guglielmin M. (2013) Diversity trends of bryophytes in continental Antarctica. *Polar Biology*, **36**, 259-271.
- Cannone N., Ellis-Evans J. C., Strachan R. J., Guglielmin M. (2006) Interactions between
 climate, vegetation and the active layer in soils at two Maritime Antarctic sites.
 Antarctic Science, 18, 323-333.
- Chambers F. M., Booth R. K., De Vleeschouwer F. *et al.* (2011) Development and refinement of proxy-climate indicators from peats. *Quaternary International*, **268**, 21-33.
- 751 Chapin III F. S. (2003) Effects of plant traits on ecosystem and regional processes: A
 752 conceptual framework for predicting the consequences of global change. *Annals of*753 *Botany*, **91**, 455-463.
- Chapin III F. S., Berman M., Callaghan T. V. et al. (2005) Millennium Ecosystem
 Assessment: Polar Systems. In: Ecosystems and Human Well-being: Current State and
 Trends, Volume 1. (eds Hassan R, Scholes R, Ash N) pp 717-743. Washington D.C.,
 Island Press.
- 758 Chapin III F. S., McGuire A. D., Ruess R. W. *et al.* (2010) Resilience of Alaska's boreal forest to climatic change. *Canadian Journal of Forest Research*, **40**, 1360-1370.
 - Charman D. J., Beilman D. W., M. B. *et al.* (2013) Climate-related changes in peatland carbon accumulation during the last millennium. *Biogeosciences*, **10**, 929-944.
- Chown S. L., Convey P. (2007) Spatial and temporal variability across life's hierarchies in the
 terrestrial Antarctic. *Philosophical Transactions of the Royal Society B: Biological* Sciences, 362, 2307-2331.
 - Clarke L. J., Robinson S. A., Hua Q., Ayre D. J., Fink D. (2012) Radiocarbon bomb spike reveals biological effects of Antarctic climate change. *Global Change Biology*, **18**, 301-310.
- Colesie C., Green T. G. A., Türk R., Hogg I. D., Sancho L. G., Büdel B. (2014) Terrestrial
 biodiversity along the Ross Sea coastline, Antarctica: lack of a latitudinal gradient and
 potential limits of bioclimatic modeling. *Polar Biology*, 37, 1197-1208.
- Convey P. (2001) Terrestrial ecosystem response to climate changes in the Antarctic. In:
 "Fingerprints" of Climate Change Adapted Behaviour and Shifting Species Ranges.
 (eds Walther G-R, Burga CA, Edwards PJ). New York, Kluwer.
- Convey P. (2013) Antarctic Ecosystems. In: *Encyclopedia of Biodiversity*. (ed Levin SA).
 San Diego, Elsevier.
- Convey P., Chown S. L., Clarke A. *et al.* (2014) The spatial structure of Antarctic
 biodiversity. *Ecological Monographs*, 84, 203-244.
- Convey P., Hopkins D. W., Roberts S. J., Tyler A. N. (2011) Global southern limit of flowering plants and moss peat accumulation. *Polar Research*, 30, doi:10.3402/polar.v3430i3400.8929.
- Convey P., Smith R. L., Hodgson D. A., Peat H. (2000) The flora of the South Sandwich
 Islands, with particular reference to the influence of geothermal heating. *Journal of Biogeography*, **27**, 1279-1295.
- Convey P., Stevens M. I., Hodgson D. A. *et al.* (2009) Exploring biological constraints on the glacial history of Antarctica. *Quaternary Science Reviews*, **28**, 3035-3048.
- Cook A. J., Fox A. J., Vaughan D. G., Ferringo J. G. (2005) Retreating glacier fronts on the Antarctic Peninsula over the past half-century. *Science*, **308**, 541-544.
- 788 Crawford R. M. M. (2008) Cold climate plants in a warmer world. *Plant Ecology & Diversity*, **1**, 285-297.

- 790 Crawford R. M. M., Jeffree C. E., Rees W. G. (2003) Paludification and forest retreat in northern oceanic environments. *Annals of Botany*, **91**, 213-226.
- Da Silveira L., Sternberg L., Mulkey S. S., Wright S. J. (1989) Ecological Interpretation of Leaf Carbon Isotope Ratios: Influence of Respired Carbon Dioxide. *Ecology*, **70**, 1317-1324.
- Daley T. J., Barber K. E., Street-Perrott F. A., Loader N. J., Marshall D. J., Crowley S. F.,
 Fisher E. H. (2010) Holocene climate variability revealed by oxygen isotope analysis
 of Sphagnum cellulose from Walton Moss, northern England. *Quaternary Science Reviews*, 29, 1590-1601.
- Daley T. J., Thomas E. R., Holmes J. A. *et al.* (2011) The 8200yr BP cold event in stable isotope records from the North Atlantic region. *Global and Planetary Change*, **79**, 288-302.
- Damm A., Guanter L., Laurent V. C. E., Schaepman M. E., Schickling A., Rascher U. (2014)
 FLD-based retrieval of sun-induced chlorophyll fluorescence from medium spectral
 resolution airborne spectroscopy data. *Remote Sensing of Environment*, **147**, 256-266.
 - Davey M. C., Rothery P. (1997) Interspecific variation in respiratory and photosynthetic parameters in Antarctic bryophytes. *New Phytologist*, **137**, 231-240.

- Dawson T. E., Mambelli S., Plamboeck A. H., Templer P. H., Tu K. P. (2002) Stable isotopes in plant ecology. *Annual Review of Ecology and Systematics*, **33**, 507-559.
- DeNiro M. J., Epstein S. (1979) Relationship between the oxygen isotope ratios of terrestrial plant cellulose, carbon dioxide and water. *Science*, **204**, 51-53.
- Edwards T. W. D. (1993) Interpreting past climate from stable isotopes in continental organic
 matter. In: *Climate Change in Continental Isotopic Records*. (eds Swart PK,
 Mckenzie JE, Lohmann KC, Savin S). Washington D. C., American Geophysical
 Union.
- Edwards T. W. D., Graf W., Trimborn P., Stichler W., Lipp J., Payer H. D. (2000) δ¹³C
 response surface resolves humidity and temperature signals in trees. *Geochimica at Cosmochimica Acta*, 64, 161-167.
- Elmendorf S. C., Henry G. H. R., Hollister R. D. *et al.* (2012) Global assessment of experimental climate warming on tundra vegetation: Heterogeneity over space and time. *Ecology Letters*, **15**, 164-175.
- Fabiszewski J., Wojtun B. (1997) The occurrence and development of peat mounds on King George Island. *Acta Societatis Botanicorum Poloniae*, **66**, 223-229.
- Farquhar G. D., Ehleringer J. R., Hubick K. T. (1989) Carbon isotope discrimination and photosynthesis. *Annual Review of Plant Physiology*, **40**, 503-537.
- Favero-Longo S. E., Cannone N., Worland M. R., Convey P., Piervittori R., Guglielmin M. (2011) Change in lichen diversity and community structure with fur seal population increase on Signy Island, South Orkney Islands. *Antarctic Science*, **23**, 65-77.
- Fenton J. H. C. (1980) The rate of peat accumulation in Antarctic moss banks. *Journal of Ecology*, **68**, 211-228.
- Fenton J. H. C. (1982) The formation of vertical edges on Antarctic moss peat banks. *Arctic* and *Alpine Research*, **14**, 21-26.
- Fenton J. H. C., Smith R. L. (1982) Distribution, composition and general characteristics of the moss banks of the maritime Antarctic. *British Antarctic Survey Bulletin*, **51**, 215-236.
- Fenton N. J., Bergeron Y. (2006) Facilitative succession in a boreal bryophyte community driven by changes in available moisture and light. *Journal of Vegetation Science*, **17**, 65-76.

856

857

863

864

865

866

- Franzke C. (2012) Significant reduction of cold temperature extremes at Faraday/Vernadsky station in the Antarctic Peninsula. *International Journal of Climatology*, **33**, 1070-1078.
- Fretwell P. T., Convey P., Fleming A. H., Peat H. J., Hughes K. A. (2011) Detecting and mapping vegetation distribution on the Antarctic Peninsula from remote sensing data. *Polar Biology*, **34**, 273-281.
- Friedli H., Lotscher H., Oeschger H., Siegenthaler U. (1986) Ice core record of the ¹³C/¹²C ratio of atmospheric CO₂ in the past two centuries. *Nature*, **324**, 237-238.
- Gallego-Sala A. V., Prentice I. C. (2013) Blanket peat biome endangered by climate change. *Nature Clim. Change*, **3**, 152-155.
- Gat J. R. (2000) Atmospheric water balance the isotopic perspective. *Hydrological Processes*, **14**, 1357-1369.
- Gimingham C. H., Smith R. L. (1971) Growth form and water relations of mosses in the Maritime Antarctic. *British Antarctic Survey Bulletin*, **25**, 1-21.
- Green T. G. A., Sancho L. G., Pintado A., Schroeter B. (2011) Functional and spatial pressures on terrestrial vegetation in Antarctica forced by global warming. *Polar Biology*, **34**, 1643-1656.
 - Guanter L., Zhang Y., Jung M. *et al.* (2014) Global and time-resolved monitoring of crop photosynthesis with chlorophyll fluorescence. *Proceedings of the National Academy of Sciences*, **111**, E1327-E1333.
- Guglielmin M., Dalle Fratte M., Cannone N. (2014) Permafrost warming and vegetation changes in continental Antarctica. *Environmental Research Letters*, **9**, 045001.
- Guglielmin M., Worland M. R., Cannone N. (2012) Spatial and temporal variability of ground surface temperature and active layer thickness at the margin of maritime Antarctica, Signy Island. *Geomorphology*, **155-156**, 20-33.
 - Helliker B. R., Griffiths H. (2007) Toward a plant-based proxy for the isotope ratio of atmospheric water vapor. *Global Change Biology*, **13**, 723-733.
 - Hill P. W., Farrar J., Roberts P. *et al.* (2011) Vascular plant success in a warming Antarctic may be due to efficient nitrogen acquisition. *Nature Climate Change*, **1**, 50-53.
 - Hogg A. G., Hua Q., Blackwell P. G. *et al.* (2013) SHCal13 Southern Hemisphere calibration, 0-50,000 years cal BP. *Radiocarbon*, **55**, 1889-1903.
- Kaislahti Tillman P., Holzkamper S., Kuhry P., Sannel A. B. K., Loader N. J., Robertson I. (2010) Long-term climate variability in continental subarctic Canada: A 6200-year record derived from stable isotopes in peat. *Palaeogeography Palaeoclimatology Palaeoecology*, **298**, 235-246.
- Kirchgaßner A. (2010) An analysis of cloud obervations from Vernadsky, Antarctica. *International Journal of Climatology,* **30**, 1431-1439.
- Kolber Z., Klimov D., Ananyev G., Rascher U., Berry J., Osmond B. (2005) Measuring
 photosynthetic parameters at a distance: Laser induced fluorescence transient (LIFT)
 method for remote measurements of photosynthesis in terrestrial vegetation.
 Photosynthesis Research, 84, 121-129.
- Korhonen H., Carslaw K. S., Forster P. M., Mikkonen S., Gordon N. D., Kokkola H. (2010)
 Aerosol climate feedback due to decadal increases in Southern Hemisphere wind
 speeds. *Geophysical Research Letters*, **37**, L02805.
- Koven C. D., Ringeval B., Friedlingstein P. *et al.* (2011) Permafrost carbon-climate feedbacks accelerate global warming. *Proceedings of the National Academy of Sciences*, **108**, 14769-14774.
- La Farge C., Williams K. H., England J. H. (2013) Regeneration of Little Ice Age bryophytes emerging from a polar glacier with implications of totipotency in extreme environments. *Proceedings of the National Academy of Sciences*, **110**, 9839-9844.

- Lang S. I., Cornelissen J. H. C., Hölzer A., Ter Braak C. J. F., Ahrens M., Callaghan T. V.,
 Aerts R. (2009) Determinants of cryptogam composition and diversity in *Sphagnum*dominated peatlands: The importance of temporal, spatial and functional scales. *Journal of Ecology*, **97**, 299-310.
- Lang S. I., Cornelissen J. H. C., Shaver G. R. *et al.* (2012) Arctic warming on two continents has consistent negative effects on lichen diversiry and mixed effects on bryophyte diversity. *Global Change Biology*, **18**, 1096-1107.
- Loader N. J., Santillo P. M., Woodman-Ralph J. P. *et al.* (2008) Multiple stable isotopes from oak trees in southwestern Scotland and the potential for stable isotope dendroclimatology in maritime climatic regions. *Chemical Geology*, **252**, 62-71.
- Loisel J., Gallego-Sala A. V., Yu Z. (2012) Global-scale pattern of peatland Sphagnum growth driven by photosynthetically active radiation and growing season length. *Biogeosciences*, **9**, 2737-2746.
- Loisel J., Yu Z. (2013) Holocene peatland carbon dynamics in Patagonia. *Quaternary Science Reviews*, 69, 125-141.
- Longton R. E. (1982) Bryophyte Vegetation in Polar Regions. In: *Bryophyte Ecology*. (ed
 Smith AJE) pp 123-165. London, Chapman and Hall.
- Longton R. E. (1988) *Biology of polar bryophytes and lichens*, Cambridge, Cambridge
 University Press.
- Lovelock C. E., Robinson S. A. (2002) Surface reflectance properties of Antarctic moss and their relationship to plant species, pigment composition and photosynthetic function. *Plant, Cell and Environment,* **25**, 1239-1250.
- Lucieer A., Turner D., King D. H., Robinson S. A. (2013) Using an Unmanned Aerial
 Vehicle (UAV) to capture micro-topography of Antarctic moss beds. *ITC Journal*, 27,
 53-62.
- Lynch H. J., Naveen R., Trathan P. N., Fagan W. F. (2012) Spatially integrated assessment
 reveals widespread changes in penguin populations on the Antarctic Peninsula.
 Ecology, 93, 1367-1377.
- McGuire D. A., Anderson L. G., Christensen T. R. *et al.* (2009) Sensitivity of the carbon cycle in the Arctic to climate change. *Ecological Monographs*, **79**, 523-555.
- Melick D. R., Seppelt R. D. (1997) Vegetation Patterns in Relation to Climatic and
 Endogenous Changes in Wilkes Land, Continental Antarctica. *Journal of Ecology*, 85,
 43-56.
- 921 Ménot-Combes G., Burns S. J., Leuenberger M. (2002) Variations of ¹⁸O/¹⁶O in plants from 922 temperate peat bogs (Switzerland): implications for palaeoclimatic studies. *Earth and* 923 *Planetary Science Letters*, **202**, 419-434.
- Ménot G., Burns S. J. (2001) Carbon isotopes in ombrogenic peat bog plants as climatic
 indicators: calibration from an altitudinal transect in Switzerland. *Organic Geochemistry*, 32, 233-245.
- Meyer M., Seibt U., Griffiths H. (2008) To concentrate or ventilate? Carbon acquisition,
 isotope discrimination and physiological ecology of early land plant forms.
 Philosophical Transactions of the Royal Society B: Biological Sciences, 363, 2767-2778.
- Minayeva T., Sirin A. (2010) Arctic peatlands. In: Arctic Biodiversity Trends 2010 Selected
 indicators of change. (ed.) pp Page, Akureyri, Iceland, CAFF International
 Secretariat.
- Nijp J. J., Limpens J., Metselaar K., van der Zee S. E. A. T. M., Berendse F., Robroek B. J.
- 935 M. (2014) Can frequent precipitation moderate the impact of drought on peatmoss carbon uptake in northern peatlands? *New Phytologist*, **203**, 70-80.
- O'Leary M. H. (1988) Carbon isotopes in photosynthesis. *BioScience*, **38**, 328-336.

950

- Ochyra R., Bednarek-Ochyra H., Smith R. I. L. (2008) *Illustrated Moss Flora of Antarctica*, Cambridge, Cambridge University Press.
- Orsenigo S., Mondoni A., Rossi G., Abeli T. (2014) Some like it hot and some like it cold, but not too much: plant responses to climate extremes. *Plant Ecology*, **215**, 677-688.
- Pannewitz S., Green T. G. A., Maysek K. *et al.* (2005) Photosynthetic responses of three common mosses from continental Antarctica. *Antarctic Science*, **17**, 341-352.
- Peat H. J., Clarke A., Convey P. (2007) Diversity and biogeography of the Antarctic flora.
 Journal of Biogeography, 34, 132-146.
- Pendall E., Markgraf V., White J. W. C., Dreier M., Kenny R. (2001) Multiproxy record of
 Late Pleistocene-Holocene climate and vegetation changes from a peat bog in
 Patagonia. *Quaternary Research*, 55, 168-178.
 - Pieruschka R., Klimov D., Berry J. A., Osmond C. B., Rascher U., Kolber Z. S. (2012) Remote chlorophyll fluorescence measurements with the laser-induced fluorescence transient approach. *Methods in Molecular Biology*, **918**, 51-59.
- Piotrowska N., Blaauw M., Mauquoy D., Chambers F. M. (2011) Constructing deposition chronologies for peat deposits using radiocarbon dating. *Mires and Peat*, 7, Art. 10.
- Proctor M. C. F. (2000) The bryophyte paradox: tolerance of desiccation, evasion of drought.
 Plant Ecology, **151**, 41-49.
- Proctor M. C. F., Tuba Z. (2002) Poikilohydry and homoihydry: antithesis or spectrum of possibilities? *New Phytologist*, **156**, 327-349.
- Quayle W. C., Peck L. S., Peat H., Ellis-Evans J. C., Harrigan P. R. (2002) Extreme responses to climate change in Antarctic lakes. *Science*, **295**, 645.
- Quinton W. L., Hayashi M., Chasmer L. E. (2011) Permafrost-thaw-induced land-cover
 change in the Canadian subarctic: implications for water resources. *Hydrological Processes*, 25, 152-158.
- Rice S. K., Giles L. (1996) The influence of water content and leaf anatomy on carbon isotope discrimination and photosynthesis in *Sphagnum*. *Plant, Cell and Environment*, **19**, 118-124.
- Roads E., Longton R. E., Convey P. (2014) Millennial timescale regeneration in a moss from Antarctica. *Current biology : CB*, **24**, R222-R223.
- Robinson S. A., Erickson III D. J. (in press) Not just about sunburn the ozone hole's
 profound effect on climate has significant implications for Southern Hemisphere
 ecosystems. *Global Change Biology*.
- Robinson S. A., Wasley J., Popp M., Lovelock C. E. (2000) Desiccation tolerance of three
 moss species from continental Antarctica. *Australian Journal of Plant Physiology*, 27,
 379-388.
- Royles J. (2012) Environmental isotopic records preserved in Antarctic peat moss banks.
 Unpublished PhD University of Cambridge, Cambridge, UK.
- Royles J., Amesbury M. J., Convey P., Griffiths H., Hodgson D. A., Leng M. J., Charman D.
 J. (2013a) Plants and soil microbes respond to recent warming on the Antarctic
 Peninsula. *Current Biology*, 23, 1702-1706.
- Royles J., Ogée J., Wingate L., Hodgson D. A., Convey P., Griffiths H. (2012) Carbon isotope evidence for recent climate-related enhancement of CO₂ assimilation and peat accumulation rates in Antarctica. *Global Change Biology*, **18**, 3112-3124.
- Royles J., Ogée J., Wingate L., Hodgson D. A., Convey P., Griffiths H. (2013b) Temporal separation between CO2 assimilation and growth? Experimental and theoretical evidence from the desiccation tolerant moss *Syntrichia ruralis*. *New Phytologist*, **197**, 1152-1160.
- Royles J., Sime L. C., Hodgson D. A., Convey P., Griffiths H. (2013c) Differing source water
 inputs, moderated by evaporative enrichment, determine the contrasting

- 988 δ^{18} O_{CELLULOSE} signals in maritime Antarctic moss peat banks. *Journal of Geophysical* 989 *Research: Biogeosciences*, **118**, 184-194.
- Rubino M., Etheridge D. M., Trudinger C. M. *et al.* (2013) A revised 1000 year atmospheric
 δ13 C-CO2 record from Law Dome and South Pole, Antarctica. *Journal of Geophysical Research D: Atmospheres*, 118, 8482-8499.
- 993 SCAR (2014) Antarctic Climate Data: READER project. pp Page, British Antarctic Survey (http://www.antarctica.ac.uk/met/READER/).
- 995 Schlensog M., Green T. G. A., Schroeter B. (2013) Life form and water source interact to 996 determine active time and environment in cryptogams: an example from the maritime 997 Antarctic. *Oecologia*, **173**, 59-72.
- 998 Schlensog M., Pannewitz S., Green T. G. A., Schroeter B. (2004) Metabolic recovery of continental antarctic cryptogams after winter. *Polar Biology*, **27**, 399-408.
- Schuur E. A. G., Vogel J. G., Crummer K. G., Lee H., Sickman J. O., Osterkamp T. E. (2009)
 The effect of permafrost thaw on old carbon release and net exchange from tundra.

 Nature, 459, 556-559.
- Sistla S. A., Moore J. C., Simpson R. T., Gough L., Shaver G. R., Schimel J. P. (2013) Longterm warming restructures Arctic tundra without changing net soil carbon storage. *Nature*, **497**, 615-618.
- Smith R. I. L. (1988a) Destruction of Antarctic terrestrial ecosystems by a rapidly increasing fur seal population. *Biological Conservation*, **45**, 55-72.
- Smith R. I. L. (1988b) Recording bryophyte microclimate in remote and severe environments. In: *Methods in Bryology*. (ed Glime JM) pp 275-284. Nichinan, Hattori Bot. Lab.
- Smith R. I. L. (1990) Signy Island as a paradigm of biological and environmental change in Antarctic terrestrial ecosystems. In: *Antarctic Ecosystems, Ecological Change and* Conservation. (eds Kerry KR, Hempel G) pp 32-50. Berlin, Springer.
- Smith R. I. L., Clymo R. S. (1984) An extraordinary peat-forming community on the Falkland Islands. *Nature*, **309**, 617-620.
- Smith R. I. L., Corner R. W. M. (1973) Vegetation of the Arthur Harbour-Argentine Islands region of the Antarctic Peninsula. *British Antarctic Survey Bulletin*, **33-34**, 89-122.
- Smith R. L. (1972) Vegetation of the South Orkney Islands with particular reference to Signy Island. In: *British Antarctic Survey Scientific Reports 68.* pp Page.
- Stanton D. E., Merlin M., Bryant G., Ball M. C. (2014) Water redistribution determines photosynthetic responses to warming and drying in two polar mosses. *Functional Plant Biology*, **41**, 178-186.
- Sternberg L., Ellsworth P. F. V. (2011) Divergent biochemical fractionation, not convergent temperature, explains cellulose oxygen isotope enrichment across latitudes. *PLoS ONE*, **10.1371/journal.pone.0028040**.
- Sternberg L., Pinzon M. C., Anderson W. T., Jahren A. H. (2006) Variation in oxygen isotope fractionation during cellulose synthesis: intramolecular and biosynthetic effects. *Plant, Cell and Environment,* **29**, 1881-1889.
- Strauss S. L., Ruhl C. T., Day T. A. (2009) Trends in soil characteristics along a recently deglaciated foreland on Anvers Island, Antarctic Peninsula. *Polar Biology*, **32**, 1779-1031
- Street L. E., Subke J.-A., Sommerkorn M., Sloan V., Ducrotoy H., Phoenix G. K., Williams M. (2013) The role of mosses in carbon uptake and partitioning in arctic vegetation.

 New Phytologist, 199, 163-175.
- Stuiver M., Reimer P. J. (1993) Extended ¹⁴C database and revised CALIB radiocarbon calibration program. *Radiocarbon*, **35**, 215-230.

- Surdu C. M., Duguay C. R., Brown L. C., Fernández Prieto D. (2014) Response of ice cover
 on shallow lakes of the North Slope of Alaska to contemporary climate conditions
 (1950-2011): radar remote-sensing and numerical modeling data analysis. *The Cryosphere*, 8, 167-180.
- Tingley M. P., Huybers P. (2013) Recent temperature extremes at high northern latitudes unprecedented in the past 600 years. *Nature*, **496**, 201-205.
- Turetsky M. R., Bond-Lamberty B., Euskirchen E., Talbot J., Frolking S., McGuire A. D.,
 Tuittila E. S. (2012) The resilience and functional role of moss in boreal and arctic
 ecosystems. *New Phytologist*, **196**, 49-67.
- Turetsky M. R., Mack M. C., Hollingsworth T. N., Harden J. W. (2010) The role of mosses in ecosystem succession and function in Alaska's boreal forest. *Canadian Journal of Forest Research*, **40**, 1237-1264.
- Turetsky M. R., Wieder R. K., Vitt D. H., Evans R. J., Scott K. D. (2007) The disappearance of relict permafrost in boreal North America: effects of peatland carbon storage and fluxes. *Global Change Biology*, **13**, 1922-1934.
- Turner D., Lucieer A., Malenovský Z., King D. H., Robinson S. A. (2014) Spatial coregistration of ultra-high resolution visible, multispectral and thermal images acquired with a micro-UAV over antarctic moss beds. *Remote Sensing*, **6**, 4003-4024.
- Turner J., Bindschadler R., Convey P. *et al.* (2009) *Antarctic climate change and the environment*, Cambridge, Scientific Committee on Antarctic Research.
- Van der Putten N., Mauquoy D., Verbruggen C., Björck S. (2012) Subantarctic peatlands and their potential as palaeoenvironmental and palaeoclimatic archives. *Quaternary International*, **268**, 65-76.
- Wall D. H., Berry Lyons W., Chown S. L., Convey P., Howard-Williams C., Quesada A.,
 Vincent W. F. (2011) Long-term ecosystem networks to record change: An
 international imperative. *Antarctic Science*, 23, 209.
- Waluda C. M., Gregory S., Dunn M. J. (2010) Long-term variability in the abundance of Antarctic fur seals Arctocephalus gazella at Signy Island, South Orkneys. *Polar Biology*, **33**, 305-312.
- Wasley J., Robinson S. A., Lovelock C. E., Popp M. (2006) Some like it wet Biological
 characteristics underpinning tolerance of extreme water stress events in Antarctic
 bryophytes. *Functional Plant Biology*, 33, 443-455.
- Wasley J., Robinson S. A., Turnbull J. D., King D. H., Wanek W., Popp M. (2012)
 Bryophyte species composition over moisture gradients in the Windmill Islands, East
 Antarctica: Development of a baseline for monitoring climate change impacts.
 Biodiversity, 13, 257-264.
- Williams T. G., Flanagan L. B. (1996) Effects of changes in water content on photosynthesis, transpiration and discrimination against ¹³CO₂ and C¹⁸O¹⁶O in *Pleurozium* and *Sphagnum. Oecologia*, **108**, 38-46.
- 1076 Xu L., Myneni R. B., Chapin Iii F. S. *et al.* (2013) Temperature and vegetation seasonality diminishment over northern lands. *Nature Clim. Change*, **3**, 581-586.
- Yu Z., Loisel J., Brosseau D. P., Beilman D. W., Hunt S. J. (2010) Global peatland dynamics since the Last Glacial Maximum. *Geophysical Research Letters*, **37**, LI3402.
- Zhang Y., Guanter L., Berry J. A. *et al.* (2014) Estimation of vegetation photosynthetic
 capacity from space-based measurements of chlorophyll fluorescence for terrestrial
 biosphere models. *Global Change Biology*, DOI: 10.1111/gcb.12664.

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Figure Legends

Figure 1 a) Mean growing season temperatures (November-March) over fifty years at meteorological stations around Antarctica (SCAR, 2014). Box encompasses maritime Antarctic (M), remainder is continental Antarctica (C); b) Major sites of moss peat banks and the associated oldest basal dates. At Signy Island (Fenton, 1982), Elephant Island (Björck *et al.*, 1991b), Norsel Point and Lazarev Bay (Convey *et al.*, 2000, Royles *et al.*, 2013a) the growth is continuous and consists of *Chorisodontium aciphyllum* and/or *Polytrichum strictum*. Norsel Point sample processed following same methods as (Royles *et al.*, 2012). All dates re-calibrated from published ¹⁴C ages using Caib (v. 7.0.2) (Stuiver & Reimer, 1993) and the SHCall3 dataset (Hogg *et al.*, 2013). Dates presented are the median probability years, with associated 2σ ranges in brackets.

Figure 2: a) Signy Island *Chorisodontium aciphyllum* moss peat bank; b) Cross section through top of *C. aciphyllum* core showing individual moss stems aligned below 5 mm of green photosynthetic tissue (scale bar represents 5 cm); c) Ridged surface of *C. aciphyllum* moss bank with moribund areas and epilithic lichens (scale bar represents 10 cm; d) Signy Island *C. aciphyllum* moss peat bank; e) Cross-section through deep *C. aciphyllum* peat; f) Signy Island *C. aciphyllum* hummock.

Figure 3: Age-depth models developed from bomb-spike dating of moss growth from the Windmill Islands (*Ceratodon purpureus*, 3 blue lines) and Vestfold Hills (*Bryum pseudotriqutrum* and *Bryoerythrophyllum recurvirostre*, 2 purple lines) (Clarke *et al.*, 2012) in continental Antarctica (C) and from the maritime Antarctic (M) *Chorisodontium aciphyllum* from Signy Island (green line) (Royles *et al.*, 2012) and *Polytrichum strictum* from Lazarev Bay (red line) (Royles *et al.*, 2013a).

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1110	Figure 4: Peat accumulation rate over time on Signy Island and at Lazarev Bay (Royles et al.,
1111	2013a). Calculated from bulk density measurements and age depth model, smoothed over
1112	three successive time-points, this takes into account the increasing effect of compression as
1113	the accumulations become deeper.
1114	
1115	Figure 5a): Measured $\delta^{13}C$ of Ceratodon purpureus (3 blue lines) organic matter from
1116	continental (C) Antarctica (Clarke et al., 2012). In the maritime Antarctic (M) at Lazarev Bay
1117	(red) (Royles $\it{et~al.}$, 2013a) and Signy Island (green) (Royles $\it{et~al.}$, 2012) $\delta^{13}C$ was measured
1118	in cellulose. Comparative measurements of organic matter and cellulose showed a highly
1119	significant linear relationship between $\delta^{13}C_{OM}$ and $\delta^{13}C_{C}$ (R²=0.73, p<0.0001, n=40, F=108,
1120	y=0.75x $-$ 4.58) from which $\delta^{13}C_{OM}$ were derived; b) Source-independent discrimination
1121	$(\Delta^{13} \text{C})$ was calculated for each measurement (see text for details; NB. Different y-axis
1122	scales).
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1124	Figure 6a) Measured carbon isotope composition of cellulose $(\delta^{13}C_{\text{C}})$ and b) Source
1125	independent discrimination ($\Delta^{13}C$; see text for details) over time from Signy Island
1126	Chorisodontium aciphyllum (3 green lines) (Royles et al., 2012) and Lazarev Bay
1127	Polytrichum strictum (red) (Royles et al., 2013a).
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1129	Figure 7: Extent of source independent carbon ($\Delta^{13}C$) and oxygen ($\delta^{18}O$) isotope
1130	discriminations measured in cellulose for Signy Island Chorisodontium aciphyllum growing

in large banks (green diamonds, orange circles) and a small hummock (blue upward triangle).

1132	In addition, Polytrichum strictum hummocks from Signy Island (Red down triangles) and
1133	Lazarev Bay (purple squares). $\delta^{13}C$ and $\delta^{18}O$ measured on separate sub-samples of cellulose
1134	as described in (Royles et al., 2013a, Royles et al., 2012, Royles et al., 2013c). Arrows
1135	represent direction and hypothetical mechanisms for isotopic variation.
1136	Figure 8: Desiccation response of <i>Syntrichia ruralis</i> : a) Max electron transport rate (ETR) as
1137	a function of relative water content (RWC). Error bars: 1 SE (n=4); b) Light response curves
1138	for desiccating Syntrichia ruralis: mean ETR as a function of light intensity. Six successive
1139	curves were measured as the tissue RWC declined from 350% (line 1, red) through to 50%
1140	(purple, line 6). Equivalent RWC (1-6) marked on (a) (n=4, dashed lines=SE)
1141	

1142

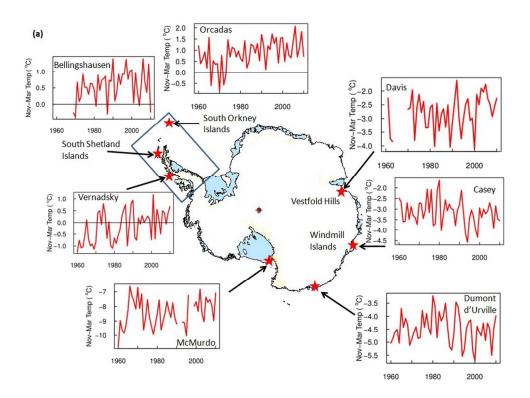


Figure 1a) Map of Antarctica and summer temperature trends $261 \times 189 \, \text{mm}$ (96 x 96 DPI)

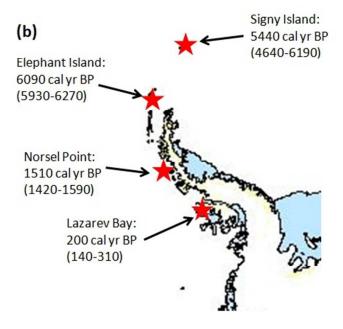


Figure 1b) Basal age of maritime Antarctic peat banks $179 \times 112 \text{mm}$ (96 x 96 DPI)

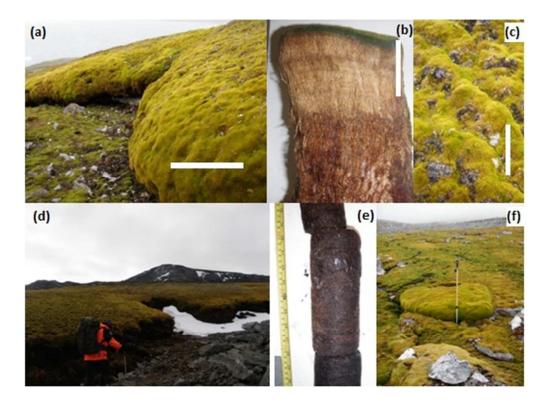


Figure 2 a) Signy Island Chorisodontium aciphyllum moss peat bank; b) Cross section through top of C. aciphyllum core showing individual moss stems aligned below 5 mm of green photosynthetic tissue (scale bar represents 5 cm); c) Ridged surface of C. aciphyllun moss bank with moribund areas and epilithic lichens (scale bar represents 10 cm; d) Signy Island C. aciphyllum moss peat bank; e) Cross-section through deep C. aciphyllum peat; f) Signy Island C. aciphyllum hummock .

136x100mm (96 x 96 DPI)

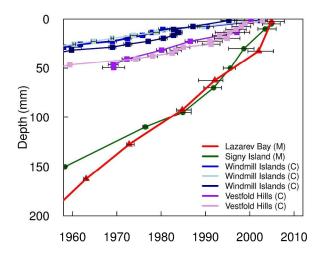


Figure 3) Age-depth models developed from bomb-spike dating of moss growth from the Windmill Islands (Ceratodon purpureus, 3 blue lines) and Vestfold Hills (Bryum pseudotriqutrum and Bryoerythrophyllum recurvirostre, 2 purple lines) (Clarke et al., 2012) in continental Antarctica (C) and from the maritime Antarctic (M) Chorisodontium aciphyllum from Signy Island (green line) (Royles et al., 2012) and Polytrichum strictum from Lazarev Bay (red line) (Royles et al., 2013a).

279x361mm (300 x 300 DPI)

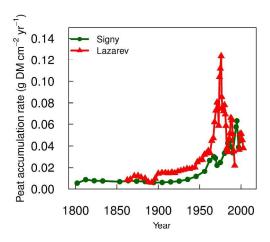


Figure 4) Peat accumulation rate over time on Signy Island and at Lazarev Bay (Royles et al., 2013a). Calculated from bulk density measurements and age depth model, smoothed over three successive time-points, this takes into account the increasing effect of compression as the accumulations become deeper. $279 \times 361 \text{mm}$ (300 x 300 DPI)

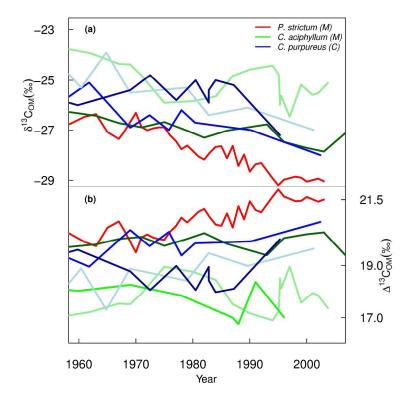


Figure 5a): Measured δ 13C of Ceratodon purpureus (3 blue lines) organic matter from continental (C) Antarctica (Clarke et al., 2012). In the maritime Antarctic (M) at Lazarev Bay (red) (Royles et al., 2013a) and Signy Island (green) (Royles et al., 2012) δ 13C was measured in cellulose. Comparative measurements of organic matter and cellulose showed a highly significant linear relationship between δ 13COM and δ 13CC (R2=0.73, p<0.0001, n=40, F=108, y=0.75x - 4.58) from which δ 13COM were derived; b) Source-independent discrimination (Δ 13C) was calculated for each measurement (see text for details; NB. Different y-axis scales).

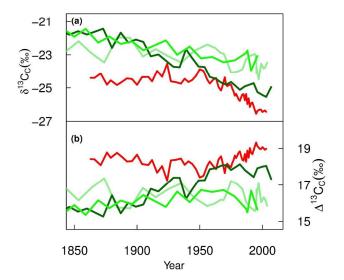


Figure 6a) Measured carbon isotope composition of cellulose (δ 13CC) and b) Source independent discrimination (Δ 13C; see text for details) over time from Signy Island Chorisodontium aciphyllum (3 green lines) (Royles et al., 2012) and Lazarev Bay Polytrichum strictum (red) (Royles et al., 2013a). 279x361mm (300 x 300 DPI)

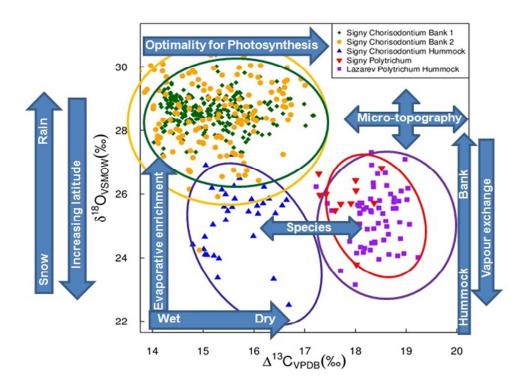


Figure 7: Extent of source independent carbon ($\Delta 13C$) and oxygen ($\delta 18O$) isotope discriminations measured in cellulose for Signy Island Chorisodontium aciphyllum growing in large banks (green diamonds, orange circles) and a small hummock (blue upward triangle). In addition, Polytrichum strictum hummocks from Signy Island (Red down triangles) and Lazarev Bay (purple squares). $\delta 13C$ and $\delta 18O$ measured on separate sub-samples of cellulose as described in (Royles et al., 2013a, Royles et al., 2012, Royles et al., 2013c). Arrows represent direction and hypothetical mechanisms for isotopic variation. $179 \times 128 \, \text{mm}$ (96 x 96 DPI)

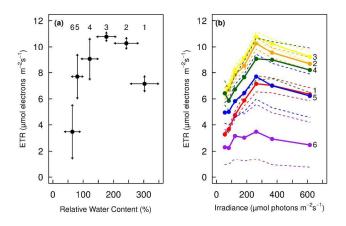


Figure 8: Desiccation response of Syntrichia ruralis: a) Max electron transport rate (ETR) as a function of relative water content (RWC). Error bars: 1 SE (n=4); b) Light response curves for desiccating Syntrichia ruralis: mean ETR as a function of light intensity. Six successive curves were measured as the tissue RWC declined from 350% (line 1, red) through to 50% (purple, line 6). Equivalent RWC (1-6) marked on (a) (n=4, dashed lines = SE) $279 \times 361 \text{mm} (300 \times 300 \text{ DPI})$