Role of the potential landscape on the single-file diffusion through channels

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Transport of colloid particles through narrow channels is ubiquitous in cell biology as well as becoming increasingly important for microfluidic applications or targeted drug delivery. Membrane channels in cells are useful models for artificial designs because of their high efficiency, selectivity and robustness to external fluctuations. Here we model the passive channels that let cargo simply diffuse through them, affected by a potential profile along the way. Passive transporters achieve high levels of efficiency and specificity from binding interactions with the cargo inside the channel. This however leads to a paradox: why should channels which are so narrow that they are blocked by their cargo evolve to have binding regions for their cargo if that will effectively block them? Using Brownian dynamics simulations, we show that different potentials, notably symmetric, increase the flux through narrow passive channels – and investigate how shape and depth of potentials influence the flux. We find that there exist optimal depths for certain potential shapes and that it is most efficient to apply a small force over an extended region of the channel. On the other hand, having several spatially discrete binding pockets will not alter the flux significantly. We also explore the role of many-particle effects arising from pairwise particle interactions with their neighbours and demonstrate that the relative changes in flux can be accounted for by the kinetics of the absorption reaction at the end of the channel.

PACS numbers: 83.10.Mj, 83.10.Rs, 87.16.dp

I. INTRODUCTION

Transport of macromolecules through nano-sized pores and narrow protein channels is essential for cell function¹ while also becoming increasingly important in microfluidic applications^{2,3} or to understand drug delivery.⁴ Channels in cell membranes are remarkable for their high efficiency, selectivity and robustness with respect to fluctuations of their environment⁵ and come in two flavours. Active transporters move their cargo by using cellular energy, e.g. from hydrolysing adenosine triphosphate or by harvesting concentration gradients of cell metabolites across the membrane. On the other hand, passive transporters are driven by the growth of entropy of the system as they translocate their specific cargo. Initially thought of as molecular sieves that select via the pore size to let the ‘right’ cargo simply diffuse through the channel, it is now well established that passive transporters achieve high levels of efficiency and specificity from binding interactions with the cargo inside the channel. A well-characterised example is the bacterial channel Maltoporin, where oligosaccharide transport is facilitated by an extended binding region.⁶ Although many more examples of this phenomenon have since been discovered using a plethora of methods (e.g. ex-situ crystallographic studies,⁷ indirect measurements of ionic currents⁸ and molecular dynamics simulations,⁹) the exact details of the mechanisms of passive transporters are still poorly understood.¹⁰

Our work is motivated by a seeming paradox that arises when one considers the flux through a narrow channel, such as Maltoporin, which prevents particles from overtaking each other. Increasing the binding affinity between the channel interior and the cargo will prolong the time each particle spends inside the channel, hence reduc-

ing the flux and effectively blocking it. Why then would channels evolve to have binding regions for the molecules they have evolved to translocate? In this paper, we combine the results of Brownian dynamics simulations with theoretical arguments to show how developing binding regions inside a channel can indeed increase flux through narrow channels. We model the binding regions using a variety of potential-energy landscapes along the channel and investigate the dependence of the flux through the channel on the shape and depth of these potentials.

We first consider a single freely diffusing particle to tune our Brownian dynamics simulations in the setting where an exact analytical solution for the transport exists, applying various tests to the simulation procedure to ensure its proper reflection of the physical situation. We then investigate single-file diffusion through a channel to analyse the dependence of particle flux on the shape and depth of applied potentials. Finally, we demonstrate that we can account for the relative changes in flux by considering the diffusion-limited reaction kinetics of the absorption in a scheme¹¹ based on the osmotic pressure along the channel alone.

In order to investigate the dynamics of colloid particles diffusing freely or in a potential, we need to solve the Langevin or corresponding Fokker-Planck equation. While there exist elegant analytical solutions for the transport of *non-interacting* particles,¹² here we consider the many-body problem of transport through crowded channels, which is a more realistic boundary condition for cellular environments¹³ or microfluidic devices¹⁴. We perform Brownian dynamics (BD) simulations¹⁵ using the LAMMPS package¹⁶ rather than using mean-field approaches¹⁷ for this genuine many-body problem.

II. FREE BROWNIAN PARTICLE

The one-dimensional motion of a Brownian particle is described by the Langevin equation $m\dot{v} + \alpha v = \zeta(t)$, where m is the particle mass, v its velocity and α is the viscous drag coefficient. We further assume white noise $\zeta(t)$ with $\langle \zeta(t)\zeta(t') \rangle = \Gamma\delta(t-t')$, where $\Gamma = 2\alpha k_B T/m$ is the intensity of the stochastic force, satisfying the fluctuation-dissipation theorem. The general solution for the root mean square displacement of the particle is¹⁸:

$$\langle \Delta x^2 \rangle = \frac{2k_B T m}{\alpha^2} (t/\tau_v - 1 + \exp(-t/\tau_v)) \quad (1)$$

with the velocity relaxation time $\tau_v = m/\alpha$. In the overdamped (or diffusive) regime, with $t \gg \tau_v$, we recover the Einstein result: $\langle \Delta x^2 \rangle = 2(k_B T/\alpha)t$, where one defines the diffusion constant $D \equiv k_B T/\alpha$. On the other hand, in the inertial regime $t \ll \tau_v$, the displacement grows linearly with time: $\langle \Delta x^2 \rangle = (k_B T/m)t^2$.

Brownian dynamics simulation

To verify that LAMMPS yields particle trajectories with the right statistical properties, we first simulated freely diffusing spherical Brownian particles of different sizes at different temperatures. The goal was to identify the inertial and the diffusive regimes and the fluctuation-dissipation theorem.

Integration of the Langevin equation and the application of thermostat conditions was done via the `fix_langevin` routine.¹⁹ The free particles were simulated in a box with periodic boundary conditions and were assigned initial velocities drawn from a uniform distribution for the given temperature. The viscous drag coefficient α was computed using Stoke's law for a spherical particle at low Reynolds number: $\alpha = 6\pi\eta R$ where $R = \sigma/2$ is the particle radius and η is the fluid viscosity.

Figure 1 shows the average root mean squared displacement of the particles computed as the average of 100 simulated trajectories per particle. Since there is no energy scale for the free particle, we used natural units and in this case set the diameter of the particles to $\sigma = 1\mu\text{m}$ and their mass density to that of water, yielding a mass of $m \sim 4.2 \cdot 10^{-15}\text{kg}$. We found that the statistical relative errors for the average trajectories were negligible for this number of simulations.

The crossover time was determined by inspection from the graphs. We can read off a crossing-over time of $\sim 0.5\mu\text{s}$ for particle #3 where the inertial response has fully died down; this is on the same order as the characteristic time scale $\tau_v = m/\alpha \sim 220\text{ns}$.

We simulated particles of different sizes ($1 - 20\mu\text{m}$) at different temperatures ($293 - 400\text{K}$). Here it may be necessary to account for the fluid viscosity variation with temperature, and we used the empirical formula for water²⁰:

$$\eta(T) = 2.141 \cdot 10^{-5} \cdot 10^{247.8/(T-140)}. \quad (2)$$

FIG. 1. **Inertial to diffusive crossover for the single particle.** Points are rms displacements computed from the simulated trajectories of 100 freely diffusing single particles for each set of parameters. Solid lines are the inertial and the diffusive limits of the Langevin equation solution (1). Particle #2 initially follows the trajectory of particle #3, with has the same mass, before crossing over to the trajectory of particle #1, which has the same drag coefficient. Errors were computed from the statistical distribution of particle displacements, but are too small to appear on this scale.

We have confirmed that the particle trajectories generated by LAMMPS had the statistical properties expected from theory. We were also able to confirm that the crossover time τ_v is practically independent on the heat bath temperature, since the viscosity only depends weakly on temperature in the range that we covered in our simulations.

To verify the fluctuation-dissipation theorem, which is used to derive the Einstein relation $D = k_B T/\alpha$, we computed the diffusion constant from a linear fit of the last three decades of each trajectory, i.e. for $t > 10^5\text{ns}$, obtained from 100 simulated particles. Figure 2 shows the product $D\alpha$ that was computed theoretically using the Stokes relation (see above) as a function of temperature, compared with the measured data. The predicted trend is observed, with a very small systematic offset that has been observed for a number of other integration schemes in Brownian dissipative dynamics.²¹ Essentially, this is an artefact that arises from the coarse-graining of the microscopic properties of the fluid using a random force ζ while imposing overall momentum conservation; this error is not significant for our purposes for two reasons: the algorithm produces trajectories in almost perfect agreement with the theory across a broad range of temperatures and furthermore, previous studies have shown that the effects due to integrator artefacts are only significant when the conservative forces of interest are comparable to the thermal fluctuations – all the potentials we apply will exceed energies of a few $k_B T$, so we expect no effect of this small factor.

FIG. 2. **Testing the fluctuation-dissipation theorem.** Diffusion constants D_{sim} were obtained from average trajectories and are plotted as points, multiplied by the drag coefficient $\alpha = 6\pi\eta\sigma$, for simulations at different temperatures. The different colours correspond to the types of particles, with different radii as indicated on the plot. The corresponding Einstein relations $D\alpha = k_B T$ are plotted as solid lines.

III. FREE PARTICLE IN A CHANNEL

Having established the dynamics of Brownian particles and verified that LAMMPS generates trajectories with the desired statistics, we now turn our attention to the diffusion of particles confined in a narrow channel. We considered particles of diameter $\sigma = 2R$ freely diffusing through a cylindrical channel of radius σ and length L .

Note that from here on, we will use Lennard-Jones (LJ) units which render all quantities dimensionless by assuming that particle interactions follow the standard Lennard-Jones potential $V(r) = 4\epsilon[(\sigma/r)^{12} - (\sigma/r)^6]$ and setting the particle mass m , the Boltzmann constant k_B and ϵ and σ as defined above equal to 1.²² For example, all lengths are from here on to be understood as multiples of the particle diameter σ , while other variables can be transformed to a dimensionless form by a scaling with an appropriate combination of m , k_B , σ and ϵ , e.g. for time: $t_{LJ} = t \cdot \sqrt{\epsilon m^{-1} \sigma^{-2}}$. For a full list of conversion formulae see the LAMMPS manual²². LJ units are widely used in computational physics and offer the advantage of treating systems of different size and energy scales in one framework.

In our simulations, particles are modelled as spheres with a Lennard-Jones 12/6 type repulsion and no attractive interaction tail, that is, the LJ potential of pair interaction is truncated at the point of its minimum, $r^* = 2^{1/6}\sigma$. The channel radius is too small to allow particles to overtake each other, thus producing the *single-file diffusion* and reflecting the experimental fact that many metabolites will completely block their channels during the transport due to their tight fit.²³

Figure 3(a) shows a 2D-projection of the simulation geometry (all simulations were carried out in full three

FIG. 3. **Channel geometry and an applied potential.**

(a) The simulation box is split in three regions, with the ‘channel’ of length L in the middle aligned along the z -axis, containing spherical particles. Particles are inserted in the blue region to the left and removed from the simulation once they have crossed the channel and entered the blue removal region to the right. (b) A typical applied potential (red), in this case a uniform well, and the corresponding force exerted on a particle (blue), to scale.

dimensions). The simulation box is a cylinder of radius σ , aligned along the z -axis, whose walls interact with particles using the repulsive part of the Lennard-Jones potential, same as described above, thereby ‘softly’ preventing contact. Particles are inserted at the very left end of the simulation box, the ‘insertion region’ (blue), if there is enough space. They then diffuse inside the cylinder. What we will call the ‘channel’ of length L is the area shown in white in the middle of the simulation box. Once the particles have crossed the channel and entered the ‘removal region’ to the right (blue), they are removed from the simulation. Underneath, in Fig. 3(b) is a plot of two example potentials $V(z)$ and their corresponding force landscapes, to scale. To investigate the first passage time distribution, no potential was applied.

Distribution of first passage times

To check the physics of our channel setup, we looked at a classical problem: the distribution of first passage times, $f(\tau)$, of particles freely diffusing through the channel, i.e. individually and without any applied potential. The first solution of this problem is due to Lord Kelvin, obtained by the methods of images¹⁴. However, the easy exponential solution it is only applicable when the particle is free to diffuse as far as necessary to the left of $z = 0$, while the first passage time is being tested by arriving at $z = L$ to the right of its entry point, see Fig. 3(a). In our case the passage is blocked to the left, so

to find the probability for a particle $p(z, t)$ one needs to solve the one-dimensional free diffusion equation with the boundary conditions: reflective wall, $\nabla p = 0$ at $z = 0$, absorbing wall, $p = 0$ at $z = L$, and the initial condition for insertion: $p(x, t = 0) = \delta(z)$. The explicit solution is:

$$p(z, t) \propto \sum_{n=0}^{\infty} \cos \left[\frac{\pi z}{L} (n + 1/2) \right] \exp \left(-\frac{\pi D^2 (n + 1/2)^2}{L^2} t \right). \quad (3)$$

The survival probability for the particle to remain anywhere between 0 and L , having started at $z = 0$ is obtained by integration: $Q(t) = \int_0^L p(x, t) dx$. Given the boundary conditions, $Q(t)$ does not depend on anything happening outside the $(0 - L)$ interval. Given the definition of the survival probability, the fraction of particles equal to $-dQ(t)/dt$ is absorbed between t and $t + dt$. This means that $f(t) = -dQ(t)/dt$ is actually the probability density of the time t that takes the particle to reach $z = L$ for the first time. This distribution function is plotted in Fig.4, and it gives *average* first passage time $\tau_{\text{diff}} \approx 4.92L^2/\pi^2 D$. A solution based on the inversion of a Laplace transform^{24,25} gives the average first passage time $\tau_{\text{diff}} = L^2/2D$, numerically indistinguishable from the mode-sum version above.

We measured the passage times in 500 simulations of a single particle diffusion (to ensure no pair-interaction events could take place), where we inserted a particle in the insertion area and allowed it to freely diffuse to the end of the channel, the time for which we measured. We fit the distribution of first passage times to the resulting histogram of particle travel times in Fig.4, where only the diffusion constant D and a normalisation act as fitting parameters. The data are in good agreement with the model, and we find a bare diffusion constant $D_0 = 0.28$.

Interestingly, we also found that when we considered a similar experiment where we inserted several particles into the channel one by one at a certain (very low) frequency, the distribution of first passage times severely deviated from the free-diffusion result even at concentrations of just 2-3 particles inside a channel of length $L = 30$ at any one time. This shows that many-particle effects caused by particle-particle interactions cannot realistically be ignored even at the lowest of concentrations, a point to which we will return at the end of this paper. In this particular case, the average first passage time was significantly increased at these concentrations, suggesting a smaller diffusion constant or higher effective resistance.

IV. POTENTIAL ALONG THE CHANNEL

Having established that our simulation setup produces physically meaningful results, we now turn to the dependence of the flux through a passive channel on the potential landscape inside it. We therefore made a series of experiments, in each of which we simulated the insertion of 100 particles in the channel. Since this is now a genuine multi-particle problem, simulation time increases

FIG. 4. **Distribution of first passage times for free diffusion.** The histogram was obtained from the first passage time of 500 single-particle simulations. The solid line is the theoretically calculated distribution $f(\tau)$, fitted using the diffusion constant $D = 0.284$ and a normalisation constant as the fitting parameters.

accordingly. We therefore made use of the parallel computing capacities of LAMMPS.

Particle insertion was attempted at a rate $0.01\tau^{-1} = 1/T_{\text{in}}$ where τ is time in LJ units (see early section III). Particles were only inserted if there was enough space in the insertion region; if a particle could not be inserted due to crowding at the channel entry, the insertion was skipped and the next attempt was made after a time interval of T_{in} . The intended insertion interval T_{in} is significantly smaller than the mean first passage time $\tau_{\text{diff}} \approx L^2/2D \approx 1600$, leading to a system driven far from equilibrium. Since we have analysed the effects of different insertion rates earlier¹⁴, we will keep the high insertion rate fixed for the remainder of the paper and focus our attention on the effects of the potentials. Inside the channel, one out of a number of different potential shapes was applied with potential depth between $V_{\text{min}} \in [5, 70]$ in LJ units. The shapes of the potential are shown in the insets of Fig. 6 and include ‘continuous’ potentials, where the channel is modelled having a homogeneous attractive interaction with the particle along its entire length (single/double tanh, triangular potentials) as well as ‘discrete’ potentials, where the channel provides a number of discrete, spatially well-defined binding pockets, modelled as a Gaussian with a depth V_{min} and a standard deviation of 0.5σ , where σ is still the particle diameter as introduced in section III. Note that potential ‘steps’ are modelled using tanh functions, hence the names ‘Double tanh’ etc.

Despite the binding pockets being narrow, we can clearly see particle trapping occurring by looking at individual particle trajectories such as the one shown in Fig. 5, where displacement of a single particle along the channel is plotted on the x-axis in red, with time on the

FIG. 5. **Example trajectory of a single particle in a channel with four binding pockets.** Time is plotted on the y-axis and displacement along the channel in red on the x-axis. The potential inside the channel (blue) has four binding pockets, and the trajectory clearly displays trapping of the particle in every pocket, spending most time in the second pocket. The overall flux, however, is unaltered compared to a channel with no potential at all (see Figs. 6 and 7).

y-axis. The applied potential, a series of four spatially discrete binding pockets, each modelled as a Gaussians, is plotted in blue. We can clearly see that the particle is trapped by every binding pocket, spending most time in the second pocket from the left. However, the continuous insertion of particles to the left of the channel leads to a consistent movement of the tagged particle to the right.

We measured the cumulative number of particles that have crossed the channel as a function of time. An example of these measurements is shown in Fig. 6, where the cumulative number of translocated particles is plotted as a function of time for different potential shapes, all with the same $V_{\min} = -20$. Insets show the exact form of the applied potentials for each experiment.

We can already make a number of observations from Fig. 6. First of all, a simple potential drop at the entrance of the channel leads to the expected acceleration and hence significantly increased flux (red). Symmetrical potentials produce an increased flux compared to a channel with no potential ($V = 0$) or different numbers of discrete binding pockets, which surprisingly do not change the flux significantly compared to the $V = 0$ case. The number of translocated particles saturates for all potentials with $V_{\min} = -20$ at a value lower than 100, the total number of particles inserted during the simulation, because as particle insertion stops, exiting the channel becomes increasingly harder for the remaining particles since the pressure inside the channel decreases. The fact that the translocation number for a triangular potential saturates at a higher value than for the double-tanh potential supports this interpretation, since the double-tanh potential has a steeper wall at the end of the channel

FIG. 6. **Cumulated number of translocated particles for different potential shapes with $V_{\min} = -20$.** The insets show the applied potential in the colour of the translocation curve that it produced, e.g. the ‘Double tanh’ potential, plotted in blue, was applied along the channel in a simulation that yielded the blue translocation curve. Discrete binding pockets don’t change the flux compared to the free ($V = 0$) channel, plotted in pink. Symmetric potentials (blue, green) enhance the flux, but as we’d expect not as much as a simple potential drop at the beginning of the channel (red).

which will effectively block the channel. The number of particles trapped in the channel at the end of the simulation increases as the potential depth increases: for the double tanh potential, particles are trapped for a time $t \gg T$, the time of the simulation, at a potential depth of $V_{\min} \sim -10$, while for the other potentials trapping only occurs at potential depths $V_{\min} > -15$.

Let us now discuss how the flux through the channel depends on the depth of the potential for the different potential functions discussed so far. We therefore define flux as the slope of a fit to the linear portion of the translocation plots in Fig. 6:

$$J = \left\langle \frac{dN}{dt} \right\rangle, \quad (4)$$

and repeat the analysis above for different potential depths. The average flux was computed from five experiments for given potential shape and depth, that simulated the translocation of 100 particles each. Simulating this number of particles stabilised the linear fits and resulted in the small relative errors that are plotted in Fig. 7 and enable us to refine some of the observations already made:

1. Symmetric potentials increase the flux of single-file diffusion. This is surprising at first sight, since the overall work done on the particle is zero. However, the symmetry of the potentials is broken by the pressure that the newly inserted particles exert on the particles near the end of the channel at the potential wall.¹⁴ It should be noted that this pressure emerges purely from the free diffusion inside the channel and has significant effects even at low

FIG. 7. **Normalised flux as a function of potential depth for different profiles.** Fluxes are defined as the slopes of a linear fit to the cumulative plot of the number of translocated particles as a function of time (e.g. Fig. 6). Fluxes were measured from five experiments per potential shape/depth that simulated the translocation of 100 particles each and were normalised to the flux through a channel with $V = 0$. The dashed lines for the three characteristic types of behaviour are to guide an eye.

colloid concentrations inside the channel. We are only inserting particles if there is free space at the beginning of the channel, as described above, so we are not actively pushing particles through the channel.

Furthermore, the flux through channels with symmetric potentials does not go below the flux through a free channel even for deep potentials.

2. The triangular potential profile outperforms the double-tanh potential. This is due to the fact that in the overdamped limit, after an impulse is delivered to a particle, it very quickly relaxes back to normal diffusion. Effectively, Newton's second law does not hold anymore and a small force over a longer time, pushing the dense region forward at the entry half of the channel, is more effective than a strong force over a short period of time.

3. We find that there are optimal potential depths that maximize flux ($V_{\min} \geq 30$ for the triangular potential, $V_{\min} \sim 15$ for the double tanh potential). This observation is in agreement with theoretical predictions for diffusion of individual particles along a channel.^{26,27}

4. The increase in flux with symmetric potentials is not due to some sort of Kramers-type barrier hopping. This is shown by the fact that the flux through a channel with a tanh step at its end ('tanh wall') (see inset in Fig. 7) goes to zero for $V_{\min} \sim 25$, where double-tanh and triangular potentials still outperform the $V = 0$ case.

5. Narrow binding pockets do not alter the flux significantly, even though it is clear from individual particle trajectories that particles do get trapped in the binding pockets (Fig. 5). This is in contrast to theoretical results that were obtained for individual, non-interacting particles¹⁰ and highlights the importance of many-body

FIG. 8. **Binding potentials keep particles in the channel.** The snapshots (a), (b) and (c) for different potential profiles give particle positions at the end of a simulation run, with channel and particle diameters to scale. This illustrates that sufficiently deep attractive potential will retain some particles, in regions shaded in the plots, when no additional influx from the left occurs (explaining the saturation plateau below 100 in Fig. 6).

interactions in the crowded channel: here, the pressure emerging from the free diffusion inside the channel and removal of particles at its end pushes bound particles out of the pockets and hence establishes a constant flux.

Figure 8 gives the snapshots of final simulation frames to illustrate what is an 'equilibrium' situation in each potential profile $V(z)$. It shows that for a sufficiently deep attractive potential well, particles are retained in such a well, while the particles facing weaker binding forces escape and diffuse out of the channel. The final number of retained particles explains why the plateaus of different curves in Fig. 6 are below 100. These snapshots also help understand why the flux increases with the depth of continuous potentials (double-tanh or triangular). The process is analogous to the enzymatic action: although the energy barrier at the end of channel is prohibitively high (as illustrated by the complete vanishing of diffusive flux for the 'tanh-wall' potential in Fig. 7), when particles are confined at a high density in front of such a wall – they are forced to escape, pushed by the neighbours

from the left.

It is also interesting to observe that at a constant temperature of our heat bath, when these potentials become excessively deep, the channel does get blocked: this occurs at $V_{\min} < 50 - 60$ for the double-tanh potential, and has to be inferred to occur at a much greater depth for the triangular potential, see Fig. 7.

V. THEORETICAL CONSIDERATIONS

We would like to gain a better insight into the data shown in Fig. 7 from a theoretical perspective. Since the basic features of single-file diffusion have already been discussed extensively elsewhere,^{28–30} we will focus our discussion here on the relative change in flux through a channel when we apply a potential.

It turns out that an effective way to pose this problem is to describe translocation as a reaction $A + B \rightarrow B$ where the colloidal particles A are absorbed by a “trap” B , i.e. the channel exit, upon encounter, although it should be noted that other approaches have been applied successfully³¹. The problem of finding the flux through the channel becomes the problem of computing the rate κ of this reaction in a crowded single-file environment with applied potentials. For systems with spherical symmetry in the limit of infinitely diluted reactants A , this is a classical problem of diffusion-controlled reaction kinetics, which was solved exactly by Smoluchowski,³² producing the rate $\kappa_s = 2\pi D_0 \sigma \rho_0$ where $\rho(r)$ is the density profile of reactants A around the trap reaching the value ρ_0 at infinity.

In general, the reaction dynamics is governed by the diffusive Fokker-Planck equation:

$$\frac{\partial \rho}{\partial t} = D_c \nabla \cdot \left(\nabla \rho - \frac{F}{k_B T} \rho \right), \quad (5)$$

taking the many-body effects into account through the inhomogeneous density profile $\rho(r)$ along the channel, which generates an osmotic pressure $\Pi(r)$ that acts as to spread the density profile via a force per particle $F(\rho(r)) = -1/\rho(r) \cdot \nabla \Pi(\rho)$.³³ These two parameters are non-linearly coupled via the collective diffusion coefficient $D_c = D_0 \cdot \partial \Pi / \partial \rho$. An exact solution of this problem is a formidable task even for numerical methods, which makes us look for reasonable approximations. We assume first that the density profile $\rho(z)$ is computed from the simulations. Secondly, we reformulate the problem as a *homogeneous* collective diffusion problem in the superimposed field $F(\rho(r))$ just introduced, i.e. we ignore the ρ -dependence of D_c when we take the derivative in (5). This approximation works since the dependence of D_c on ρ is less non-linear than the dependence of Π which survives the approximation and dominates the final result.³⁴

We take $\rho(z)$ as the number of colloid particles per unit length along the channel at a given time (hence always $\rho < 1$), and we initially ignore the force due to the applied potential. With increasing ρ , it takes a given single

FIG. 9. **Binding potentials keep steady-state density highly non-linear.** The snapshots (a), (b) and (c) for different potential profiles give the average density of particles in each case, in the steady-state transport regime (constant flux).

particle longer to reach the channel exit, but once it is in the vicinity of the exit, its chance of escaping increases since it cannot diffuse far away from the exit. Quantitatively, Dorsaz and co-workers¹¹ showed numerically that the reaction rate can be approximated well by the following expression:

$$\kappa \approx \kappa_s \frac{\beta \Pi(\rho_0)}{\rho_0} \cdot \exp \left(\frac{-\beta \Pi(\rho_\Delta)}{\rho_\Delta} \right) \quad (6)$$

where $\beta = 1/k_B T$, ρ_0 is the density at the beginning of the channel, and $\rho_\Delta = \rho(\Delta)$ is the density a characteristic ‘encounter distance’ Δ from the channel end at which the density of colloid particles acquires structure due to interactions (in other words, where the ideal-gas linear relationship $\Pi = k_B T \rho$ stops being valid), see Fig. 9. Equation (6) has since been derived from first principles by Zaccone,³⁴ who finds a prefactor of $\beta(d\Pi/d\rho)_{\rho_0}$ instead of $\beta \Pi(\rho_0)/\rho_0$, but notes that the two prefactors have the same dependence on ρ_0 which would indicate that the solution is qualitatively correct.

FIG. 10. **Reaction rates (6) for an absorption reaction $A+B \rightarrow B$ correctly predict the flux through a channel with applied potential.** The trends seen in the simulations (Fig. 7) are correctly predicted and numerical agreement is also good, although there is a systematic offset of ~ 0.1 . Since the form of the potentials does not enter the model at any point, we conclude that all the information is encoded in the equilibrium density distribution, which we sample at only two discrete points.

In order to evaluate (6), we write the osmotic pressure $\beta\Pi$ as a virial expansion in the density along the channel:

$$\beta\Pi = \rho + B_2\rho^2 + B_3\rho^3 + \dots \quad (7)$$

and compute ρ from 1000 randomly selected snapshots of the simulations after the flux has equilibrated to its steady-state value. The virial coefficients B_2 and B_3 account for two- and three-body interactions between the particles in the channel which captures the essential dynamics since in the effectively 1-D system of the channel, the motion of a particle is dependent on the particle in front and the particle behind it.¹⁴ B_2 and B_3 were computed for the Lennard-Jones 12/6 potential with $\epsilon = 1, \sigma = 1$ that was used to model particle-particle interactions as described in section III. While $B_2 = 2\pi \int_0^\infty r^2 [1 - e^{-\beta V(r)}] dr$; computation of B_3 is more involved, but values are available.³⁵

A plot of the reaction rates computed from (6) is shown in Fig. 10. All values are normalised with respect to the reaction rate computed for no potential: $\kappa \equiv 1$ at $V = 0$. It is clear from the graph that these rates correctly predict the trends seen in the flux from the simulations (Fig. 7): there is no significant flux change with discrete pockets but a considerable increase with continuous potentials; the double tanh potential performs best at small potential depths while the triangular potential trumps for deeper potentials. The numerical range of the relative changes is good although it is systemically low by ~ 0.1 . This is a remarkable agreement given that at no point we explicitly introduced the form of the potentials

and evaluate ρ only at two discrete points, *i.e.* the beginning of the channel and very close to its exit. This shows that all the information about many-particle effects and the channel translocation with an applied potential is encoded in the steady-state density distribution, which in turn is controlled by the two virial coefficients B_2 and B_3 (or the two values ρ_0 and ρ_Δ sampled near the beginning and near the end of the channel).

VI. CONCLUSION

Our result that continuous, symmetrical potentials increase the flux significantly confirms earlier speculation that membrane channels in cells are most likely to provide a “molecular slide”⁶ by organising discrete binding sites in succession, since having them isolated one after the other would provide little to no increase in flux as shown. Furthermore, our results can offer guidance for the design of artificial channels in microfluidic applications, where improving flux is often important and clogging can be a problem¹⁴.

Any theoretical description of particle translocation has to account for both the applied potential and the crowding inside the channel. We have shown that it is possible to account for the relative changes in flux by considering the kinetics of the “absorption reaction” of particles exiting the channel, thus mapping the many-body problem to a two-body-interaction where crowding is modelled by the osmotic pressure inside the channel, without knowledge of the applied potential. However, this is more of an explanation *a posteriori* since it requires knowledge of the density profile along the channel. Further theoretical work will therefore have to focus on the development of methods to calculate these distributions not just for periodic boundary conditions²⁹, but for more realistic geometries and boundary conditions in the presence of potentials in an attempt to predict particle flux without resorting to simulations.

ACKNOWLEDGEMENTS

We are grateful to Anna Lappala for many stimulating discussions. Simulations were funded by the Cavendish Laboratory teaching committee and performed using the Darwin Supercomputer of the University of Cambridge High Performance Computing Service (<http://www.hpc.cam.ac.uk/>), provided by Dell Inc. using Strategic Research Infrastructure Funding from the Higher Education Funding Council for England.

¹B. Alberts et al., *Molecular Biology of the Cell*, Garland Science, New York, 2008.

²R. Shah et al., *Mater. Today* **11**, 18 (2008).

³S. Pagliara, C. Schwall, and U. F. Keyser, *Adv. Mater.* **25**, 844 (2013).

⁴K. Sugano et al., *Nat. Rev. Drug Discov.* **9**, 597 (2010).

⁵B. Hille, *Ion channels of excitable membranes*, Sinauer Sunderland, MA, 2001.

- ⁶T. Schirmer, T. Keller, Y. Wang, and J. Rosenbusch, *Science* **267**, 512 (1995).
- ⁷J. J. Kasianowicz, T. L. Nguyen, and V. M. Stanford, *Proc. Natl. Acad. Sci.* **103**, 11431 (2006).
- ⁸C. Hilty and M. Winterhalter, *Phys. Rev. Lett.* **86**, 5624 (2001).
- ⁹M. O. Jensen, S. Park, E. Tajkhorshid, and K. Schulten, *Proc. Natl. Acad. Sci.* **99**, 6731 (2002).
- ¹⁰A. Kolomeisky, *Phys. Rev. Lett.* **98**, 048105 (2007).
- ¹¹N. Dorsaz, C. De Michele, F. Piazza, P. De Los Rios, and G. Foffi, *Phys. Rev. Lett.* **105**, 120601 (2010).
- ¹²W. R. Bauer and W. Nadler, *Proc. Natl. Acad. Sci. U. S. A.* **103**, 11446 (2006).
- ¹³P. Bressloff and J. Newby, *Rev. Mod. Phys.* **85**, 135 (2013).
- ¹⁴A. Lappala, A. Zacccone, and E. M. Terentjev, *Sci. Rep.* **3**, 3103 (2013).
- ¹⁵J. C. Chen and A. S. Kim, *Adv. Colloid Interface Sci.* **112**, 159 (2004).
- ¹⁶S. J. Plimpton, *J. Comput. Phys.* **117**, 1 (1995).
- ¹⁷A. Zilman and G. Bel, *J. Phys. Condens. Matter* **22**, 454130 (2010).
- ¹⁸G. Uhlenbeck and L. Ornstein, *Phys. Rev.* **36**, 823 (1930).
- ¹⁹T. Schneider and E. Stoll, *Phys. Rev. B* **17**, 1302 (1978).
- ²⁰T. Al-Shemmeri, *Engineering Fluid Mechanics*, Ventus Publishing ApS, 2012.
- ²¹I. Vattulainen, M. Karttunen, G. Besold, and J. M. Polson, *J. Chem. Phys.* **116**, 3967 (2002).
- ²²The LAMMPS Team, Lammmps documentation, <http://lammps.sandia.gov/doc/units.html>, 2014.
- ²³E. M. Nestorovich, C. Danelon, M. Winterhalter, and S. M. Bezrukov, *Proc. Natl. Acad. Sci.* **99**, 9789 (2002).
- ²⁴P. G. Harrison, *J. Appl. Prob.* **27**, 74 (1990).
- ²⁵S. M. Bezrukov, A. M. Berezhkovskii, M. A. Pustovoi, and A. Szabo, *J. Chem. Phys.* **113**, 8206 (2000).
- ²⁶A. M. Berezhkovskii, M. A. Pustovoi, and S. M. Bezrukov, *J. Chem. Phys.* **116**, 9952 (2002).
- ²⁷A. M. Berezhkovskii and S. M. Bezrukov, *Biophys. J.* **88**, L17 (2005).
- ²⁸D. Levitt, *Phys. Rev. A* **8**, 3050 (1973).
- ²⁹M. Kollmann, *Phys. Rev. Lett.* **90**, 180602 (2003).
- ³⁰P. S. Burada, P. Hänggi, F. Marchesoni, G. Schmid, and P. Talkner, *Chemphyschem* **10**, 45 (2009).
- ³¹A. Zilman, J. Pearson, and G. Bel, *Phys. Rev. Lett.* **103**, 128103 (2009).
- ³²M. Smoluchowski, *Z. phys. Chem.* **92**, 129 (1917).
- ³³A. Zacccone and E. M. Terentjev, *Phys. Rev. E* **85**, 061202 (2012).
- ³⁴A. Zacccone, *J. Chem. Phys.* **138**, 186101 (2013).
- ³⁵R. B. Bird, E. L. Spotz, and J. O. Hirschfelder, *J. Chem. Phys.* **18**, 1395 (1950).