

OF TIME AND TAPHONOMY: PRESERVATION IN THE EDIACARAN

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ABSTRACT.—The late Neoproterozoic witnessed a revolution in the history of life: the transition from a microbial world to the one we know today. The enigmatic organisms of the Ediacaran hold the key to understanding the early evolution of metazoans and their ecology, and thus the basis of Phanerozoic life. Crucial to interpreting the information they divulge is a thorough understanding of their taphonomy: of what is preserved and how it is preserved, and also of what is not preserved. Fortunately, this Period is also recognized for its abundance of soft-tissue preservation, which is viewed through a wide variety of taphonomic windows. Some of these, such as pyritization and carbonaceous compression, are also present throughout the Phanerozoic, but the abundance and variety of moldic preservation of body fossils in siliclastic settings is unique to the Ediacaran. In rare cases, one organism is preserved in several preservational styles which, in conjunction with our increased understanding of the taphonomic processes involved in each style, allow us to more confidently interpret aspects of the biology and ecology of the organisms preserved. Several groundbreaking advances in this field have been made since the 1990s, and have paved the way for increasingly thorough analyses and elegant interpretations.

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INTRODUCTION: THE IMPORTANCE OF TAPHONOMY

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Taphonomic analyses can provide valuable insights into a spectrum of paleobiological questions, including biology, tissue composition, and paleoecology, as well aspects of the paleoenvironment such as sediment pore-water composition and ocean water geochemistry. In celebrated examples, unique taphonomic windows and exceptional preservation have allowed fundamental questions about a group to be resolved. Ediacaran macrofossils are amongst the least-well understood of any macrobiotic assemblage in terms of their biology, paleoecology, and phylogenetic affinity. A thorough understanding of the processes involved in their preservation is required to distinguish between taphonomic artifact and genuine morphological features. Accordingly, there is a long history of actualistic taphonomic analyses undertaken on this biota, combining field-based observations with petrographic and experimental data. Furthermore, recent research has demonstrated that the coincidence of multiple taphonomic windows operating on the same organisms allow us to resolve fundamental questions regarding the taphonomic processes themselves; each window provides a test-bed against which the others can be compared. Of all Ediacaran macrofossil assemblages, those traditionally referred to as the ‘Ediacara biota’ (e.g. Gehling, 1999; Narbonne, 2005) have been the most intensively scrutinised, and thus form the focus of this discussion.

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The most notable difference between the Ediacaran and the Phanerozoic is the abundance of moldic preservation of soft parts in sand-grade sediments, but the

differences in geochemistry and taphonomic processes between the two are not yet fully understood. There remain fundamental questions regarding life and its preservation in the Ediacaran, including the composition of the soft parts of the organisms, the taphonomic biases imparted by the various paleoenvironments in which the organisms lived, and the relationship between organism, microbial mat, and sediment. Only by understanding the taphonomy of these organisms can we begin to confidently elucidate other aspects of their biology. Herein, we review Ediacaran diversity and paleobiology, discussing the dominant preservational styles, the importance of microbial mats, and a few intriguing oddities. We explore the limitations to our current knowledge, and suggest future directions for research in this fascinating Period.

Ediacaran macrofossils – it's life, but not as we know it!

The Ediacaran Period (635-541 Ma; Knoll et al., 2004) hosts the earliest macrofossils currently known from the rock record. These include the ~600 Ma Lantian biota (Yuan et al., 2011), and the much more widely-studied assemblages commonly and collectively referred to as the Ediacara biota (e.g. Gehling, 1999; Narbonne, 2005; see MacGabhann, 2014 for a discussion). Assemblages included in the latter biota span some 40 Ma of Earth history, from immediately after the Gaskiers glaciation to the base of the Cambrian (Fedonkin et al., 2007; Narbonne et al., 2012), and shape our ideas of early metazoan evolution and the development of Phanerozoic ecosystem structure. They also provide a means of calibrating molecular clocks and the origination of metazoan clades (Erwin et al., 2011). They are thought by many to include some of the earliest metazoans (e.g. Narbonne, 2005), but interpretations regarding their phylogenetic affinity are as

varied as the organisms themselves. They have been interpreted as, or allied to, stem- or crown-group metazoans (e.g. Glaessner, 1979; Clapham et al., 2003; Sperling and Vinther, 2010; Sperling et al., 2011), algae (Ford, 1958), xenophyophores (large benthic foraminifera; Seilacher et al., 2003), fungal-grade organisms (Peterson et al., 2003), an extinct Kingdom (the Vendobionta; Seilacher, 1984, 1992) and even lichens (Retallack, 1994). Part of this confusion is, at least in part, attributable to the historic treatment of the biota as one phylogenetic group, when it almost certainly includes representatives from many disparate clades (Xiao and Laflamme, 2009; Erwin et al., 2011). Treatment of the organisms on a ‘case by case basis’ will, no doubt, lead to a clearer understanding of the diversity and biology of the organisms present.

Most workers now recognize several distinct groups (Fedonkin et al., 2007; Laflamme et al., 2013), including: 1) rangeomorphs, characterised by repeated branching which creates a pseudo-fractal, “modular” architecture (Narbonne, 2004; Brasier et al., 2012); 2) arboreomorphs, which are similar in gross appearance to rangeomorphs but have a different (non-fractal) branching pattern; 3) kimberellomorphs, which are bilaterally symmetrical, display clearly defined anterior-posterior differentiation and have at least three concentric zones; 4) erniettomorphs, which are modular and consist of tubular units, and 5) dickinsoniomorphs, which are also modular but show anterior-posterior differentiation. Four additional groups are defined based purely on their symmetry: bilateralomorphs (which are likely polyphyletic; Laflamme et al., 2013), triradialomorphs, tetradialomorphs, and pentaradialomorphs (Laflamme et al., 2013). Although considered phylogenetically distinct, individuals of all described groups comprise serially-repeated units.

Three assemblages have been proposed: the Avalon, White Sea and Nama assemblages (Waggoner, 2003). Whilst these correlate broadly with current understanding of paleogeography and age, debate as to the extent of the influence of depositional environment on biotic composition of any given site persists (Grazhdankin, 2004; Droser et al., 2006; Gehling and Droser, 2013). Understanding the paleoenvironment of a site, and thus the biases imposed on taphonomic processes, is of critical importance in assessing the fidelity of the fossil assemblage to the life assemblage (e.g. Grazhdankin et al., 2008).

Many sites record diverse subaqueous communities (e.g. Xiao et al., 2013), comprising upright and flat-lying forms, and some thought to be wholly or partially buried in the sediment; the majority were benthic, sessile, and epifaunal (Narbonne, 2005; Laflamme and Narbonne, 2008). Whilst scant evidence for motility has been documented from the oldest of these sites (Liu et al., 2010), it is not until the latest Ediacaran that bioturbation (e.g. Chen et al., 2013; Meyer et al., 2014c) and macro-benthic predation (e.g. Hua et al., 2003) become obvious. Microbial mats are widespread and well-documented from these assemblages, and have been implicated in their paleoecology (e.g. Seilacher, 1999).

Peeking through the taphonomic windows at a soft-bodied world

The Ediacaran is conspicuous for its sheer abundance of soft-tissue preservation. Dozens of sites are known from across five continents, and include representatives from multiple paleoenvironmental settings hosting abundant and diverse communities. They allow a glimpse onto a world populated almost entirely by soft-bodied organisms.

Without such abundant preservation of these soft parts, we would have little knowledge of macro-benthic life during most of this critical interval of Earth history; mineralization in macro-organisms only evolves in the terminal Ediacaran (e.g. Grant, 1990; Grotzinger et al., 2000; Penny et al., 2014), and the makers of known trace fossils largely remain elusive.

Many aspects of the Ediacaran paleoenvironment differed to the Phanerozoic, and different taphonomic biases held sway: sediment surfaces were sealed by microbial mats, scavenging and deep bioturbation were absent, and the organisms were largely sessile and immotile. Consequently, there was a lack of disturbance and an attendant lack of significant time-averaging. Microbes are often cited as key to preservation (e.g. Briggs, 2003; Raff and Raff, this volume), and they formed ubiquitous mats in the Ediacaran (see *Binding it all together*, below). It also had a potentially very different sedimentary and oceanic chemistry, with lower seawater sulfate concentrations (Canfield et al., 2008), abundant labile dissolved organic carbon in the deep oceans (Sperling et al., 2011), and a condensed sediment-water geochemical profile, favoring early diagenetic mineralization (Callow and Brasier, 2009b).

The multiple views provided onto the community by the different taphonomic windows, coupled with the lack of significant time-averaging, give us greater confidence in the relative completeness of ecosystems. This has allowed inferences about community successions (Clapham et al., 2003), evolutionary progressions (Xiao and Laflamme, 2009) and ecological interactions to be made (Clapham and Narbonne, 2002; Clapham et al., 2003; Droser et al., 2006; Darroch et al., 2013). Nevertheless, details of the anatomy of many Ediacaran organisms are largely unknown. Only external and, rarely, internal

surfaces are preserved (Meyer et al., 2014a, 2014b); in marked contrast to the Phanerozoic, convincing evidence of preserved internal structures is virtually absent (though see Dzik, 2002, 2003; Narbonne, 2004).

VARIETY IS THE SPICE OF LIFE

The taphonomic windows through which we can view the biota are many and varied, each giving us a different perspective on their biology. Crucially, some organisms are preserved in more than one taphonomic mode, allowing the biases induced by each mode to be ascertained (e.g. Grazhdankin et al., 2008). Ediacaran preservation is dominated by three major taphonomic modes: 1) moldic; 2) replication by early diagenetic minerals; and 3) carbonaceous compression, each of which are detailed below. Two or more modes sometimes combine in an individual specimen (Fig. 1; Cai et al., 2012). The mode in which a fossil is preserved depends on a variety of factors, many of which are still incompletely understood, but which include the nature of the burial sediment and depositional environment (Narbonne, 2005), the nature of the microbial community (Gehling, 1999; Gehling et al., 2005), and the chemistry of the pore waters (Mapstone and McIlroy, 2006; Callow and Brasier, 2009b).

Upsides and downsides: moldic preservation

Moldic preservation is the most abundant and typical preservational style of the Ediacaran (Fig. 2–4; e.g. Gehling, 1999; Steiner and Reitner, 2001; Narbonne, 2005; Grazhdankin et al., 2008; Cai et al., 2012) but, barring a handful of exceptional examples

(e.g. MacGabhann et al., 2007), is largely unknown outside of this Period. Terms used to describe the nature of the molds, and their relationship to the beds preserving the fossils, were introduced by Glaessner and Wade (1966). Features observed on the top surface of a bed are termed epirelief (Fig. 2), and those seen on the base of a bed are hyporelief (Fig. 3). Features which form hollows or depressions have negative relief (Fig. 2a,c,d,f; Fig. 3a,b,c) and those which protrude above the substrate surface have positive relief (Fig. 2b,e; 3e). The sense of relief is thought to involve an interplay between the relative resistance of the soft parts to collapse, and the timing of substrate lithification (Gehling, 1999; Narbonne, 2005): more robust or recalcitrant parts collapse or decay more slowly, and so are cast by still-soft material from the underlying bed being injected upwards (creating negative hyporelief; Fig. 3a,b,c/positive epirelief impressions; Fig. 2b,e), whilst more fragile, fluid-filled or labile parts collapse or decay quickly, creating impressions which are filled and presumably cast by material from the overlying bed subsiding into the void (resulting in positive hyporelief; Fig. 3e/negative epirelief impressions; Fig. 2a,c,d,e; Fig. 4). The latter process likely requires stabilization of the lower surface of the organism prior to complete decay in order to retain the observed level of morphological detail (Darroch et al., 2012). Both senses of relief may be seen even within a single specimen, and in different specimens of the same taxon from different localities (Fig. 2b–e). In the Avalon Assemblage sites of Newfoundland and Charnwood Forest, the fossils are only seen preserved as epirelief impressions (Fig. 2; Fig. 4); the counterparts are as yet unknown.

The influence of the nature of the surrounding sediment and the depositional environment on the taphonomy of an organism is apparent from the four styles of moldic

preservation proposed by Narbonne (2005), but is not yet completely understood. He considers “Conception-style” preservation to be attributable to early diagenesis of reactive minerals in a smothering volcanic ash. In contrast, the classic White Sea Assemblage sites of Australia preserve their fossils on the bases of event beds (hyporelief), for which Narbonne (2005) coined the term “Flinders-style” (Fig. 3c,e). This style is also observed in the sandstone-and-shale facies of the White Sea (Grazhdankin, 2004), and has been suggested to be characteristic of shallow-marine environments between fair- and storm-wave base. In both regions, fossils may in whole or in part be preserved as positive or negative relief features. In his “Fermeuse-style preservation”, only the bases of holdfasts, and trace fossils, are preserved (Narbonne, 2005). In “Nama-style preservation” (Narbonne, 2005), named after its occurrence in the Nama Group of Namibia, three-dimensional internal and external molds of fossils (Fig. 3d,f,g) are preserved within storm event beds and channel-fill deposits deposited above fair-weather wave base. Whilst it has also been reported from the Spaniard’s Bay locality of Newfoundland (Fig. 4; Narbonne, 2004), other authors have interpreted the taphonomic mode there to be more akin to death mask preservation, with the higher relief of the fossils attributable to their preservation within scours in the underlying bed (Brasier et al., 2013). Regardless, the exquisite preservation on this bed is remarkable, and is dependent on a so-far unique combination of taphonomic artifice. This likely includes the burial of the frond within sediment (Brasier et al., 2013), which exposed the entirety of the frond to mineralizing pore waters, and unusually rapid and thorough mineralization, which was potentially a function of pore water chemistry with favorable iron and sulfur ion concentrations (see discussion of death mask preservation below).

208 Several taphonomic experiments have sought to replicate moldic preservation of
209 soft parts, but these have typically used specimens from only one species (Darroch et al.,
210 2012) or one phylum (Seilacher, 1984; Norris, 1989; Bruton, 1991). This limits their
211 usefulness in interpreting the soft-part composition or phylogentic affinity of the fossils,
212 as limited comparison between tissue types or phyla can be made (under the same set of
213 experimental parameters). Indeed, the different responses and preservation potential of
214 the cnidarian taxa studied by Norris (1989) hints at the importance of the structure and
215 composition of an organism on its taphonomy. Under favorable conditions, medusoids
216 and chondrophorans produce simple concentric or radial impressions; pennatulids appear
217 more susceptible to contraction even under burial and compression, with the stalk and
218 polyps producing clear impressions, but most branches retracting into an indistinct mass.
219 Interestingly, no impression of musculature was produced in these experiments (Norris,
220 1989); its preservation in the Phanerozoic typically involves phosphatization (Briggs,
221 2003 and refs. therein), which seems to play only a minor role in the Ediacaran outside of
222 Doushantuo-type preservation (Schiffbauer et al., this volume). Is the absence of
223 musculature in the Ediacaran original, or due to failure of preservation? Additionally,
224 only the influence of the content of organic matter in the sediment was investigated
225 (Norris, 1989; Darroch et al., 2012); other parameters (e.g. grain size, composition) exert
226 an as yet unquantified influence. These experiments have also been conducted under an
227 array of environmental conditions, from water-covered (Bruton, 1991; Darroch et al.,
228 2012) to water-saturated with compression (Norris, 1989) to dry (beach strandline;
229 Bruton, 1991). Interestingly, experiments conducted under water but without mats or
230 compression result in poor or absent impressions, as the buoyancy of the decaying

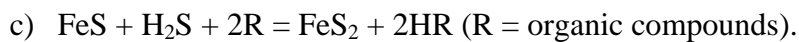
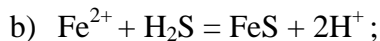
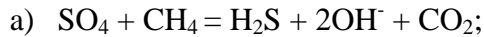
organism lifts it off the sediment surface (Norris, 1989; Bruton, 1991). Whilst these experimental approaches represent great leaps forward in our taphonomic understanding, greater finesse is required to determine the relative influences of the many variables (see Sansom, this volume).

Breaking the mold: Gehling's death mask hypothesis.—

Once formed, the impressions must be rapidly stabilized in order to be preserved. Groundbreaking work into understanding this mode of preservation, and particularly how biological structures could be preserved in their original positive relief, was made by Gehling (1999), based on observations from the Flinders Ranges. The elegant “death mask” hypothesis he proposed consists of four main stages:

- 1) Organisms living on a microbial mat were smothered by sediment;
- 2) Labile or fluid-filled organisms/tissues decayed rapidly, leaving impressions which were infilled by sediment from the overlying bed, while more robust organisms/tissues persisted;
- 3) Sulfur-reducing bacteria exploited the organic material of both carcasses and mat, releasing reduced sulfur compounds which combined with iron in the sediment, resulting in the formation of pyrite. This pyrite coated the lower surface of the now-collapsed labile organisms/tissues, and the upper surface of the recalcitrant organisms/tissues, stabilizing the impressions and forming the so-called “death mask”;
- 4) Death masks which formed over more recalcitrant tissues were infilled from below by still-unlithified sediment.

The pyrite thus formed is observed on the base of the event bed, comprising a sole veneer of sediment grains infilled by interstitial pyrite; this layer is typically no more than a few sand grains thick (Gehling et al., 2005; Mapstone and McIlroy, 2006). The reactions involved can be summarized thus (Gehling et al., 2005):



Several factors influence pyrite precipitation, which in turn influences the anatomical fidelity of the resulting impression (Darroch et al., 2012; Meyer et al., 2014b). In pyritization, a balance exists between the quantity, quality and distribution of organic matter, and the availability of sulfate and iron ions in the system (see Farrell, this volume). Several factors may have contributed to the comparative prevalence of this taphonomic mode in Ediacaran systems. First, the buried microbial mat provided both a diverse population of decay bacteria and a ready supply of organic matter at the horizon which hosts the fossils (Gehling, 1999). Secondly, sealing of the sediment by the microbial mat re-establishing on top of the event bed may have been particularly important in isolating the now anoxic/dysoxic pore waters from the oxic water column above, controlling the availability of sulfate ions and maintaining anoxic pore waters in even porous sediments (Gehling et al., 2005; Callow and Brasier, 2009b).

This model has since been expanded upon (Gehling et al., 2005; Mapstone and McIlroy, 2006; Laflamme et al., 2011), as its ubiquity and variability has become

276 appreciated. Comparable pyrite sole veneers have been described from the Amadeus
277 Basin of Australia (Mapstone and McIlroy, 2006), and inferred from hematite partings
278 (Gehling et al., 2005) and Fe and S within preserved microbial mats (Laflamme et al.,
279 2011) from Newfoundland. Fossils from the Nama Group were originally thought to have
280 been preserved via a different process (Narbonne, 2005), but this has since been
281 questioned (Meyer et al., 2014a, 2014b). These fossils have a thin coating of pyrite that
282 lines the voids left by the external walls of the organism. The thinness of the pyrite
283 coating has been attributed to the dearth of organic matter in the system; in the absence of
284 a microbial mat, the sole source of organic matter was the carcass and sediment (Meyer et
285 al., 2014a, 2014b). Pyritization has also been implicated in the preservation of specimens
286 from the White Sea (Dzik, 2003) and the Gaojiashan biota (Cai et al., 2012), and replaces
287 the organic wall of frond stems from Siberia (Steiner and Reitner, 2001).

288 Formation of a pyritiferous sole veneer or parting is also key in the recovery of
289 the fossils: it provides a surficial horizon which is more susceptible to weathering than
290 the bulk rock (Mapstone and McIlroy, 2006; Meyer et al., 2014a, 2014b), allowing the
291 rocks to split along those horizons where fossils have been preserved. The red iron-oxide
292 and iron-oxyhydroxide staining resulting from this weathering also aids field
293 identification of likely fossil-bearing surfaces (Fig. 3e,f; Gehling et al., 2005).

294 If pyrite is the only mineral involved in making the death mask, then the level of
295 morphological detail retained in fossils is difficult to reconcile with their collapse and/or
296 decay prior to pyritization (which does not occur until step 3 of Gehling's 1999 model).
297 Recent evidence suggests that either authigenic aluminosilicate templating (see below) or

the microbial mat (see *Binding it all together*, below) may have stabilized the impression during the earliest stages of its formation (Darroch et al., 2012).

Coatings of clay – the timing of aluminosilicate mineralization.

The role and importance of clay minerals in soft tissue preservation is controversial, but they have been generated experimentally during early stages of decay (Darroch et al., 2012). Clays are most famously invoked in the preservation of Burgess Shale fossils, although the precise timing of its formation is debated (Orr et al., 1998; Gaines et al., 2005; Butterfield et al., 2007; Page et al., 2008; Anderson et al., 2011). Authigenic clays were first suggested to have played a role in the preservation of Ediacaran fossils by Wade (1969), and they have recently been noted in association with fossils in several Ediacaran localities. In the Amadeus Basin, aluminosilicates occur as part of a complex suite of very early diagenetic minerals, which consist of authigenic quartz, illite, K-feldspar, chlorite, smectite, glauconite, and lastly hematite after pyrite (Mapstone and McIlroy, 2006). These cements fill interstitial space between the sand grains of the enclosing sediment, and stabilized the fossil impressions throughout burial. In addition to the interstitial minerals, these authors noted a “matted” fabric of clay minerals, hematite and silt grains which form a superficial coating on the fossils. Rather than representing a death mask, which would have additionally included pyrite framboids, they interpreted this coating to record fines falling out of suspension onto the associated mat prior to burial (Mapstone and McIlroy, 2006).

Clay minerals and pyrite appear involved in the preservation of fossils from the Doushantuo and Dengying formations (Anderson et al., 2011) and from the Gaojiashan

Lagerstätte (Cai et al., 2012; Meyer et al., 2012). In the Gaojiashan Lagerstätte, three-dimensional fossils which have the finest level of three-dimensional morphological detail are pervasively pyritized (Fig. 1a,b,f); two-dimensional carbonaceous compression fossils are stabilized by Fe-rich clay minerals (Fig. 1c,d,e,g). Based on their composition, spheroidal aggregate habit, and the fact that the fossils they template retain poorer morphological detail, these clays are thought to reflect relatively late diagenetic replacement of an earlier mineral which stabilized the fossil but which formed later in the diagenetic sequence than pyrite (which is associated with the best fossils).

An association of pyrite and Fe- and Mg-rich aluminosilicates has also been documented from the Fermeuse Formation in Newfoundland (Laflamme et al., 2011), which is notable for its abundance of discoidal fossils assigned to *Aspidella terranovica* Billings (Billings, 1872; Gehling et al., 2000). This study found that *Aspidella* specimens interpreted as holdfasts were preserved in three dimensions, and were coated in a thin layer of finer-grained material. The interior of the holdfast and the surrounding sediment is sand-grade and principally composed of Si and Na (analyzed using EDS); the interior sediment was slightly richer in Al, Ca, K, Fe, Na, Mg, P, and Mn, but poorer in C. This was thought to record a vital effect, with sediment inferred to have been incorporated into the holdfast during life (Laflamme et al., 2011). However, it could alternatively record post-mortem precipitation of clay minerals within the holdfast as a function of its specific decay microenvironment. The finer-grained material coating the fossils had higher concentrations of Al, Mg, Fe, Ti, K, Mg, and S than both the holdfast interior and the exterior sediment. This finer-grained layer, which is of variable thickness, was interpreted

to record authigenic aluminosilicate and pyrite growth in a decaying biofilm which had completely surrounded the holdfast during life (Laflamme et al., 2011).

Authigenic chlorite also preserves compressed discs in the Jinxian biota, which are found in stratigraphic association with carbonaceous compressions of *Chuaria*, *Shousienia* and *Tawuia* (Zhang et al., 2006).

Caught on film: carbonaceous compressions

Carbon compression fossils are best known from Chinese localities, with examples from the Gaojiashan Lagerstätte (Fig. 1c,d; Cai et al., 2012), Doushantuo Formation (Fig. 5a; Anderson et al., 2011), Miaohé (Xiao et al., 2002; Zhu et al., 2008), Lantian (Fig. 5c–f; Yuan et al., 2011) and Jinxian (Zhang et al., 2006) biotas, and also from the Denying Formation (Sun, 1986). Preservation of fossils in this mode has enabled inferences about the nature of the original composition of the organism (i.e., recalcitrance) and, by extension, phylogenetic affinity (Zhu et al., 2008) to be made. Assemblages of the ~550 – 590 Ma Miaohé biota are found in black shales in the Yangtze Gorges, and host a diverse range of macrofossils, many of which are interpreted as algae based on the sub-millimetric resolution of preservation and the observation of delicate thalli-like structures (Xiao et al., 2002). Specimens preserved in a comparable manner to the Miaohé fossils are documented from the White Sea and from Siberia, where they occur in finely-laminated, silicified calcareous mudstones of the Khatyspyt Formation (Steiner and Reitner, 2001; Grazhdankin et al., 2008). Kerogenization of carbonaceous compressions has been documented from the Gaojiashan, and has been inferred to have contributed to stabilization of the fossils (Cai et al., 2012).

Although the paleoenvironments are very different, carbonaceous compression fossils do not seem to differ from typical Phanerozoic fossils such as Carboniferous plants (see Locatelli, this volume) preserved in this way: at first glance, they appear to be a simple film of organic carbon compressed onto the sediment surface. However, the extent of microbial influence in their preservation is uncertain, particularly for those fossils which also have a moldic component. Although comparisons have been made between Ediacaran carbonaceous compressions which are associated with clay minerals and/or pyrite, and preservation in the Burgess Shale (e.g. Anderson et al., 2011; Cai et al., 2012; Meyer et al., 2012), the presence or influence of any differences in sedimentology, pore water chemistry or even nature of microbial community on preservation between the sites is unknown. Given the uncertainty regarding the phylogenetic affinity and biological composition of Ediacaran organisms, it is also unclear to what degree the composition of tissues would affect the quality or type of preservation within this taphonomic spectrum.

Bridging the taphonomic void

The large number of biotas now known enables the preservation of single taxa to be compared across different paleoenvironments and taphonomic windows, and thus the biases imparted by each to be elucidated (Fig. 5; Grazhdankin et al., 2008; Zhu et al., 2008). This promises to allow original ecological variability in assemblages to be distinguished from secondary, taphonomic effects. The exclusion of certain organisms from deposits in which fossils are preserved as carbonaceous compressions has been shown to be a purely taphonomic artifact, and may record differences in original tissue composition (Grazhdankin et al., 2008). In the Khatyspyt Formation of Siberia, fossils are

preserved in two, facies-dependent, modes: as carbonaceous compressions and by authigenic carbonate cementation. *Charnia* is commonly found in the latter, but is only observed in the former as “phantoms” where it fortuitously distorts co-occurring carbonaceous films (Grazhdankin et al., 2008). In some cases, individual fossils are preserved in multiple modes, suggesting a complex taphonomic pathway (Cai et al., 2012). The fine balance between these modes may provide insights into the nature of the original organic material, and/or the chemistry of the host sediment. In order to preserve a carbonaceous compression, decay by sulfate-reducing bacteria, and hence pyrite formation, must be halted early. This may be accomplished by overwhelming the system with disseminated organic carbon, or by limiting diffusion of sulfate from seawater into the sediment.

BINDING IT ALL TOGETHER: THE IMPORTANCE OF MICROBIAL MATS

In contrast to much of the Phanerozoic, microbial mats played a conspicuous role in the preservation of soft parts in the Ediacaran, and may be at least partly responsible for the comparative abundance of soft-tissue preservation at this time (Seilacher, 1984; Gehling et al., 2005). Their ubiquity on the sea-floor is widely supported by a myriad of field and petrographic fabrics (e.g. Gehling, 1999; Steiner and Reitner, 2001; Noffke et al., 2002; Gehling et al., 2005; Grazhdankin and Gerdes, 2007; Callow and Brasier, 2009b; Wilby et al., 2011; Lan and Chen, 2012). These fabrics include microbially-induced sedimentary structures (“MISS”, *sensu* Noffke et al., 2001) such as wrinkle marks, reticulate network fabrics, old elephant skin textures and pustular fabrics (see

Gehling, 1999, and refs. therein), as well as “bubble trains” (Laflamme et al., 2012; though see Brasier et al., 2013 for an alternative explanation of these structures). Despite this abundant evidence, there are only scant reports of microbial body fossils outside of the Doushantuo Formation of China (Hofmann et al., 1979; Callow and Brasier, 2009a).

The timing of pyritization relative to collapse of the organism as per the death mask model (see discussion above), and the attendant difference in sense of relief of the resultant fossil, testifies to the importance of microbes in Ediacaran preservation (Gehling et al., 2005; Narbonne, 2005). Sites in the Flinders Ranges with thicker microbial mats (evidenced by prominent surface textures) show negative hyporelief preservation of fossils: pyrite formed before collapse of the organisms. By comparison, sites with thinner microbial mats (relatively subdued microbial textures) exhibit composite and shallow, positive hyporelief preservation: pyrite formed after collapse of the organisms. Therefore, a thicker mat induced earlier formation of a death mask. Similarly, Narbonne (2005) attributes “Flinders-style” preservation to rapid mineralization related to the presence of the thick mats which could develop in the environments where this style is common (Seilacher, 1984; Gehling, 1999). In contrast, he suggests that organisms preserved either by poor examples of “Flinders-style” preservation or by “Nama-style” preservation are attributable to their occurrence in environments above fair-weather wave base, which would have lacked a well-developed mat. In contrast, “Fermeuse-style” preservation has been suggested to have arisen from failure of the overlying bed to lithify before complete decay of the organism, due to either an absence of a mat, or the presence of a mat composed entirely of heterotrophic and/or sulfur-oxidizing bacteria (Narbonne, 2005).

434 Additionally, the extracellular polymeric substance (EPS) produced by the
435 microbial mat likely played an important role in the initial formation and stabilization of the
436 fossil impression by binding the sediment grains prior to precipitation of authigenic
437 minerals (Darroch et al., 2012). In a series of experiments, the impressions which retained
438 the finest morphological detail and lasted the longest were those that contained microbial
439 mats (Darroch et al., 2012). These samples preserved exceptional detail (as scored on
440 their taphonomic index) for two weeks after death of the subject, compared to one week
441 in samples with no mat. In mat-hosted samples, precipitation of pyrite precursors initiated
442 after just one day and reached maximum extent after two weeks. In samples without
443 mats, pyrite precursors initiated after two weeks in unsterilized sand, and failed to initiate
444 in sterilized sand. The importance of microbial mats in preservation is highlighted when
445 this recent experimental approach is compared with those conducted in the 1980s and
446 1990s, particularly given the relatively low return rate of recognizable impressions in
447 experimental runs with otherwise similar conditions (Norris, 1989).

448 449 ***ASPIDELLA* EVERYWHERE! THE ABUNDANCE OF HOLDFASTS**

450
451 Without doubt, the most abundant macrofossils found throughout the Ediacaran
452 are discoidal. The majority of these belong to the taxon *Aspidella*, which brings under its
453 umbrella many different, and previously taxonomically distinguished, forms (Gehling et
454 al., 2000). They are thought to principally represent holdfasts (Gehling et al., 2000;
455 Laflamme et al., 2011; MacGabhann, 2007), and may be so profuse that they entirely
456 cover bedding surfaces, as seen in the Fermeuse Formation of Newfoundland (Fig. 6e).

Only rarely are holdfasts preserved on the same horizon as their fronds (e.g. in Charnwood Forest, Fig. 2b,d; Wilby et al., 2011); in most localities, either one or the other is preserved. Most holdfasts were likely buried within the sediment, in whole or in part, and their frequent concentric rings are generally interpreted as collapse structures. On certain surfaces, only fronds of certain taxa are associated with holdfasts (e.g. *Charniodiscus* in Mistaken Point, Newfoundland). In these cases, the holdfasts are typically positive epirelief, and have been inferred to record upper surfaces of holdfasts which protruded above the mat surface in life and were preserved before their collapse by rapid mineralization of the ash (Conception-style preservation; Narbonne, 2005). Fronds on these surfaces which appear without associated holdfasts (e.g. *Charnia*) may have had ones which remained entirely buried within the sediment during their life and death (Laflamme et al., 2007), and so are out of the plane of preservation. Some localities preserve only holdfasts and trace fossils (“Fermeuse-style” preservation; Narbonne, 2005).

There may be several taphonomic reasons for the relative abundance of holdfasts to fronds. First, their *in vivo* position within the sediment means that, even if the holdfasts collapse after death or are tugged out, they may still leave an expression, such as collapse structures formed as sediment falls back into the void left after decay (Narbonne, 2005), and shear structures (Tarhan et al., 2010), respectively. Secondly, their anchorage within (Laflamme et al., 2011) or beneath (Mapstone and McIlroy, 2006) the mat means that they are less likely than the frond to be tugged out and removed by the current. Thirdly, their presence within or close to the zone of active diagenesis (the “mixed layer”; Callow and Brasier, 2009b) gives them a higher preservation potential. Finally, there is some

evidence that holdfasts were made of more resistant material; they may be preserved as carbonaceous compressions while stems are replicated by pyrite (Steiner and Reitner, 2001), or as positive epirelief structures while fronds are preserved in negative epirelief (Fig. 2e; 4a,b,d,f; Narbonne, 2005). Their comparative persistence may lead to their preservation even in conditions conducive to only slow mineralization (cf. Darroch et al., 2012).

UNRAVELLING THE RAVAGES OF TIME: BIOSTRATINOMY AND THE GRADATION OF FORMS

Biostratinomy encompasses the effects of post-mortem compaction, contraction (whether by dehydration or bacterial decay), folding, and transport (Gehling et al., 2005), which necessarily influence the final morphology of the fossil. Compared to the Phanerozoic, fewer biostratinomic processes operated in Ediacaran times. Scavengers consume or disarticulate carcasses, but are unknown from the Period, and bioturbation was limited. Those processes which would have endured throughout the Proterozoic into the Phanerozoic are microbial decay and abiotic (physical) disturbance. Syn- or post-mortem distortion of the morphology of the organism by physical processes has been recorded from localities around the globe, and includes wrinkling (Gehling, 1991), folding (Seilacher, 1992) and ripping (Runnegar and Fedonkin, 1992). If different parts of an organism had dissimilar rheologies, they will be affected differently by shared biostratinomic processes. For example, the crenellated part of *Kimberella* shows comparatively greater deformation or wrinkling than the rest of the organism, and is

503 accordingly inferred to have been a broad, flattened “foot” which was less robust than a
504 surrounding, unmineralized shell (Fig. 3a; Fedonkin and Waggoner, 1997).

505 In fossils from Newfoundland, the quality of preservation is seen to decrease
506 along their length; there is greater opportunity for sediment to settle beneath the more
507 distal and lateral parts of the frond during felling (Laflamme et al., 2007). Fronds of
508 increasing size may show variations in susceptibility to current-induced stacking of
509 branches and compression of overall form. The observed change from I to V to U and to
510 O plan-view morphology in *Bradgatia* with overall increase in size may thus reflect
511 increased resistance to this process (Brasier et al., 2013), rather than a purely ontogenetic
512 signal (Flude and Narbonne, 2008). On a finer scale, the branching pattern of
513 rangeomorphs may also be affected during the burial event, for example the current-
514 induced imbrication of primary branches recorded in specimens from Spaniard’s Bay,
515 Newfoundland (Fig. 4; Brasier et al., 2013). If this interpretation is correct, it is
516 unnecessary for these branches to have been either constrained in a sheath or attached to
517 each other, as has been proposed for some taxa (Narbonne et al., 2009).

518 519 **Sweeping up: interpreting “mops”**

520 The potential extent of modification of form by physical disruption is further
521 evidenced by so-called “mop” structures (Tarhan et al., 2010). They have a highly
522 variable appearance and relief, but are consistently aligned parallel to ripped-up stems on
523 the same bed, have well-defined distal margins, and subparallel internal lineations that
524 are orthogonal to the margin. There is a gradation from typical *Aspidella* holdfasts to full
525 “mop”, consistent with these structures representing an effect imposed upon an organism

rather than a discrete taxon. “Mops” are thus interpreted to be the result of current shear on frond holdfasts; the fronds themselves are not preserved, and are suggested to have been either torn off by the current or held above the preservational surface (Tarhan et al., 2010). They may alternatively have been held out of the plane of preservation. Upstream parts of “mops” have a range of distal margin shapes, and are interpreted as buckled and compressed parts of the holdfast, whereas downstream, linear structures are interpreted as torsion-induced stretch marks. The smallest “mops” are expressed solely as distortion of the surrounding sediment, suggesting that the smallest fronds were removed entirely. This may provide an explanation for the lack in many communities of specimens below a few centimeters. Alternatively, it may be that the entire mop spectrum represents disturbance and distortion of the microbial mat arising from plucking out of the holdfast, and are therefore a form of MISS (Laflamme, pers. comm.. 2014).

Is it all rot? Iveshediomorphs

Forms currently referred to as “iveshediomorphs” (Fig. 6a–d) are contentious, and include a wide spectrum of morphologies. These were originally described from Charnwood Forest as discrete taxa, and include *Ivesheadia*, *Blackbrookia*, *Pseudovendia* and *Shepshedia* (Boynton and Ford, 1979, 1995); similar forms in Newfoundland are referred to as “pizza discs”, “lobate discs” and “bubble discs” (Narbonne et al., 2001; Laflamme et al., 2012). A full spectrum between such forms and fronds exhibiting fine detail has been documented from several bedding planes in Newfoundland, leading to the interpretation of “ivesheadiomorphs” as the remnants of dead organisms which were in the process of microbial decay at the time of burial (Liu et al., 2011). The irregular,

549 unusually high relief and often network-like internal features of these forms were
550 suggested to represent a conflation of sediment trapped by EPS and gas derived from the
551 decay process (Liu et al., 2011). However, other authors have suggested alternative
552 explanations: Laflamme et al. (2012) interpret these structures as purely microbial in
553 origin, and Wilby et al. (2011) propose that at least some of the forms may be created by
554 differential loading on the fossil-bearing surface following collapse of organisms within
555 the overlying bed.

556 557 **All full up: the timing of sand infills**

558 Resolution of both the mechanism and timing (*in vivo* or post-mortem) of
559 sediment infill is of great significance for current interpretations of the biology of the
560 organisms in which they are found. Rangeomorphs and erniettomorphs have been
561 interpreted as osmotrophs, absorbing dissolved organic carbon from the water column
562 (Laflamme et al., 2009). The proposed model requires the organisms to have had a very
563 small (< 2 %) volume of metabolically-active material, which could potentially have been
564 achieved by *in vivo* incorporation of sediment (Laflamme et al., 2009). If this can be
565 demonstrated, then an osmotrophic mode of life is plausible; if not, then the inert material
566 must be a fluid, which might be expected to exert its own influence on the rheology of the
567 organism and on its taphonomic behaviour.

568 Sand fills observed in stalks of fronds from the Amadeus Basin exhibit ripple
569 cross-lamination contiguous to that of the surrounding sediment, and are convincingly
570 demonstrated to record post-mortem slumping of sediment into the stalk (Mapstone and
571 McIlroy, 2006). In contrast, it has been suggested that sand in the holdfasts (“bulbs”) and

central stalks of fossils of *Rangea* from Namibia was incorporated into the body of the organism during life, based on the similarity of the sediment within the “bulb” and stalk to that infilling the gutter casts below the fossil (Vickers-Rich et al., 2013). A similar *in vivo* fill has been interpreted in holdfasts from the Fermeuse Formation of Newfoundland (see *Aspidella* section above; Laflamme et al., 2011).

If this *in vivo* interpretation holds, then such a fill could have served to stabilise or anchor the organisms. However, a post-mortem infilling might explain the recorded presence of a gap in the sediment fill part way up the stalk (Vickers-Rich et al., 2013), their Fig. 7.3,7.4), which is hard to reconcile with the *in vivo* interpretation. In contrast, a taphonomic sand infill is more difficult to argue for specimens of taxa such as *Ernietta* which are preserved as three-dimensional sediment casts. There are no obvious tears or punctures through which sediment could enter, and it might be expected that a post-mortem fill would be less pervasive than an *in vivo* one, either restricted to a few (damaged) individuals or to partial infills. If these fossils represent templates of the external surface which have been infilled by uncemented sediment (Meyer et al., 2014a, 2014b), how did this sediment penetrate a more-or-less continuous veneer after its formation? Perhaps these organisms did have an *in vivo* fill; in that case, how was this achieved?

Fully understanding the biostratinomy of the organisms can be hugely useful in determining various aspects of their biology. The observation of torsion-induced stretch marks in holdfasts indicates both that these were firmly anchored in the sediment, and that they were sufficiently elastic to deform rather than snap or tear. This, in turn, casts doubt on interpretations of structures such as “mops” and isolated *Aspidella* specimens

which invoke ripping off the frond. We can infer that tissues which do show evidence of tearing under the same conditions were likely more rigid. That organisms can fold over indicates a certain degree of flexibility, and the style of fold can provide additional information on body rheology: assuming no decay had occurred, those that kink over at a sharp line (e.g. dickinsoniids) were likely more rigid/solid than those which show more sinuous or irregular folds (e.g. the rangeomorph, *Fractofusus*).

KNOW YOUR LIMITS: PRESERVATIONAL BIASES AND UNKNOWNABLE UNKNOWNNS

Although soft-tissue preservation is reasonably common in the Ediacaran, with bed after bed preserving fossils, it is by no means perfect, and has its own inherent problems and biases. Two-dimensional records of three-dimensional organisms are by their very nature either composite (with structures at multiple levels in the organism compressed together into one plane) and/or partial; usually, only one side of an organism is cast. This is most problematic for organisms with a highly three-dimensional morphology, such as multifoliate rangeomorphs (*sensu* Laflamme and Narbonne, 2008) and erniettomorphs, as opposed to flatter organisms such as unifoliate rangeomorphs (*sensu* Laflamme and Narbonne, 2008) and dickinsoniids. Imagine how the two-dimensional impression of a bushy plant would compare to its three-dimensional morphology, and how much more complex and confused, and less representative of the living morphology, this would be in contrast to the compression of a plant like a fern.

617 This is exemplified in the changing morphology and decreased clarity of branching of
618 *Bradgatia* with increasing size (see *Biostratinomy* section above).

619 As in any Lagerstätte, we must still consider what has not been preserved, in
620 terms of both parts of the organisms and of whole organisms. For those specimens which
621 have been transported and deposited in death assemblages (Namibian channel-fill
622 deposits), we have lost information regarding their *in situ* ecology, such as their relative
623 abundances and spatial distributions, as well as their position relative to the sediment-
624 water interface. The limits of the resolution of preservation also render invisible any
625 meiofauna which may have been present, and so we have no knowledge of these
626 components of the ecosystem (Sperling et al., 2013). Any organism capable of motility
627 (Liu et al., 2010) has a greatly reduced chance of being captured in the ecosystem than its
628 sessile compatriots. Despite early (now-refuted) suggestions of discoidal fossils as
629 medusoids (e.g. Sprigg, 1947, 1949; Glaessner and Wade, 1966), there is to date no
630 convincing evidence of pelagic forms. As such, we have no knowledge whatsoever of the
631 macroscopic life which may have been present in the water column during the Ediacaran.
632 By extension, if these organisms did have a planktonic dispersal stage (discussed in
633 Darroch et al., 2013), it is unlikely that this would be captured. Apparent support for such
634 a life stage is the lack of preserved individuals smaller than a centimeter, despite sub-
635 millimetric preservation, and that even the very smallest fossils currently known are
636 identical in morphology to adult forms (Liu et al., 2012, 2013).

637 For the organisms that we do see, the overwhelming dominance of two-
638 dimensional, external moldic preservation means that we have little, if any, evidence of
639 internal anatomy. Reported examples of internal “struts” in rangeomorphs from

Spaniard's Bay (Narbonne, 2004) have subsequently been explained as branches filled by sediment casting the underside of the upper surface and creating positive epirelief impressions (Fig. 4b,c), rather than casting the lower surface and creating negative epirelief impressions (Fig. 4a,d,e,f), as is typical at this locality (Brasier et al., 2013). The ridges originally interpreted as struts are accordingly inferred to be the divisions between higher-order branches (Brasier et al., 2013). The differential collapse of branches within the frond (Fig. 4b) could be taken to suggest the *in vivo* presence of a hydrostatic skeleton which became punctured and subsequently deflated, but could alternatively simply record relaxation in response to decay (Brasier et al., 2013). Discrimination between the competing hypotheses will remain challenging until further evidence comes to light.

Three-dimensional fossils are rare, restricted to occurrences of "Nama-style preservation" (Narbonne, 2005) and within certain carbonate deposits (Xiao et al., 2005; Grazhdankin et al., 2008): most fossils have relief but are essentially two-dimensional (in the sense that only one side, or a composite of both sides, of the organism is captured). This means that, for the vast majority of specimens, there is no evidence for what the other side looked like. There is also little indication of the original volume of the organism, due to both biostratinomic and burial compaction.

Explanations for the absence of internal structures in Ediacaran fossils include: 1) decay outpaced their fossilization; 2) the microenvironment created by their decay inside the body cavity was not conducive to fossilization; 3) the extrinsic environment was incompatible with their fossilization, perhaps because of sediment sealing; 4) comparatively low oceanic sulfate concentrations (e.g. Canfield et al., 2008) favored pyritization of the organisms' external surface; or 5) the organisms lacked substantive

internal structures. Until sufficient and suitable fossils are available to permit the undertaking of destructive analyses (e.g. Laflamme et al., 2011), resolution of this question may remain elusive.

CONCLUSIONS AND QUANDARIES

Exciting progress is being made in several fields of Ediacaran endeavour, including paleoecology (Clapham and Narbonne, 2002; Clapham et al., 2003; Darroch et al., 2013), anatomy (e.g. Fedonkin and Waggoner, 1997; Narbonne et al., 2009; Brasier et al., 2012; Vickers-Rich et al., 2013), ontogeny (e.g. Laflamme et al., 2004; Antcliffe and Brasier, 2007) and phylogentic relationships (e.g. Erwin et al., 2011; Laflamme et al., 2013). Taphonomic processes and biases impact all aspects of paleobiology; an understanding of these is therefore paramount if we are to further elucidate the nature of the original organisms and their communities. Great strides are being made, with elegant experimental work (McIlroy et al., 2009; Darroch et al., 2012) enhancing detailed petrographic and field-based studies (e.g. Gehling, 1999; Xiao et al., 2005; Grazhdankin et al., 2008; Laflamme et al., 2011). Taphonomic modes as disparate as moldic preservation, pyritization, carbonaceous compression, clay mineral precipitation, and carbonate mineralization are all observed in Ediacaran sites across the globe, sometimes with multiple modes within a single fossil (Cai et al., 2012). There does appear to be a broad correlation between taphonomic style and depositional environment (e.g. Narbonne, 2005; Grazhdankin et al., 2008), with moldic and death mask preservation most common in siliciclastic and volcanoclastic settings, and carbonaceous compression

fossils occurring predominantly in shales and carbonates. Clay mineralization appears to occur in all environments. However, the extent of the influence exerted by paleoenvironment, and of the disparate factors this includes, remains uncertain. Fifteen years on from the proposal of the death mask model (Gehling, 1999), perhaps it is time to consider how the plethora of biotas featuring pyritization relate to this model, and to one another.

The location of an organism with respect to the sediment-water interface imparts a significant taphonomic bias, as those structures which were located within the sediment during life were more readily preserved in death. This bias is a likely cause of the greater abundance of holdfasts with respect to fronds (see *Aspidella* section above). Other taphonomic biases are much more poorly understood, with a few rare exceptions (e.g. the scarcity of rangeomorphs in Miaohu-type preservation; Grazhdankin et al., 2008). Elucidation of these biases is of absolute import for paleoecological studies: as they become known, we can begin to appreciate which site-to-site differences in assemblage composition are original (ecological), and which are secondary (taphonomic overprint).

Perhaps the most appropriate way to investigate these biases is through expansive experimental work. Such studies may also help to answer why moldic preservation is so prevalent in the Ediacaran as opposed to the Phanerozoic. The effects of a range of physical and chemical parameters have been tested in various taphonomic experiments (see e.g. Briggs, 2003, and refs. therein; Sansom, this volume). However, such studies have yet to be systematically extended to investigate systems which would be more applicable to the Ediacaran, e.g. those with microbial mats (with the exception of Darroch et al., 2012). Trace metals such as molybdenum, which are limiting nutrients for life (e.g.

(Glass et al., 2012), are becoming widely used as tracers of productivity and ocean redox conditions in the Proterozoic (e.g. Scott et al., 2008), but nothing is known about their specific effects on decay-related microbial activity, and therefore on taphonomy.

A major caveat to such taphonomic experimentation is the enduring uncertainty surrounding the original biological composition of Ediacaran organisms, which will likely only be resolved upon discovery of an assemblage preserving cellular-level detail. Currently, inferences regarding relative degrees of robustness and rigidity may be made based on biostratigraphic grounds, but these can only tell us so much. Taphonomic experiments sampling a wide variety of tissue and cell types from as many branches of the tree of life as possible may provide our best hope: by comparing the behaviours of different biological compositions to features seen in fossils, it may be possible to relate the two, and consequently to infer the original composition of the organism or its parts. Of course, this must be repeated for the many potential variables already discussed in order for any such inferences to be made with any degree of confidence, rendering the number of experiments required unfeasible.

One quandary peculiar to Avalon Assemblage localities is the true nature(s) of “ivesheadiomorphs”. Do all “ivesheadiomorphs” as currently defined (Liu et al., 2011) have the same genesis? Are they all taphomorphs of known taxa? Are they all microbial colonies (Laflamme et al., 2012)? Do any represent discrete macro-organisms (Boynton and Ford, 1979, 1995)? Do some have a different origin? The answers to these questions have great potential impacts for several aspects of paleoecological studies, including living biomass, species diversity and disparity, and spatial distributions: if these structures are dead and decaying organisms, they must inherently have a different effect

on the rest of the community than if they were living organisms. Whatever they are, why is their occurrence seemingly restricted to Avalonian, deep-water systems? Is this an environmental signal, or a function of the microbial community present? If they really do include taphomorphs, could different organisms have created distinguishably different forms, and could these be related back to their progenitors (cf. Liu et al., 2011)?

Paradigm shifts in understanding will probably depend not on the development of analytical techniques, but rather on the discovery of new, higher-resolution preservational windows.

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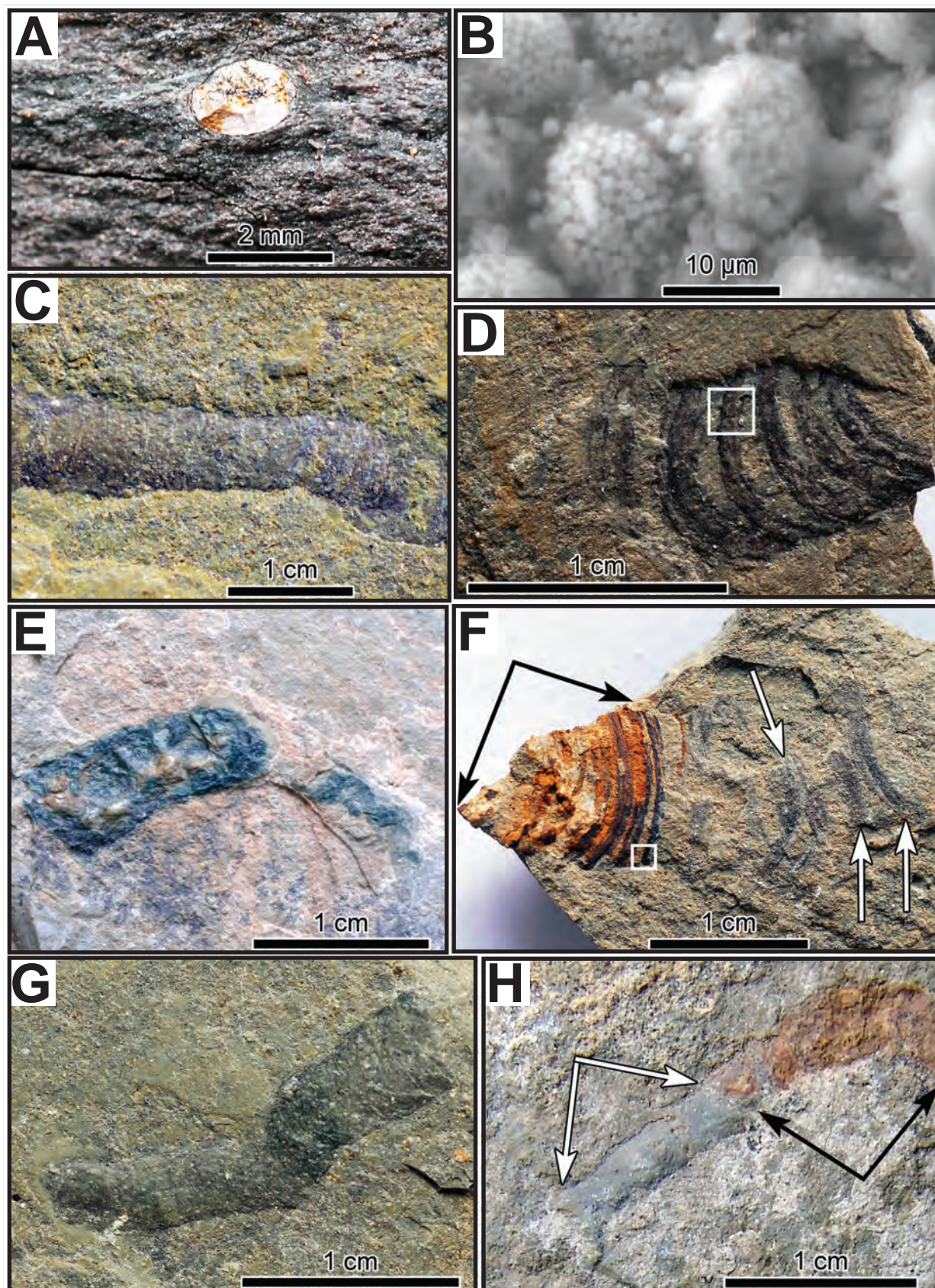


FIGURE 1.—Preservation of single fossils from the Gaojiashan biota in multiple taphonomic styles, reproduced from Cai et al., 2012 with the permission of Elsevier. All reflected light photographs except b (SEM photomicrograph). a) Cross-section through a pyritized *Conotubus hemiannulatus* specimen; b) pyrite framboids on the exterior of the specimen in a); c, d) carbonaceous compression fossils; e) replication by green-coloured aluminosilicate minerals; f) specimen exhibiting both pyritization (rust-coloured area, black arrows) and carbonaceous compression (grey parts, white arrows); g) specimen preserved in both carbonaceous compression and greenish aluminosilicate minerals (central part of the tube); h) specimen exhibiting all three preservational styles (pyritization between black arrows, carbonaceous compression and aluminosilicate mineral replication between white arrows). Rectangles in c, d, and f indicate areas analyzed via ESEM EDS.

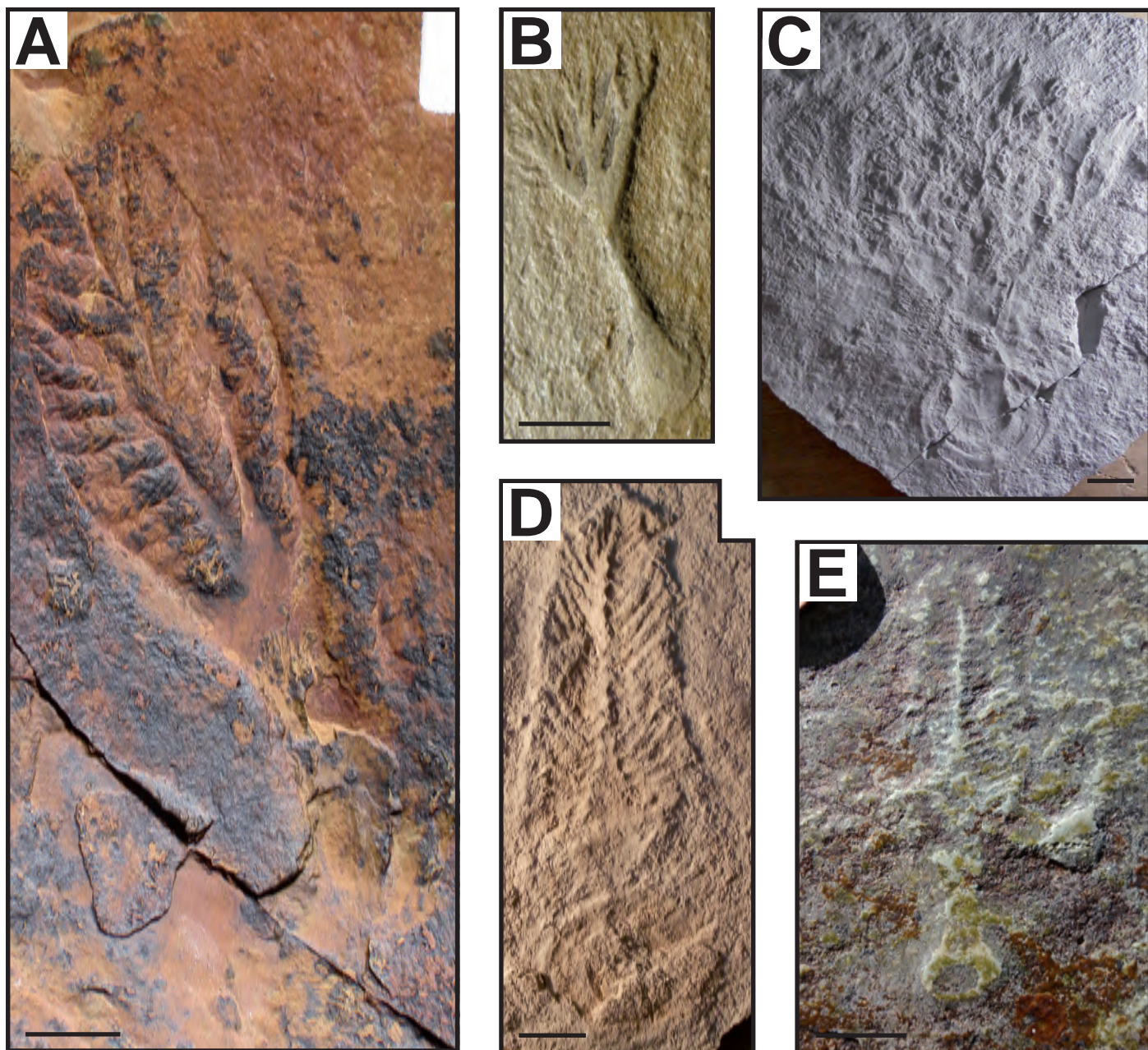


FIGURE 2.—a) Exceptional preservation in the Avalon Assemblage - a *Culmofrons* specimen from the Bonavista Peninsula, Newfoundland. Note that the frondose portion of the fossil is preserved in negative epirelief, and the stalk is preserved in positive relief. Photo courtesy of Alex Liu. b) Small *Primocandelabrum* specimen from the Bonavista Peninsula, with holdfast, stalk and branch axes preserved in positive epirelief. The frondose portions are not evident in this fossil. c) A *Primocandelabrum* specimen from Charnwood Forest, UK. Note that in contrast to b, the stalk and holdfast are preserved in negative epirelief, indicating a difference in the relative timings of collapse and lithification of the beds in the two sites. Plaster cast of GSM 105969; d) *Charniodiscus* specimen from Charnwood Forest, with central and branch axes preserved in low positive epirelief, and the holdfast in negative epirelief. Plaster cast of GSM 106069; e) *Charniodiscus* specimen from the Bonavista Peninsula, Newfoundland, with central and branch axes and the holdfast all preserved in positive epirelief. Scale bars are 2 cm (a, c, d) and 1 cm (b, e).

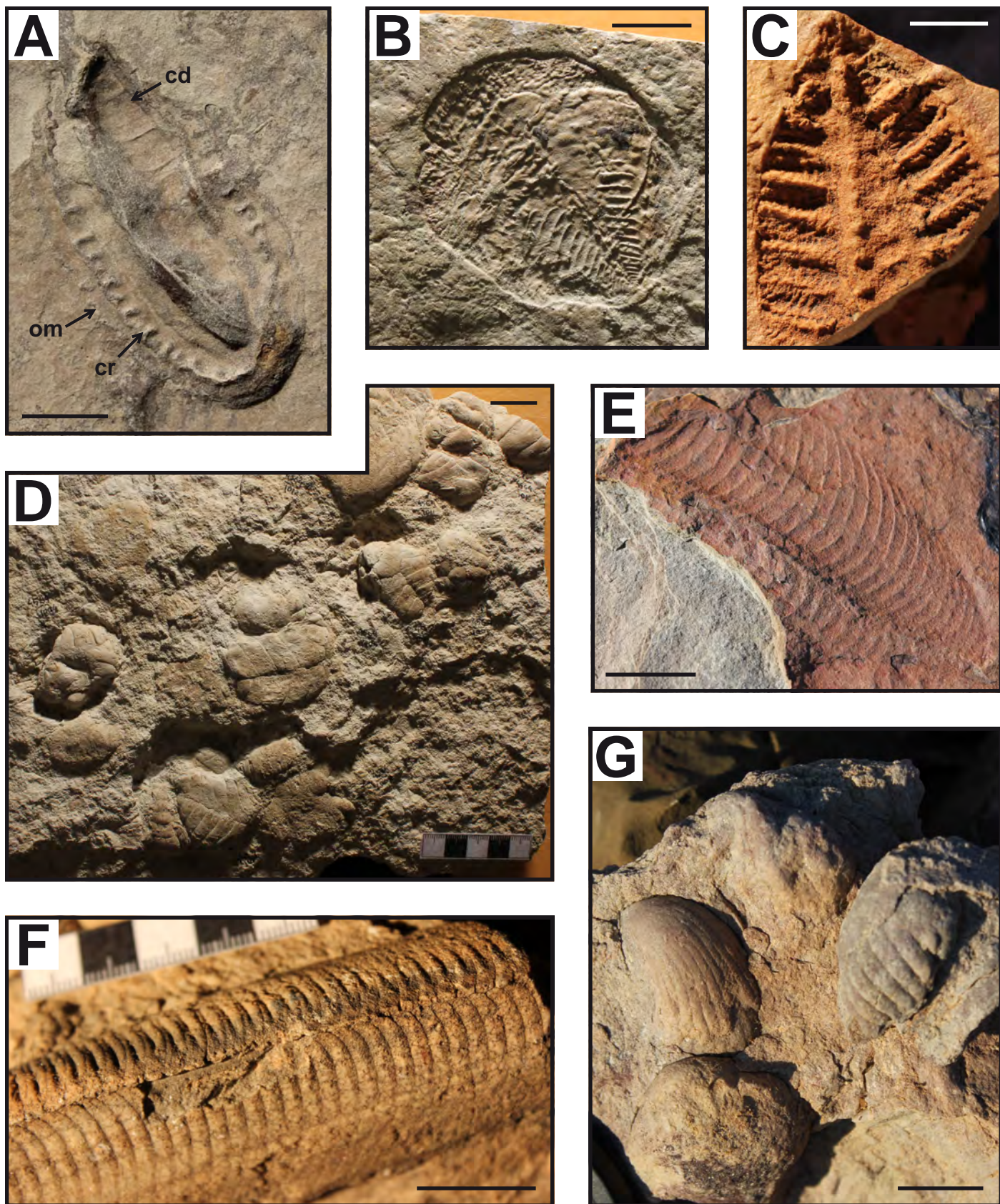


FIGURE 3.—Examples of mouldic preservation from the White Sea (a, b, d) and Namibia (c, e, f, g). a) *Kimberella* displaying three distinct zones (om = outer margin; cr = crenellated part; cd = central depression); b) *Yorgia* with a disturbed proximal region (upper left); c) *Rangea* with clear rangeomorph branching; d) death assemblage of *Ventogyrus*; e) *Pteridinium* specimen with prominent red staining on the bed surface; f) *Pteridinium*; g) *Ernietta*. Scale bars are 1 cm (a, b) and 2 cm (c – g). All images courtesy of Marc Laflamme.

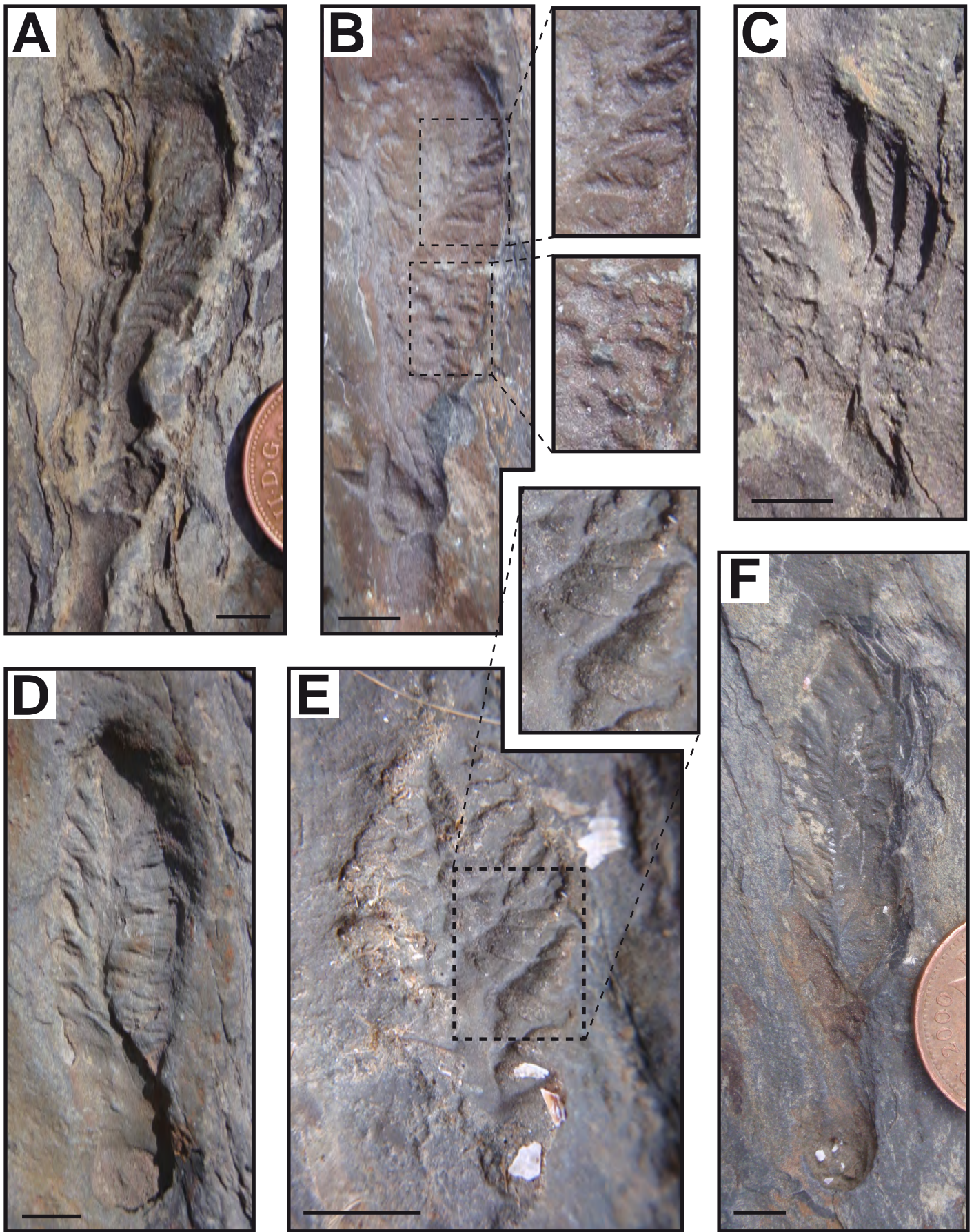


FIGURE 4.—Specimens from Spaniard's Bay, Newfoundland. a) *Bradgatia* with exquisite preservation down one side only; b) *Beothukis* with good, negative epirelief, distal (upper insert) and poorer, positive epirelief, proximal preservation (lower insert); c) unidentified frond with relatively high relief on the right, and lower relief on the left; d) *Beothukis* with a prominent disc structure at the base and either a basal sheath or current scour (smooth area); e) *Charnia* with negative epirelief preservation; f) *Trepassia* with exquisite preservation and a basal sheath/ current scour. All scale bars: 5 mm.

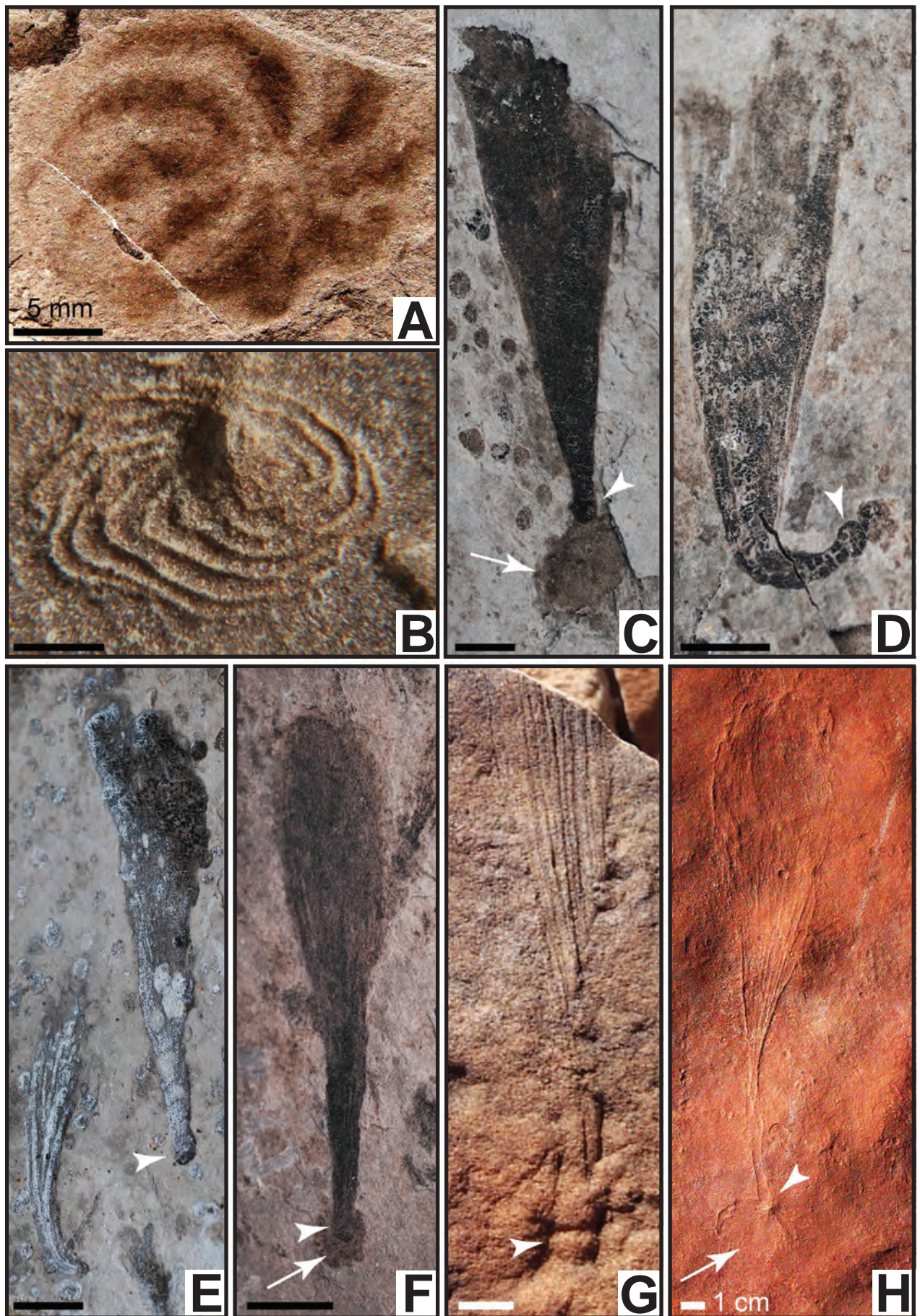


FIGURE 5.— One species preserved in multiple taphonomic modes. Reproduced from Xiao et al., 2013 with permission from GSA. a) *Eoandromeda octobrachiata* preserved in black shale, from the uppermost Doushantuo Formation at Wenghui (China); b) *E. octobrachiata* preserved as a siliclastic mold from the Ediacara Member sandstone (South Australia); c–f): *Flabelophyton lantianensis* preserved in black shales of the lower Lantian Formation (South China); g, h) similar forms from the Ediacara Member sandstone (Australia). Arrowheads point to globose holdfasts, and arrows point to organic mass at base. Black scale bars = 5 mm, white scale bars = 1 cm.

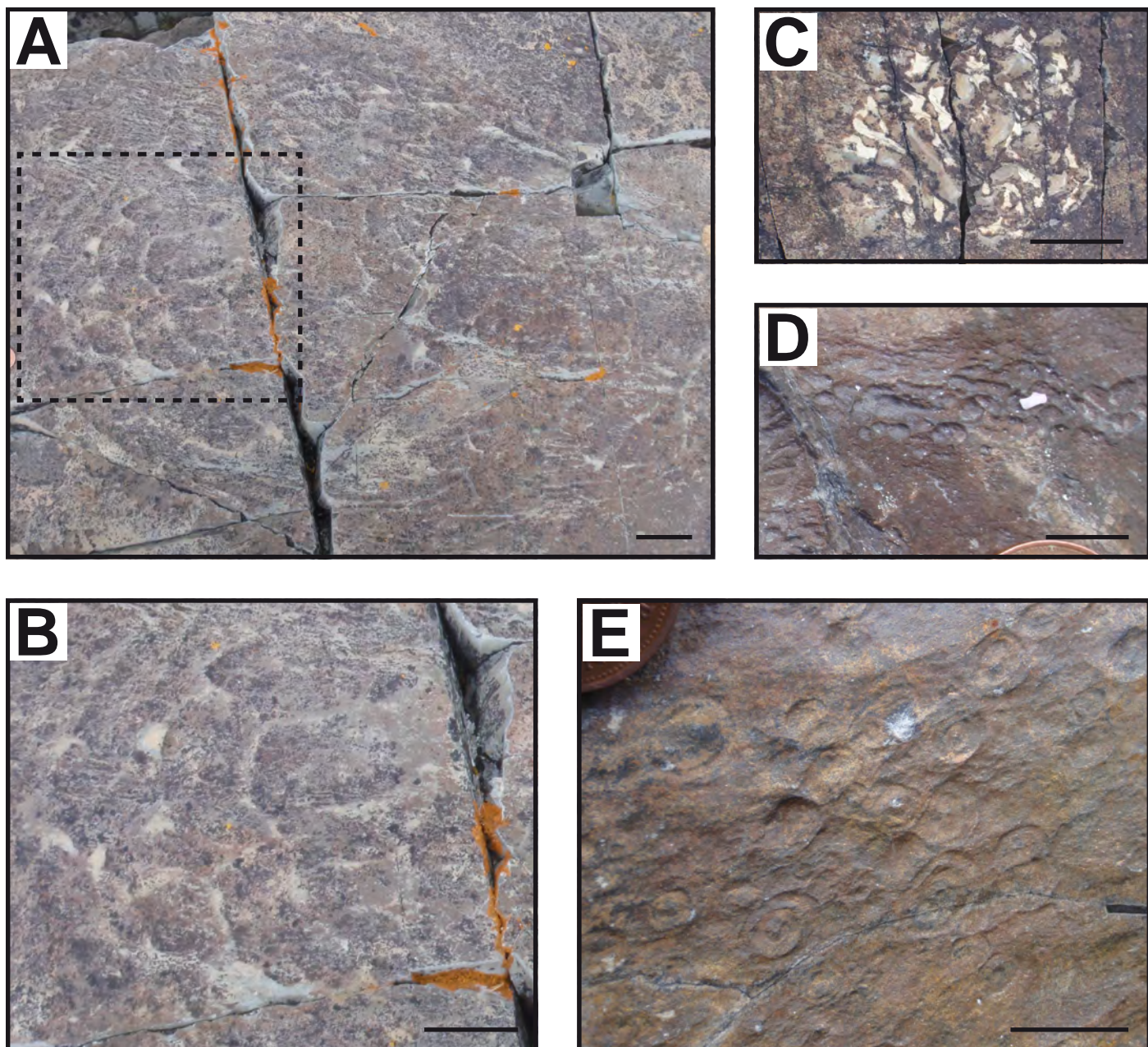


FIGURE 6.—a) Large iveshedidiomorph from Bonavista Peninsula, Newfoundland; b) inset of a) showing reticulate network texture; c) “pizza disc” *sensu* Narbonne 2001 from the Mistaken Point Ecological Reserve, Newfoundland; d) “bubble strip” *sensu* Laflamme et al. 2012 from Spaniard’s Bay, Newfoundland; e) numerous small *Aspidella* holdfasts from Ferryland, Newfoundland. Scale bars are 5 cm (a, b, c) and 1 cm (d, e).