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Journal: Palaeontology

Manuscript ID: PALA-12-17-4127-OA.R1

Manuscript Type: Original Article

Date Submitted by the Author: n/a

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Key words: Romer’s gap, rhizodont, tetrapod, fish, floodplain, Carboniferous, Tournaisan

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Micropalaeontology data.csv
CA axis scores.csv
Raw diversity.dat
NMDS axis scores.csv
Raw diversity dataset.csv
Relative diversity dataset.csv
Relative diversity.dat
A FISH AND TETRAPOD FAUNA FROM ROMER’S GAP PRESERVED IN
SCOTTISH TOURNAISIAN FLOODPLAIN DEPOSITS

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Abstract: The end-Devonian mass extinction has been framed as a turning point in vertebrate evolution, enabling the radiation of tetrapods, chondrichthyans, and actinopterygians in the Carboniferous and beyond. Until very recently ‘Romer’s Gap’ rendered the Early Carboniferous a black box standing between the Devonian and the later Carboniferous, but now new Tournaisian localities are filling this interval. Recent work has recovered tetrapod and lungfish diversity in contrast with previous expectations. However, the composition of Tournaisian faunas remains poorly understood.

Here we report on a Tournaisian vertebrate fauna from a well-characterized, narrow stratigraphic interval from the Ballagan Formation exposed at Burnmouth, Scotland. Microfossils suggest brackish conditions and the sedimentology indicates a low-energy debris flow on a vegetated floodplain. A range of vertebrate bone sizes are preserved. Rhizodonts are represented by the most material, which can be assigned to two taxa. Lungfish are represented by several species, almost all of which are currently endemic to the Ballagan Formation. There are two named tetrapods, Aytonerpeton and Diploradus, with at least two others represented by additional specimens. Gyracanths, holocephalans, and actinopterygian fishes are represented by rarer fossils. This material compares well with vertebrate fossils from other Ballagan deposits elsewhere.

The faunal provides an opportunity to revisit previous work on the end-Devonian extinction. Faunal similarity analysis using an updated dataset of Devonian-Carboniferous (Givetian-Serpukhovian) sites corroborates a persistent Devonian/Carboniferous split. Separation of the data into marine and nonmarine partitions indicates more Devonian-Carboniferous faunal continuity in nonmarine settings compared to marine settings. These results agree with the latest fossil discoveries, and suggest that the Devonian-Carboniferous transition proceeded differently in different environments and among different taxonomic groups.

Key words: Romer’s gap, Tournaisian, tetrapod, fish, floodplain, Carboniferous, rhizodont
Romer's Gap (Romer 1956; Coates and Clack 1995) is a hiatus in the non-marine fossil record that currently spans the Tournaisian stage of the Carboniferous (~359-345Ma). Named for the paucity of tetrapod fossils throughout this interval, the gap has long obscured cladogenic events and ecological transitions implied by differences between the few, mostly aquatic, limbed forms present at the end of the Devonian and the diverse and disparate tetrapods known from the Visean (Wellstead 1982; Clarkson et al. 1993; Pardo et al. 2017). What little we know of Tournaisian tetrapods is drawn from not much more than isolated bones and fragments from localities such as Blue Beach in Canada (Anderson et al. 2015), but with the notable exception of the unique, substantial and partly articulated skeleton of *Pederpes* from Scotland (Clack and Finney, 2005). However, the challenge presented by Romer's Gap extends beyond tetrapods. Few Tournaisian fish localities have been known until recently (Clarkson 1985; Long 1989; Sallan and Coates 2010; Mansky and Lucas 2013; Sallan and Galimberti 2015; Mickle 2017, Richards et al. in press), and these, too, appear impoverished relative to subsequent Visean faunas.

It follows that all three major living vertebrate divisions, actinopterygians (Sallan 2014; Giles et al. 2017), chondrichthyans (Coates et al. 2017, 2018), and tetrapods (Clack et al. 2016; Pardo et al. 2017) underwent major radiations within this key interval. Moreover, it seems likely that these radiations occurred opportunistically, to refill ecological space in the aftermath of serial extinction events that occurred close to end of the Devonian (Sallan and Coates 2010;
Sallan and Galimberti 2015). But, absence of a substantial record of Tournaisian fossil vertebrates obscures the timing and true phylogenetic pattern of the Devonian-Carboniferous transition. It remains unclear whether these large-scale post-extinction diversifications arose from short (Tournaisian) or long (Devonian) fuses. We have almost no perspective on the multiple ecological transitions that took place at this time, especially the ongoing vertebrate invasions of non-marine environments including the likely multiple terrestrialisations of the Tetrapoda. Moreover, the hypothesized abruptness of the end-Devonian mass extinction (Hangenberg event), (Sallan and Coates 2010) and an inferred Lilliput effect (Sallan and Galimberti 2015) deserve to be tested. Already, Tournaisian tetrapod discoveries include a mix of morphologies (Anderson et al. 2015) that are distributed across multiple nodes in recent phylogenetic analyses (Clack et al. 2016). And, a recently-recognized Tournaisian radiation in lungfish (Smithson et al. 2015) contradicts a historical narrative of post-Devonian decline and stasis.

These various observations raise the possibility that the Devonian/Carboniferous divide recovered by Sallan and Coates (2010) might not be expressed in the same way across all environments, particularly given the uneven distribution of end-Devonian diversity loss among taxonomic groups (Sallan and Coates 2010). Thus, there may be discernable time/environment associations, with different levels of taxonomic/ecological continuity across the Devonian/Carboniferous boundary. However, widespread euryhalinity among Devonian-Carboniferous vertebrates (Ó Gogáin et al. 2016; Goedert et al. 2018)
may support hypotheses of a cosmopolitan post-extinction fauna in the
Tournaisian (Friedman and Sallan 2012),

Here we describe the vertebrate fossil fauna from the Tournaisian-age
Ballagan Formation exposed on the beach at Burnmouth, Scotland. The fossils
come from a 1m stratigraphic interval. These new data, along with those from the
Blue Beach locality (Anderson et al. 2015), are added to the dataset of Sallan
and Coates (2010) in order to test their conclusions of a Devonian/Carboniferous
faunal divide against new Tournaisian data.

GEOLOGICAL CONTEXT

The Ballagan Formation crops out in northern England and the Midland
Valley of Scotland. It has previously been referred to as the Cementstone Group
in England and the lower part of the Calciferous Sandstone Measures in
Scotland (Greig 1988), and probably spans the totality of the Tournaisian
(Smithson et al. 2012). The Ballagan is characterized by alternating dolostones
(“cementstones”), paleosols, sandstones, and siltstones. The last of these have
been identified as an important facies for preserving fossils both in the Ballagan
and elsewhere in the Carboniferous (Bennett et al. 2016; Kearsey et al. 2016).
The Ballagan has produced isolated (macro)fossils in the past (Clack 2002) but
its full fossiliferous potential has only recently been realized recently via the
TW:eed project (tetrapods.org), which has also increased our sedimentological
and palaeoenvironmental knowledge of the formation (Smithson et al. 2012,
The entire Ballagan Formation is exposed at Burnmouth, Scotland (Fig.1), as just over 500m of vertically dipping beds. Most of the tetrapod fossils at Burnmouth occur from 332-383m, though there are isolated bones known from lower horizons (see Fig.6a in Clack et al., 2016). The greatest concentration comes from a ~50cm span within a highly-sampled 1m interval at 340.5m originally discovered by TR Smithson. This 1m interval contains the Ossirarus and Aytonerpeton Beds (Clack et al. 2016) and is here designated the Tetrapod Interval Metre, or TIM (see Fig.2).

The Ballagan Formation comprises ten facies and three facies associations, each of which occur throughout the formation; 1) fluvial facies association; 2) overbank facies association; and 3) saline-hypersaline lake facies association (Bennett et al. 2016). The sandy siltstone facies occurs within the overbank facies association, and is characterised as matrix-supported, ungraded siltstones, with millimetre-sized siltstone and very fine sandstone lithic clasts. This facies is a key characteristic of the Ballagan Formation, with 71 beds reported from the Burnmouth succession. Beds are randomly distributed through the succession, laterally variable, and range in thickness from 0.2 to 140cm (Bennett et al., 2016). 71% of beds overlie palaeosols or desiccation cracks and often occur in stacked sequences with palaeosols. The palaeosols are rooted red, green or gley siltstones and only rarely contain small carbonate nodules (Kearsey et al. 2016). They represent a range of floodplain environments...
including woodland (Vertisols), scrubby vegetation (Entisols, Inceptisols) and saline marshes (gleyed Inceptisols).

The sandy siltstone beds are interpreted to have formed as a cohesive flow resulting from monsoonal-type flood events, picking up sediment clasts and fossil material from desiccated floodplain lakes and vegetated ground as the flood travelled (Bennett et al. 2016). The beds either deposited material into depressions on a dry vegetated floodplain, or into existing floodplain lakes or pools. The high degree of vertebrate and invertebrate fossil articulation within these units indicates the local origin and minimal transport of the fossil components (Bennett et al. 2016).

MATERIAL AND METHODS

Collection and preparation

A detailed sedimentary log of the section in the cliff exposure was drawn in October 2012 and enhanced by additional observations gathered from summer fieldwork in 2013-2016. Samples were taken approximately every 20cm through the studied interval and their fossil content was examined under a Leica binocular microscope at the University of Leicester. Initial fossil observations from all sandy siltstone beds identified at Burnmouth (including those in this interval) are recorded in Bennett et al. (2016, Appendix 1). To obtain a high-resolution sedimentological and palaeontological data set, the TIM was sampled by removing a 1mx1mx30cm block from the foreshore. This was recovered in June 2013, with permission from Crown Estate Scotland and Scotland National
Heritage. This sample, was divided into four equal units 25x25x30cm long-wide-deep, named A-D from left to right and 1-3 from top to bottom. From here the fossil content was recorded. Macrofossil material presented here includes samples collected from the TIM by SP Wood in 2007, TR Smithson in 2006-2008, and by the TW:eed team in 2012-2013, and material prepared from the metre square block in 2014-2015 by SPW, TRS, JAC, and BKAO. Much of this collecting was done approximately 18m laterally seaward from the meter square.

The fossils were initially revealed using a 200gm hammer and cold chisel. These were then prepared using a mounted tungsten-carbide needle, under binocular microscopes with up to x50 objectives. Dilute Paraloid B72 was used as a specimen consolidant during preparation. Specimen photography was carried out using a Panasonic Lumix DMC-LZ5 and a Sony Cyber-shot DSC-RX100 III digital camera and a Dino-Lite Pro/Pro2 AD4000 AM4000 digital microscope. Image processing was carried out in Adobe Photoshop. Select specimens were CT scanned using a Nikon Metrology XT H 225 ST High Resolution CT Scanner at the University of Cambridge Biotomography Center. The following parameters were used: isotropic voxel size 0.0609 mm, 1,080 projections, no filter, X-ray 120 kV, 125 µ A, 1,789 slices, 1,000 milliseconds per slice exposure, 24 dB gain. 3-D sectioning was carried out in Mimics Innovation Suite ((biomedical.materialise.com/mimics; Materialise, Leuven, Belgium). Additional rendering and manipulation of objects was done in Meshlab (Visual Computing Lab- ISTI-CNR).
Micopaleontological analysis

Three beds from the TIM were selected for micopaleontological processing from three different facies (Fig.3): 1) palaeosol facies, a gleyed Inceptisol that occurs just below the main fossil-bearing unit; 2) sandy siltstone facies, within the tetrapod-bearing unit; 3) laminated grey siltstone facies, with desiccation cracks containing sandy siltstone, above the tetrapod-bearing unit. All three facies are part of the overbank facies association (Bennett et al., 2016). Approximately 15-20g of each sample was processed overnight in a 5% solution of hydrogen peroxide, then wet sieved at 1000, 425, 250, 125, 65μm fractions and oven dried at 40°C. All fossil specimens present were picked from the 1000, 425, 250 and 125μm fractions and total counts recorded (Otoo et al. 2018, Micropalaeontology data). Three standard-sized, polished thin sections (20μm thick) of these beds were examined on a Leica petrographic microscope at the University of Leicester.

Faunal analyses

Sallan and Coates (2010) compiled a large dataset of gnathostome vertebrate fossil occurrences from the Middle Devonian (Givetian stage) through to the end of the Early Carboniferous (Serpukhovian stage). This dataset was updated to include new data from the Burnmouth TIM fauna (named in the analyses as “Burnmouth-TIM”), Blue Beach (Brazeau 2005; Mansky and Lucas 2013; Anderson et al. 2015) and Mill Hole (Carpenter et al. 2014), as well as revisions of rhizodont taxonomy (Johanson and Ahlberg 2001; Jeffery 2006).
Localities were characterized as marine, marginal, or non-marine, with more specific environmental categories based on descriptions in the original dataset and literature. Both raw diversity (number of species in each taxonomic group) and relative diversity (taxon diversity relative to locality diversity) were analyzed. This accounted for the effects of assemblage size and provided comparability with the Sallan and Coates (2010) analyses.

All faunal analyses were run in PAST (Hammer 2015). In each analysis sites were not grouped *a priori* by stage or environment. Correspondence analysis (CA) was used in order to recover any time-environment faunal associations and investigate which taxa were driving similarities and differences between faunas. Diversity datasets are presented in the supplementary information (Otoo et al. 2018, Raw diversity dataset, Relative diversity dataset). Non-parametric multidimensional scaling (NMDS) was used to characterize faunal similarity quantitatively and graphically using Bray-Curtis distance, a faunal similarity metric Bray-Curtis distance (Bray and Curtis 1957).

SYSTEMATIC PALAEONTOLOGY

Macrofossil distribution

The greatest portion of macrofossils from the TIM come from section C (see Fig.2). The microfossil assemblage of Sample 3 (see Microfossil Results) occurs within the same lithology as the TIM and has similar microfossil content to the TIM macrofossil content. However, macrofossils have been found elsewhere within the TIM, including the paleosol/Ossirarus bed (a lungfish body fossil) and the laminated grey siltstone (eurypterid fossils, Smithson et al. 2012).

Faunal list

A full list of vertebrate taxa represented within the TIM is presented in Table 1. The following taxa and material have been described elsewhere and will not be included here: Aytonerepeton microps (Clack et al. 2016), Diploradus austiumensis (Clack et al. 2016), Ballagadus rossi (Smithson et al. 2015). The indeterminate tetrapod material is figured in the supplement to Clack et al. 2016 and, based on tooth morphology, likely represents at least two different taxa but is not described here.

Class CHONDRICHTHYES Huxley 1880
Family GYRACANTHIDAE sensu Warren et al. 2000
Gyracanthidae indet.
**Material:** Numerous fin spines, as well as UMZC 2017.2.584 and UMZC 2017.2.582, two scapulocoracoids (Fig.4).

**Diagnosis:** Large, broad-based paired fin spines having a distinct longitudinal curvature; fin spines inserted deep into the body, their exserted portions with ornament ridges oblique to the long axis of the spine. Fin spine ridges bearing tubercular ornament. Large triangular scapulocoracoid forms part of pectoral girdle.

**Description:** The spines thus far collected from the TIM have the distinctive curvature of gyracanthid pectoral spines, as opposed to the straightness of spines from other parts of the body (Warren et al. 2000). Consistent with Warren et al.'s diagnosis (see above) at least twenty oblique ridges intersect the boundary of the insertion area. Unlike *Gyracanthides murrayi* (Warren et al. 2000), the insertion area is smooth to gently striated. Tubercles on the oblique ridges are triangular with a median ridge. Oblique ridges closest to the leading edge of spine insertion have two tubercles each; moving towards the trailing edge, this number increases to a maximum of approximately 13 before decreasing to six or less. Unlike *G. murrayi*, the insertion area is gently striated as opposed to tuberculated. UMZC 2017.2.480 preserves distal portion of a likely pectoral spine. Tubercles of oblique ridges absent or indistinct, perhaps as a result of wear and/or abrasion.
The scapulocoracoids are more complete than those of *Gyracanthides murrayi* figured by Warren et al. (2000), though they are smaller in size. They have the same ridged bone texture and triangular shape, with the proximal portion showing a distinct tapering.

**Remarks:** Gyracanth spines are common in Devonian-Carboniferous nonmarine settings (Turner *et al.* 2005; Snyder 2011). Turner *et al.* (2005) referred a number of Early Carboniferous records of *Gyracanthus* to *Gyracanthides*. However, both genera have been identified in Tournaisian collections from Blue Beach (Mansky and Lucas 2013). The two scapulocoracoids are identifiable as gyracanth on the basis of the presence of ridges and striae on both the internal and external surfaces (pers. obs. TR Smithson) as well as their resemblance to the scapulocoracoids of *Gyracanthides* (Turner *et al.* 2005), though they are not diagnostic to the generic level. The gyracanth material is thus here only referred to *Gyracanthidae* indet.

**Superorder HOLOCEPHALI** Bonaparte, 1832-1841

**Order MENASPIFORMES?** Obruchev 1953 cf. Stahl 1999

*Menaspiformes* indet.

**Material:** Two specimens, UMZC 2017.2.583 and UMZC 2017.2.526, each of one spine (Fig.4).
**Diagnosis:** Strongly asymmetric tuberculated dermal spine dividing into two major prongs, each subtriangular; larger, leading prong with prominent elliptic tubercles distributed distally, long axes oriented towards likely leading edge.

**Description:** The leading spine is slightly convex along the presumed leading edge. The trailing spine is straight and subconical. The tuberculated surfaces are characteristic of menaspiform mandibular spines, most clearly described and depicted by Patterson (1965). The spines most closely resemble the mandibular spines of *Deltotoptichius* (Fig. 28C in Patterson 1965). However, they lack the spines on the distal tip. The broken area of UMZC 2017.2.583 is more completely preserved in UMZC 2017.2.526.

**Remarks:** Menaspiform holocephalans are a Permo-Carboniferous group known from, among other places, the Visean of Scotland and Illinois (Stahl 1999). The taxonomy of the group is in need of revision. The specimens are not diagnostic to a lower level, and lack associated toothplates to aid identification. If correctly identified as menaspiform, then these mandibular represent the earliest occurrence of the group.

Class OSTEICHTHYES Huxley 1880

Subclass SARCOPTERYGII Romer 1955

Order DIPNOI Müller 1845

*cf.* *Uronemus splendens* Traquair 1873
**Material:** UMZC 2017.2.588, a single isolated tooth plate (Fig.5A).

**Diagnosis:** Tooth plate with linearly arranged conical cusps, elongate ovoid base.

**Description:** UMZC 2017.2.588 is approximately 0.5cm in length. There are three conical cusps arranged linearly along one side of an ovoid base.

**Remarks:** Specimen UMZC 2017.2.588 strongly resembles the vomerine tooth plate of *Uronemus splendens* as described and figured by Smith et al. (Fig.20 in Smith et al. 1987) in both size and morphology, differing chiefly in having three cusps per ridge instead of four. However, recent study of Tournaisian lungfish has found similar tooth plates in multiple taxa (TR Smithson, unpublished data), and it is not clear whether the marginal toothplates of *Uronemus* are unique or general.

*cf.* *Ctenodus roberti* Smithson et al. 2015

**Material:** A (partial?) pterygoid tooth plate only visible in CT scan (Fig.5C) from UMZC 2015.46b.

**Diagnosis:** Oval tooth plate with six tooth ridges without individual teeth.
**Description:** Oval-shaped tooth plate with six or seven slightly diverging tooth ridges lacking teeth. The tooth plate is flat, gently convex along the anterior edge and straight along the medial side. It is incomplete medially and posteriorly.

Maximum length 4 mm, length to width ratio 3:1.

**Remarks:** This tooth plate resembles the holotype for *Ctenodus roberti* (Fig.3D and Fig.4H in Smithson et al. 2015). It is approximately seven times smaller than the *C. roberti* holotype. It is possible that it is incomplete laterally, but even allowing for this, it is much smaller than the *C. roberti* holotype.

*cf. Ballagadus caustrimi* Smithson et al. 2015

**Material:** An incomplete skeleton represented by disarticulated cranial dermal bones, articulated ribs and fin ray supports, two sets of cranial dermal bones (Fig.6) together with two (partial?) pterygoid tooth plates only visible in CT scan (Fig.5E, F) from 2015.46b.

**Diagnosis:** Toothplate with four tooth ridges. More complete specimen (Fig.5F) shows two teeth on the pterygoid ridge and four or five on the other ridges. Ridge angle approximately 100-120 degrees.

**Description:** Both tooth plates are incomplete and appear to be lacking at least one tooth ridge. The largest specimen is c 5 mm long, the other is 3 mm long. As preserved, the length to width ratio is c 2.5:1. The largest specimen has four
intact tooth ridges with the remains of a fifth. The smaller specimen has four partial tooth ridges. The tooth ridge angle on the largest specimen is c 60° and c 90° on the smaller specimen.

The skeleton (Fig. 6C-H) also appears to belong to *B. castrimi*. The overall body length of the animal represented is approximately 8-9 cm, much smaller than *Ctenodus* spp. Fin supports are present dorsally and ventrally. Unlike *Xylognathus macrustenus*, the dorsal and anal fins do not seem to be separate. The head plates resemble those of a lungfish recovered from the surface of the palaeosol below the *Ossirarus* bed (Fig. 6A, B). While not much morphology is visible, the tail fin appears to be continuous dorsally and ventrally. The headplates are thick and subcircular, with very fine small tubercles.

**Remarks**: The tooth plates in the CT scan are similar to those of *Ballagadus castrimi* (which is in turn highly similar to *B. rossi*), particularly if they are assumed to be damaged. These tooth plates are half the size of the *B. castrimi* tooth plate figured by Smithson *et al.* (2015), but are similar in size to those present on the referred specimen UMZC 2014.1.6a collected by Stan Wood (p. 43 in Smithson *et al.* 2015). The skeleton (Fig. 6C-H) represents an animal with an overall length of approximately 8-9 cm, one of the smallest lungfish from the Ballagan Formation, and is probably similar in size to *Coccovedus celatus* (Smithson *et al.* 2015). Dorsoventral continuity of the tail fin has been considered
characteristic of post-Devonian lungfish in contrast to Devonian forms, but has recently been recognized in Late Devonian lungfish (Clack et al. in press).

cf. *Xylognathus macrustenus* Smithson *et al.* 2015

**Material:** The posterior portion of a pterygoid toothplate only visible in CT scan (Fig.5H) from UMZC 2015.46b.

**Diagnosis:** Narrow, flattened toothplate with two prominent teeth and (possibly) two more smaller teeth.

**Description:** This toothplate is clearly incomplete, and corresponds to the posterior portion of the *Xylognathus* tooth plate. It is flat and slightly curved along the long axis at an approximately 90 degree angle. Apart from damage/wear to the specimen, the surface appears to be smooth and lacking individual denticles.

**Remarks:** While the anterior portion of the toothplate is missing, the remaining posterior portion is a strong match for *Xylognathus*, particularly in preserving the distinctive two rows of teeth. It is also of comparable size to the pterygoid toothplate for *Xylognathus* as reconstructed by Smithson *et al.* (2015).

Order RHIZODONTIDA Andrews and Westoll, 1970

cf. *Strepsodus sauroides* Binney 1841 sensu Jeffery 2006
**Material:** Incomplete and fragmentary skull bones, scales, a series of articulated vertebrae, numerous partial and complete cleithra and clavicles (Fig. 7).

**Diagnosis:** Cleithrum with anastomosing ridged ornament. Clavicle with tuberculate ornament and rounded anterior tip. Simple, cylindrical vertebrae. Subcircular scales with ridged exterior surface and central boss in internal view.

**Description:** Cleithra range from almost complete specimens to fragments. When preserved on the cleithra, the dorsal laminae all have a slight ridge running down the external ascending blade with a slight prominence at the top (Z Johanson pers comm.) This不同iates the cleithrum of *Strepsodus* from the cleithrum of *Screbinodus* (Andrews 1985).

The vertebrae are articulated and exposed in probable ventral or ventrolateral view. The vertebrae (Fig. 7A) are smooth, massive, and broad-waisted. They are extremely similar to the *Strepsodus* vertebrae figured by Andrews and Westoll (1970, see Plate XIIIa). This morphology contrasts with the more pleisiomorphic rhachitimous vertebrae of more basal rhizodonts such as *Barameda* (Garvey et al. 2005), and they are much more robust than those of *Hongyu* (Zhu et al. 2017).

**Remarks:** While rhizodont cleithra often preserve due to their robustness, even in juveniles (Andrews and Westoll 1970; Andrews 1985; Davis et al. 2001), they
are highly variable within species due to age and individual variation. The dermal bone fragments are united by having either a tuberculate or ridged ornament. The quality of the preservation of the ornament varies but the ornamentation patterns are consistent. This suggest that the varied material can be assigned to *Strepsodus* despite being isolated.

aff. *Archichthys portlocki* sensu Jeffery 2006

**Material:** UMZC 2017.2.465 and UMZC 2017.2.458 from Burnmouth, UMZC 2017.2.551 and UMZC 2017.12.2 from Coldstream (Fig.8).

**Diagnosis:** Recurved, robust teeth with short, anastomosing striae. Striae form a ‘collar’ around the circumference of the tooth and are absent from the tip. 12 basal plications.

**Possible synonyms:** “cf. *Archichthys portlocki*”, GLAHM 152115, GLAHM 152121, GLAHM 152252 (Carpenter *et al.* 2014); “*Archichthys portlocki*” YPM VPPU 019334 and NSM008GF040.063 (Carpenter *et al.* 2015); “*Archichthys portlocki*” NBMG 15799, 15818, 19972 (Ó Gogáin *et al.* 2016).

**Description:** The teeth include both insolated singletons, teeth attached to jaw fragments, and a possible vomer, UMZC 2017.2.458. UMZC 2017.2.458 is heavily damaged, but the bone is approximately teardrop-shaped in occlusal
view. A wall of bone brackets approximately half the circumference of the tooth and extends straight posteriorly.

The teeth are circular to oval in cross-section. Where preserved, the bases appear to have 12 plications. The teeth vary in size, suggesting that the striation pattern is present on teeth across the jaw. None of the teeth preserve the sigmoid tip characteristic of *Strepsodus* dentary fangs. Apart from the striation pattern they are otherwise like the teeth of *Archichthys*, robust as opposed to the slender teeth of *Letognathus* which shares a similar striation pattern (Brazeau 2005). The presence of striae distinguishes them from the smooth teeth of *Rhizodus* and *Screbinodus* (Andrews 1985). The striation pattern differs from that of *Strepsodus*, which is composed of larger parallel striae (Jeffery 2006).

**Remarks:** These teeth are difficult to distinguish from those of *Archichthys* given the similarity in striation pattern, and teeth from the Ballagan on the Isle of Bute have been assigned to *Archichthys* previously (Carpenter *et al.* 2014). The situation is not helped by the current limits of reliably-assigned *Archichthys* material (Jeffery 2006). However, the presence of these teeth in the TIM, elsewhere at Burnmouth, and at Coldstream, another Ballagan locality (see Smithson *et al.* 2012 for more locality information) suggests that genuine variation is being sampled. Given the confounding effects of tooth and scale taxonomy on our understanding of rhizodonts (Andrews 1985; Jeffery 2006) and the state of ongoing work we are hesitant to erect a new taxon at this time.
Superclass TETRAPODA Goodrich 1930

aff. Pederpes Clack 2002 and Whatcheeria Lombard and Bolt 1995

Material: UMZC 2016.8, two incomplete possible frontal and prefrontal bones, possibly with partial nasal (Fig.9A-B), (Fig.9A, B, Fig.F, G in Clack et al. 2016), UMZC 2016.9, partial maxilla with teeth, digit bones, a radius, and a cleithrum.

Diagnosis: Anterior margin of orbit straight. Ornament on skull roof bones subcircular at the center of the bone and more ovoid toward the margin. Teeth anteroposteriorly compressed in cross section. Maxilla and cleithrum with fine ornament texture.

Description: The ornament of the frontals/prefrontals contrasts with that of Pederpes, which has a pattern of pits with ridges and grooves; ornament in Whatcheeria is limited to very light pitting. It is not clear whether the orbital margin is present or the edge is broken. If the former, the orbital margin is mostly straight as in Whatcheeria and Pederpes in creating a mostly straight anterior margin to the orbit, though unlike these taxa that region is not thickened or raised into a lip.

The stem of the cleithrum is narrow and straight, more similar to that of Whatcheeria than Pederpes. However, the dorsal expansion, though incomplete, does not seem to have the characteristic notch of Whatcheeria. Among the jaw
material (eg. Fig.9D), there are both a ‘large’ size fraction (approx. 9cm) and a
‘small’ size fraction (approx. 1cm). However, all of these preserve teeth with the
same anteroposteriorly compressed cross-section. The toe bones are longer
than broad, are clearly waisted, asymmetrical, and strongly concave on the flexor
surface. In this they resemble the proximal foot bones of Pederpes but contrast
with those of Whatcheeria, which are much flatter and broader than long (BKA
Otoo pers. obsv.).

Remarks: The skull bones are most like those of Pederpes and Whatcheeria,
with the apparently straight anterior orbit margin the strongest similarity. This
feature is currently unknown outside of these taxa. However, confirmation must
await more complete specimens. The other specimens are not associated with
the skull bones, but resemble Whatcheeria/Pederpes in some respects, contrast
with the colosteid-like morphology of Aytonerpeton and are too large to be
assigned to Diploradus. The jaw material is particularly interesting— the distinctive
tooth morphology seems to be unique, and allows for the unification of
specimens of multiple sizes. UMZC 2014.14, a maxilla collected from Heads of
Ayr by JA Clack in 2014 (see Clack et al., 2016, supplementary data), appears to
have a similar tooth morphology. Though there is not sufficient material for a
formal diagnosis, we suspect this represents a new taxon.

Material: UMZC 2016.7, a partial skull table, and UMZC 2016.6b, a parasphenoid (Fig.10).

Diagnosis: Tetrapod skull table with colosteid-type dermal ornament, curved posterior orbital margin, and circular parietal opening without thickened margin. Parapshenoid with short dorsum sellae and extensively denticulated posterior plate.

Description: The skull table is exposed in internal view (Fig.10A). The pattern of dermal ornament is extremely similar to that of Aytonerpeton, if not identical. Exposures of the lateral line are visible on the postfrontals and postorbital. The pattern of lateral line exposure is nearly identical to that of Greererpeton in the same region (Smithson 1982)]. The parietal opening is relatively much larger.

A lower-resolution image of the parapshenoid was included in the supplement to Clack et al. 2016. In Fig.10D-H the sculpturing on the ventral surface of the cultriform process and denticulation on the posterior plate are more clearly visible. Clack et al. (2016) pointed out the short dorsum sellae, comparing it to later temnospondyls, as well as noting the unique extent of posterior plate denticulation. The morphology of the parasphenoid contrasts strongly with that of Greererpeton, having a broader posterior plate and a relatively shorter, broader cultriform process. This is suggestive of a broader, less elongate skull.
**Remarks:** The size of both the parasphenoid and skull table are consistent with them belonging to *Aytonerpeton*. The proportions of the parasphenoid are consistent with the skull proportions of *Aytonerpeton*, which has a much shorter skull than *Greererpeton*. The colosteid-like lateral line exposure on the skull roof and the great similarity in dermal ornament to *Aytonerpeton* suggest that it belongs to that taxon. However, definitive assignment must await the discovery of more complete *Aytonerpeton* specimens against which these can be compared.

Class **ACTINOPTERYGII** Cope 1887

*Actinopterygii* indet.

**Material:** Isolated scales, rarer squamation, an isolated dermal bone, partial skull roof (Fig.11).

**Diagnosis:** Ornament-covered scales with either diamond shape or elongate trapezoidal shape. Diamond-shaped scales have anterodorsal process and dorsal peg. Dermal bones ornamented with either smooth elongate tubercles or ridges.

**Description:** Two different actinopterygian scale morphologies are present: smaller, more equidimensional scales that occur as hash (Fig.11A) and larger, dorsoventrally elongate scales that are found in isolation (Fig.11B, C).
former are shiny, whereas specimens of the latter exhibit variable amounts of luster. The morphology of the smaller scales is consistent with their being from the caudal lobe of the tail; the larger scales likely come from the flank.

UMZC 2017.2.717 appears to come from the roof of the skull. It is possible that instead of being a single bone it is a set of paired bones. UMZC 2017.2.668 is a partial skull roof, encompassing parts of the frontals and parietals. The ridged ornament pattern is common to many early actinopterygian genera (Daeschler 2000; Friedman and Blom 2006; Mickle 2011, 2017; Sallan and Coates 2013). The orbital margin is long. The frontals are more than twice as long as the parietals and incomplete anteriorly. The posterior margin of the parietals is indented at the midline. The growth center of the right frontal is visible, marked by the central whorl of ornament. The pits of the supraorbital canal are also discernable, crossing over the center of the right frontal and curving laterally near the parietal. The right parietal shows the three pit lines characteristic of actinopterygians.

Remarks: Though there are actinopterygian fossils present in numerous samples from the TIM, in absolute terms the amount of material is small and mostly consists of scales. The proportions of the frontals and parietals compares favorably with that of Limnomis (Daeschler 2000) and Cuneognathus (Friedman and Blom 2006) from the Famennian, both of which are small-bodied. However, they contrast with Avonichthys from the Tournaisian (Wilson et al. 2018).
Interestingly, the elongation of the frontals relative to the parietals has been considered a derived character. None of the taxa appear to have an indented posterior parietal margin, but such an indent appears in *Rhabdolepis macropterus* from the Permian (Schindler 2018).

The phylogenetic analysis of Blom and Friedman (2006) recovered *Limnomis* and *Cuneognathus* as the sister taxa to *Kentuckia* from the Visean. While there is not enough material for a phylogenetic analysis or taxonomic assignment, the morphology of UMZC 2017.2.668 suggests that it may be part of this Devonian-Carboniferous group.

**RESULTS**

**Sedimentology results**

The fossil-rich interval represents a change in sedimentary environment from permanently wet and marshy conditions to seasonally wet conditions (Clack et al., 2016). The 250cm thick section that contains the TIM illustrates this transition in detail (Fig.2). The lower half of the section (up to 1.4 m height) includes a dolostone bed at the base, which contains ostracods and fish debris and has a brecciated top. This is overlain by a sequence of two 40-50cm thick gleyed Inceptisols (c.f. Kearsey et al. 2016), interbedded with mottled siltstones and sandy siltstones. Desiccation cracks occur in three horizons and are infilled with sandy siltstone. Dolostone nodules are associated with the lower gleyed Inceptisol but are localised, only present in the foreshore. The upper half of the
section contains a complex sequence of fossil-rich sandy siltstones (Fig.2, Detailed Log). Overlying this is a sequence of interbedded laminated grey siltstones and red siltstones (Inceptisols), which have numerous desiccation cracks. At the top of the section a Vertisol red siltstone bed contains green drab root haloes. For full description of paleosol facies, see Kearsey et al. (2016).

The detailed log illustrates the complexity of the fossil-rich interval (Fig.2). At the base is a thin gleyed Inceptisol, the top of which is brecciated and infilled by the overlying sandy siltstone. In thin section this bed is light grey, mottled, with fish scales, plant and ostracod fragments at random orientations within a coarse siltstone matrix (Fig.3, thin section 1). The top of the gleyed Inceptisol has an irregular surface, attributed to erosion by the overlying sandy siltstone bed (Fig.3A). This sandy siltstone bed is laterally variable in thickness and in the metre square section it has desiccation cracks in the centre and at the top indicating that it is a composite of two event beds (Fig.12). This is overlain by a thin laminated grey siltstone bed which is localised to the foreshore (not present in the cliff exposure). Above this is the first tetrapod-bearing bed, containing Ossirarus, an organic-rich, structureless, black sandy siltstone (Fig.3A-B). Overlying this dark sandy siltstone bed are two very fine sandstone beds, with asymmetric ripple lamination, that fine upwards into siltstone. Their thickness in the foreshore is 2-4cm, with thinner beds observed in the cliff section (Fig.2-3).

The second tetrapod-bearing bed, with Aytonerpeton, is a 9cm, structureless, dark grey sandy siltstone. In thin section this bed is characterised by a fine-grained siltstone matrix with sub-millimetre sized clasts of siltstone,
sandstone and bioclasts (Fig.3, thin section 2), while in the metre square section, larger light green siltstone clasts up to 5mm in size are present, which are of a similar lithology to the underlying gleyed Inceptisol. Plant material is abundant and plant fossils are sometimes flow-aligned to produce a wavy fabric. Overlying this bed is a sequence of laminated grey siltstone, with millimetre-thick sandstone lenses. Within this bed is a thin sandy siltstone inter-bed that infills desiccation cracks (Fig.3, thin section 3). Above the laminated bed is another sandy siltstone, containing a green siltstone lens, which is the last sandy siltstone in the sequence. At the top of the detailed log sequence are further laminated grey siltstones, with sparse desiccation cracks.

Microfossil results

The purpose of this microfossil study is to characterise the microfossil assemblages in terms of palaeoenvironments, not to describe the taxonomy of fossil specimens. Each sample has a unique microfossil assemblage (Fig.13). Microfossils are generally well preserved, with minimal wear, abrasion, or cracks observed.

Sample 1- palaeosol. The assemblage is dominated by ostracods, with a minor component of actinopterygian scales, one actinopterygian tooth, one rhizodont scale and plant fragments (Fig.13). The majority of ostracods present are poorly preserved, and the following identified to genus level: *Beyrichiopsis*, *Cavellina*, *Glyptolichvinella*, *Paraparchites*, *Shemonaella*, *Silenites*, *Sulcella* (Fig. 14A-D). This assemblage is typical of the fine-grained clastic lithologies of the Ballagan
Formation (Williams et al. 2005). Of these, Cavellina, Shemonaella and Glyptolichvinella are most numerous within the sample (Otoo et al. 2018, Micropalaeontology data). The 1mm and 425µm fractions have a ratio 1:3 carapaces:single valves, while the 250µm and 125µm fractions contain mostly broken single valves, which is likely an artefact resulting from breakage during sieving. Ostracod juveniles with the full instar size range are present and there is no apparent size sorting of the assemblage. The actinopterygian tooth (Fig.14E) lacks the apical cap but preserves the characteristic cross-hatched surface texture (Carpenter et al. 2011).

Sample 2- sandy siltstone. This sample has the highest number of fossil specimens, at 98.7 fossils/gram, which is twofold more than sample 1 and fivefold more than sample 3 (Otoo et al. 2018, Micropalaeontology data). The assemblage is dominated by plant material (fibrous fragments, likely from plant stems) and rhizodont scale and bone fragments (Fig.13). Rhizodonts are also dominant in the underlying Ossirarus bed that was sieved and picked for microfossils as a pilot study, but not quantitatively picked (Fig.14F-H). The exterior surface of the scales has a fibrous structure, and shiny, silver colour. When broken the interior layers have a range of structural elements characteristic of rhizodonts (Andrews 1985) including sheets of either tubercules and pits or grooves and ridges that interlock together. These structures were identified by the examination of broken rhizodont scale macrofossils from the TIM interval. Also present are indeterminate megaspores (with a smooth external surface, no visible trilete marks; Fig.14I), actinopterygian scales and teeth.
Actinopterygian scales have a rhombic shape with a smooth interior surface with keel, and sculpted exterior surface with transverse ridges or grooves (Fig.14J-K). Small rows of curved unornamented actinopterygian teeth, likely pharyngeal in origin, occur solely within the 125µm fraction (Fig.14M-N). Indeterminate brown or black cuticle occurs within all size fractions, and in sample 3. One fragment of semi-lunate shaped eurypterid scale occurs in the underlying Ossirarus sandy siltstone bed (Fig.14L). Dipnoan and tetrapod microfossils have not been recorded from this bed, because of the difficulties of scale and bone fragment identification and their relative rarity compared to the other vertebrates. The amount of unidentified vertebrate bone and scale material varies per sample (sample 1: 0.5%; sample 2: 3.4%; sample 3: 15.2%; Otoo et al. 2018, Micropalaeontology data).

Sample 3- laminated grey siltstone. The sample has a lower number of specimens in each size fraction compared to the other two samples. Plant fragments, Spirorbis and actinopterygian bone and scale fragments dominate the assemblage (Fig.13). Spirorbis specimens are fragmented and pyritised (Fig.14O). Ostracod fragments, indeterminate cuticle and one rhizodont scale fragment are also present. The125µm fraction contains a higher number of actinopterygian fragments than the other size fractions, consisting of small scale and bone debris.
**Faunal similarity analyses**

In all analyses, Devonian and Carboniferous localities plot on opposite sides of the origin along the x axis (Fig.15-18). The convex hulls for each stage overlap within their respective period, but there is no Devonian/Carboniferous overlap. Attempted analyses using environmental sub-categories proved unsuitable within the constraints of the methods used here and are not included.

**Correspondence Analysis (CA).** When the total dataset is analyzed using correspondence analysis (CA), marine sites generally plot below 0 on the y axis and nonmarine (=marginal + continental) sites plot above 0 (Fig.15). Thus four broad time-setting associations emerge when the entire raw diversity dataset (Fig.15A) is analyzed:

- **Devonian marine:** Onychodontida, Ischnacanthida, Arthrodira, Ptyctodontida, other placoderms
- **Devonian nonmarine:** Phyllolepida, Dipnoi, Acanthodida, Climatiida, Porolepiformes, Osteolepididae, Antiarcha, Tristichopteridae
- **Carboniferous marine:** Elasmobranchii, Actinistia, Symmoriiformes, Holocephali, Actinopterygii
- **Carboniferous nonmarine:** Tetrapoda, Rhizodontida, Megalichthyidae, Gyracanthida

Analysis of the entire relative diversity dataset returns the same broad associations, except acanthodians move to the Carboniferous nonmarine
association (Fig.15B). In both the raw and relative diversity analyses Burnmouth-TIM emerges closest to Mill Hole (Fig.15).

When only the nonmarine sites are analyzed, marginal sites do not plot separately from continental sites in any recognizable pattern, though the relative Devonian/Carboniferous positions are maintained (Fig.16). The Givetian convex hull overlaps less with the Frasnian and Famennian hulls, and the Serpukhovian hull no longer overlaps with the Tournaisian and Visean hulls. Burnmouth-TIM plots roughly equidistant from Mill Hole, Gilmerton, and Niddrie. When raw diversity (Fig.16A) is analyzed the following associations are recovered:

- Devonian: Climatiida, Onychodontida, Acanthodida, Ptyctodontida, Ischnacanthida, Osteolepididae, Arthrodira, Phyllolepida, Actinistia, other placoderms
- Devonian: Dipnoi, Antaircha, Porolepiformes, Tristichopteridae, Phyllolepida
- Carboniferous: Actinopterygii, Megalichthyidae, Symmoriiformes, Elasmobranchi
- Carboniferous: Tetrapoda, Rhizodontida, Gyracanthida, Holocephali

When relative diversity is analyzed (Fig.16B), actinistians join the actinopterygian-megalichthyid-elasmobranch-symmoriiform association, and phyllolepids join the other Devonian association.
Non-parametric multidimensional scaling (NMDS). As with the correspondence analysis results, in the NMDS results Devonian and Carboniferous sites are separated along the x axis (Coordinate 2) and marine and nonmarine sites are separate along the y axis (Coordinate 1) (Fig.17). The relative positions of the individual sites are similar to those in the CA results. In analyses of both the total raw diversity dataset (Fig.17A) and the total relative diversity dataset (Fig.17B), Burnmouth-TIM emerges closest to Mill Hole and nearby Gilmerton, Niddrie, and Dora. When only nonmarine sites are analyzed (Fig.18), as with the CA results the overlap between the Givetian and Serpukhovian hulls and those of the other Devonian and Carboniferous stages, respectively, decreases, more so for raw (Fig.18A) than relative diversity (Fig.18B). Burnmouth-TIM maintains its relative position.

DISCUSSION

Sedimentology interpretation

The depositional environments represented in the 250cm thick section (Fig.2) are diverse. The basal dolostone represents deposition in a saline-alkaline lake, with a marine influx onto the floodplain attributed to storm activity (Bennett et al., 2017). The mottled grey siltstones and green palaeosols represent the deposition within a floodplain lake, then the growth of vegetation. Although periods of desiccation occurred in the lower part of the section, in general the soils were waterlogged and the environment was probably a saline (brackish) marsh (Kearsey et al., 2016). Sandy siltstones were deposited as
cohesive debris flows in flooding events after periods of desiccation (Bennett et al., 2016). In the 40cm thick fossil-rich interval numerous flooding events deposited a sequence of six sandy siltstone beds, some of which overlie desiccation cracks. Lateral variability in the thickness and internal structure of sandy siltstones and very fine sandstones between the cliff and foreshore exposures highlight the localised origin of these deposits.

A modern analogue for the dry/wet alternations seen in the paleosol/sandy siltstone association are mud aggregates in dryland river floodplains (Rust and Nanson, 1989; Wakelin-King and Webb, 2007a,b). However, the climate of the Ballagan Formation is interpreted as tropical, with seasonal monsoonal rainfall (Falcon-Lang 1999; Kearsey et al. 2016). Sandy siltstone deposits have not been reported from other sites in the geological record, and have likely been reported as massive siltstone beds, due to the small size of their clasts. It is difficult to find a modern analogue for these deposits. However, a similar facies occurs in the Early Cretaceous Wessex Formation, Isle of Wight. Here plant debris beds contain a similar fauna and are also interpreted to have formed as debris flow deposits during flooding events, in a tropical climate (Sweetman and Insole 2010). These plant debris beds infill local depressions on the floodplain and also contain sediment clasts and fossils within a siltstone matrix. They differ from sandy siltstones in having larger-sized clasts and fossils (including dinosaur bones), but are otherwise interpreted to have formed by a similar mechanism. Overbank deposits preserve tetrapod fossils at the Famennian site of Red Hill, Pennsylvania, but further study is needed to determine whether the standing
water taphofacies described as “green-grey siltstones with abundant plant material and an occasional occurrence of arthropod and vertebrate remains” (Cressler et al. 2010, pg.117) are analogous to sandy siltstones. This emphasizes the ecological and preservational importance of these environments in the Devonian-Carboniferous.

Between flood episodes, deposition laminated grey siltstones were deposited from suspension within shallow floodplain lakes. After the last sandy siltstone bed lacustrine conditions were re-established, with clastic input of siltstones and thin sand beds, perhaps derived from small sheetfloods into the lake. Inceptisols and large desiccation cracks formed by sub-aerial exposure of the lake and scrubby vegetation growth on the floodplain. Finally, the Vertisol represents the formation of a more established soil, alternatively due to the growth of vegetation under relatively dry, long-lived conditions or a drop in the local water table/infilling of the water body by sediment. Roots recorded from Vertisols in the Ballagan Formation can be up to 1 metre in depth and are interpreted to have been formed by isolated stands of lycopsid trees (Kearsey et al. 2016).

**Micropalaeontology interpretation**

The variation within the microfossil assemblages illustrates the different palaeoenvironment and taphonomic history of each of the beds. Taking into account the breakage of some ostracod specimens during pedogenesis, the ostracod assemblage within the palaeosol is indicative of an autochthonous
deposit due to the presence of juveniles, adults and carapaces (Boomer et al., 2003). The ostracods represent one of the oldest brackish-water floodplain assemblages of this group (Williams et al. 2005, 2006), although solely freshwater genera such as Carbonita that occur in the Visean (Bennett et al. 2012) are absent. A brackish salinity for this bed is consistent with the formation of the gleyed Inceptisol under waterlogged brackish conditions (Kearsey et al., 2016). The habitat of the ostracods may have been a saline marsh or pool on the floodplain.

The dominance of rhizodont microfossils within the Aytonerpeton bed is unusual for Ballagan Formation sandy siltstone beds, in which actinopterygian (macro)fossils are more common than rhizodonts (Bennett et al., 2016). This is the most rhizodont-rich bed observed within the Burnmouth succession to date. The evidence of clasts and flow-aligned plant debris within the bed indicates the transport of all fossil material, although the articulated nature of some specimens (eg. the Aytonerpeton holotype, rhizodont vertebral series, lungfish skeleton) indicates local transportation distances. The original habitat of the fauna within this bed was likely to have been floodplain pools, lakes or land surfaces, where animals may have died in periods of drought. The abundance of plant material within the sandy siltstone beds indicated vegetated floodplains, and rooting structures indicate scrubby vegetation to forested landscapes (Kearsey et al. 2016). Heavy rainfall on the floodplain then transported the fossil and plant material in a mud-supported debris flow, to be deposited within a floodplain lake. The microfossil sample three assemblage represents quieter deposition within a
floodplain lake, affected by desiccation, then subject to one small flood deposit that infilled the desiccation cracks.

In the Carboniferous, many groups of actinopterygians, as well as rhizodonts, have been found in paleoenvironments that span salinity gradients, suggesting that they were euryhaline or brackish-freshwater tolerant at this time (Carpenter et al. 2014, 2015; Ó Gogáin et al. 2016). Early Carboniferous eurypterids are mostly restricted to brackish or freshwater environments (Braddy 2001; Lamsdell and Braddy 2010). A brackish habit was recently confirmed for the Famennian East Greenland tetrapods, long considered to have been part of a freshwater fauna (Goedert et al. 2018). The microfossil content of the three samples is different from that identified from two dolostone beds from the Ballagan Formation on the Isle of Bute, which contain actinopterygians, sarcopterygians, dipnoans, elasmobranchs and non-gyracanth acanthodians (Carpenter et al. 2014). The latter two groups are absent from the TIM, as are Chondrites and Phycosiphon-like burrows, which are common within the dolostones (Bennett et al. 2017). The dolostones are thought to form in saline, coastal lakes subject to periodic influx of marine waters in storms. Burrows are not commonly identified within sandy siltstones in this formation (Bennett et al. 2016), however some fossils do indicate a marine influence. Palaeozoic Spirorbis has been interpreted to be of a marine origin, with a mechanism of larval transport onto coastal floodplains attributed to transport during storms or by tides (Gierlowski-Kordesch and Cassle, 2015). The laminated grey siltstone deposit
was likely formed in a floodplain lake that was influenced by a relatively minor marine input.

**Palaeoecology**

The TIM fauna lived on a vegetated floodplain, centered in and around water bodies. The vertebrates were actinopterygians, holocephalan and gyracanth chondrichthyans, and several kinds of sarcopterygians—rhizodonts, lungfish, and tetrapods (Table 1, Fig.19). Living alongside them were various microinvertebrates, as well as eurypterids and myriapods (Smithson *et al.* 2012). Rhizodonts would have been the apex predators, presumably feeding on other taxa as well as each other. Gyracanths would likely have fed on small prey in the water column, whereas tetrapods were likely demersal faunivores, and possibly semiaquatic as well. Lungfish and holocephalans would have taken a range of prey, probably overlapping in their consumption with the sediment-mining eurypterids. Myriapods lived on land, consuming plants and available organic detritus.

Ecological relationships and a food web for the Ballagan Formation have recently been inferred by Bennett *et al.* (work in progress). That reconstruction includes data from this study and other work, and emphasizes the role of diversifying nonmarine invertebrates in providing a wider base for Carboniferous biotas relative to Devonian ones. Several lines of evidence are consistent with this hypothesis. The diversification of holocephalans (*Sallan et al.* 2011, Richards *et al.* in press) and lungfish (*Smithson et al.* 2015) during the Tournaisian and
Visean has been noted previously. In this context it is worth noting that
toothplates are highly effective in immobilizing elusive, soft-bodied prey through
massive trauma (MI Coates pers. obsv.) and that upon examination multiple
Early Carboniferous holocephalans have previously-neglected marginal dentition.
These suggest that multiple taxa took advantage of an increasing abundance
and diversity of small invertebrate prey. This hypothesis should be explored
further as more fossils are described from the Ballagan and other Tournaisian
deposits.

The consistency in taxonomic identities and numerical proportions in the
micro- and macrofossil samples (Otoo et al. 2018, Micropalaeontology data)
suggests that there is a common signal being captured by both. In particular, the
low number of actinopterygian specimens and large number of rhizodont
specimens probably at least somewhat reflect genuine paucity and abundance
rather than solely preservation bias. Both a single actinopterygian and a single
rhizodont will produce numerous scales with high preservation potential, and the
that this is only the case for the TIM rhizodonts is striking. However, low relative
abundance and diversity of actinopterygians is a common feature of both
Devonian and Carboniferous nonmarine settings (Cressler et al. 2010; Sallan
and Coates 2010; Carpenter et al. 2015; Ó Gogáin et al. 2016); at least through
the early part of the Late Carboniferous, the increasing taxonomic,
morphological, and ecological diversity of actinopterygians is considered to be
largely a brackish and, in particular, marine phenomenon (Sallan and Coates
2010, 2013). More abundant and morphologically diverse actinopterygians have been reported from Blue Beach, where the succession preserves multiple marine incursions (Mansky and Lucas 2013).

The rhizodont occurrences in the TIM fit a broader pattern. Rhizodonts are often represented by two species in Carboniferous localities (Sallan and Coates 2010; Carpenter et al. 2014, 2015; Ó Gogáin et al. 2016). Common associations are *Archichthys/Strepsodus*, and *Screbinodus/Rhizodus*, which has long had a confounding effect on the taxonomy of these genera (Andrews 1985; Jeffery 2006). *Screbinodus* is a small rhizodont, with a length of 1.5-2m; *Strepsodus* and *Rhizodus* are both large rhizodonts, with length estimates of 3-5m and 5-7m, respectively (Jeffery 2012). Given the limited material, size estimates have not been produced for *Archichthys*. The TIM *Strepsodus* material indicates individuals across a range of sizes, the largest likely about 1m in length. This parallels the rhizodonts of the Foulden locality, where Andrews (1985) identified multiple ‘small form’ individuals that she suggested were juveniles of the ‘large form’ that lived elsewhere; all these were later referred to *Strepsodus* by Jeffery (2006). This suggests that size was an important ecological discriminant both within and between rhizodont species.

The small size of the referred lungfish toothplates might appear to support Sallan and Galimberti’s (2015) conclusion of a post-Devonian size reduction. However, 35m above the Devonian/Carboniferous boundary at Burnmouth there
is evidence of both very small and very large lungfish, with very large lungfish also occurring 20m above the TIM (TR Smithson pers. obsv., T Challands unpublished data). There are also gyracanth and rhizodont fossils from outside the TIM that represent larger individuals (BKA Otoo pers obsv.). Therefore, it does not seem likely that organisms experienced concerted size reduction at the extinction or in its aftermath. Further sampling will allow for greater examination of this hypothesis.

**Faunal analyses**

The diversity and taxonomic composition of the Mill Hole fauna is most similar to that of the TIM (Carpenter *et al.* 2014), including at least one shared species of rhizodont (*Strepsodus sauroides*). Mill Hole is also in the Ballagan Formation and roughly coeval with the TIM, and these qualitative observations indicate that these assemblages sample a coherent ‘Ballagan fauna’ (this study, Bennett *et al.* in prep.). However, unlike the TIM, the Mill Hole fauna is derived from a dolostone as opposed to a siltstone. The fauna also contains the elasmobranch *Ageleodus* and the sarcopterygian *Megalichthys* (Carpenter *et al.* 2014); while the actinopterygian material is similar in terms of content- mostly scales and lepidotrichia- it is the most abundant material at the locality (JA Clack pers. obsv.). These details indicate that Mill Hole is taxonomically and geologically sampling a more marine signal than the TIM.

The persistent recovery of a Devonian/Carboniferous split when the entire dataset is analyzed with both CA and NMDS supports the conclusion of Sallan
and Coates (2010). The persistence of this split when the marine sites are removed suggests that the end-Devonian extinction has a diversity signal in both marine and nonmarine settings. However, the losses were not evenly distributed among groups- placoderms account for four of the taxonomic divisions within this dataset and were eliminated completely at the end of the Devonian. The Carboniferous nonmarine gyracanth-rhizodont-tetrapod association recovered by the correspondence analysis was already in place in the Late Devonian, there often accompanied by tristichopterids and lungfish (Cressler et al. 2010), and usually porolepiformes and antiarchs as well. In nonmarine settings the end-Devonian extinction left behind a subsetted fauna, which does not seem to have been greatly disturbed; on either side of the Devonian/Carboniferous boundary rhizodonts are generally represented by the same number of species per locality, and tetrapods greatly increase in diversity relative to the Devonian. The fact that when marine sites are removed holocephalans are associated with gyracanths, rhizodonts, and tetrapods (as is the case in the TIM) as opposed to the more marine-inclined actinopterygians, elasmobranchs, and symmoriiformes is interesting, and may suggest different environmental trajectories for elasmobranchs and holocephalans in the Early Carboniferous.

The TIM and Devonian-Carboniferous faunal assembly

The TIM (vertebrate) fauna is very similar to that of the Famennian Red Hill locality, as are their respective environments- both coastal alluvial plains at low palaeolatitude, though Burnmouth experienced greater rainfall (Retallack et al. 2009; Kearsey et al. 2016). Indeed, the only major vertebrate division not
represented in similar numbers in the TIM as at Red Hill are elasmobranchs (ctenacanths) and placoderms (Cressler et al. 2010). Conversely, holocephalans are absent from Red Hill. The rest of the fauna in both cases is composed of gyracanths, tetrapods, actinopterygians, lungfish, and large predatory sarcopterygians (including rhizodonts). The Famennian vertebrate fauna from East Greenland, famous for *Acanthostega* and *Ichthyostega*, also has a similar composition (Blom et al. 2007). It appears that the TIM is sampling a faunal-environmental association that apparently passed through the end-Devonian extinction without extensive modification.

Much of this fauna is held in common with ‘typical’ Carboniferous coal swamp faunas such as Dora and Niddrie, and floodplain localities such as Gilmerton (Sallan and Coates 2010), their qualitative affinities supported by the results of the faunal analyses. The only characteristic coal swamp taxon missing from the TIM are xenacanths, which first appear in the Visean with *Diplodoselache* at Wardie (this may be superseded by Tournaisian occurrences at Burnmouth, Clack et al., work in progress). It would be interesting to know the extent to which the apparent transition of the more or less Devonian-type floodplain fauna- as seen at Red Hill, East Greenland, and, with minor changes, in the TIM at Burnmouth- to the coal swamp setting represents a physical move inland toward fresher water or if the swamps developed on top of the existing alluvial plains (the two scenarios are not mutually exclusive). With the addition of xenacanths, the key difference between the floodplain faunal association and the
coal swamp association seems to be the increased abundance and diversity of tetrapods (and arthropods), at least in part driven by the increased resources and complexity of terrestrial habitats. But overall much of the nonmarine faunal order in the Carboniferous can be described as diversification of a discernable, preexisting subset of Devonian diversity. This contrasts with marine environments, where, following the extinction of placoderms, actinopterygians and elasmobranchs diversified greatly and added large clades/taxonomic divisions (Coates et al. 2017, 2018; Giles et al. 2017) to create very differently-composed marine faunas compared to those that existed in the Devonian.

Yao et al. (2015) recovered isotope excursions in carbon and nitrogen from which they infer sea level fluctuations with a net drop through the Tournaisian, particularly in the second half of the stage. These changes begin in the mid-Tournaisian and do not stabilize until the early Visean. Further investigation of Tournaisian environments preserved at Burnmouth and elsewhere is needed to determine the environmental and biological impact of these sea level changes. The presence of macrofossil assemblages throughout the Ballagan at Burnmouth starting from very near the Devonian/Carboniferous boundary indicates that floodplain biotas were able to persist through these continued post-extinction environmental changes. The physical and physiological ability to move between different salinity regimes would doubtless have been useful, and this interval may be responsible for encouraging the apparent retention of euryhalinity in many vertebrates throughout the Carboniferous.
It is not obvious to what extent these faunal transitions were accompanied by ecological restructuring. Placoderms were extremely numerous at Red Hill, East Greenland, and other Late Devonian localities and had a diversity of morphologies and feeding ecologies- their extinction at the end of the Devonian must have had ecological impacts, but what those were is currently unclear. Also unknown are the impacts of the increased diversity and terrestrialization of tetrapods and arthropods during the Carboniferous. Future comparative ecological work will be valuable for understanding these issues.

CONCLUSIONS

Despite post-dating the end-Devonian mass extinction by only a few million years, the TIM fauna contains multiple invertebrate and vertebrate taxa across different ecological roles. This assemblage is, aside from the lack of placoderms, very similar to floodplain faunas from the Late Devonian, and closely resembles floodplain and coal swamp faunas from later in the Carboniferous. As in environmental similar Late Devonian assemblages, actinopterygians are low in diversity and abundance, with the assemblage dominated numerically and taxonomically by sarcopterygians. The TIM lacks the placoderms of the Devonian and the tetrapod diversity and xenacanths found in later Carboniferous biotas, but otherwise seems to represent a floodplain faunal association that persisted from the Late Devonian into the Carboniferous. Either ecologically or evolutionarily, this floodplain association may have formed the basis of the coal
swamp faunas that characterize many Carboniferous localities. Insofar as the Carboniferous is marked by a diversification of actinopterygians and elasmobranchs, during the Tournaisian these events may have been happening in different environmental/faunal contexts (Mansky and Lucas 2013).

This study and other work (Mansky and Lucas 2013; Carpenter et al. 2014; Anderson et al. 2015; Clack et al. 2016) have focused primarily on taxonomic and phylogenetic changes through the Devonian/Carboniferous transition and Romer’s Gap. These data are providing a basis for future work linking taxonomy and phylogeny to ecology, which will provide an enhanced perspective on the biological and environmental changes through this important period in Earth history.

Acknowledgements. This work was funded by NERC consortium grants NEJ021067/1 (BGS), NE/J022713/1 (Cambridge), NE/ J020729/1 (Leicester), NE/J020621/1 (NMS), and NE/J021091/1 (Southampton), as well as an Evan Carroll Commager Fellowship for Graduate Study in Paleontology from Amherst College. Most of this research was conducted by BKAO in 2014-2015 for an MPhil project under the supervision of JAC at the University of Cambridge. We thank Anne Brown and Colin MacFadyen at Scottish Natural Heritage for permission to collect at Site of Special Scientific Interest under their care, Paul Bancks from the Edinburgh office of The Crown Estate for permission to collect from the foreshore at Burnmouth, members of the TW:eed Project team for help and support during the excavation of the Burnmouth site, Sarah Finney and the
Department of Earth Sciences, University of Cambridge, for help and guidance and the use of lab facilities during the preparation of the bulk samples recovered from Burnmouth, Keturah Smithson for scanning the specimens and help with preparing the rendered images, Nick Fraser and Stig Walsh (NMS) and Matt Lowe (UMZC) for collections access, and the community of Burnmouth for their support and interest during the TW:eed Project. Yasmin Yonan (University of Leicester) provided the reconstruction of the Burnmouth fauna. Graham Slater provided valuable advice on computational methods. TIK publishes with the permission of the Executive Director, British Geological Survey (NERC).

DATA ARCHIVING STATEMENT

Data for this study, including faunal diversity datasets and micropalaeontological data, are available in the Dryad Digital Repository:


[please note that the data for this paper are not yet published and this temporary link should not be shared without the express permission of the author]
REFERENCES


CARPENTER, D. K., FALCON-LANG, H. J., BENTON, M. J. and HENDERSON,


pectoral fin and vertebral column of the rhizodontid fish Barameda decipiens from the Lower Carboniferous of Australia. *Journal of Vertebrate Paleontology*, 25, 8–18.


MICKLE, K. E. 2011. The Early Actinopterygian Fauna of the Manning Canyon


SALLAN, L. C. and COATES, M. I. 2010. End-Devonian extinction and a


SCHINDLER, T. 2018. Revision of Rhabdolepis macropterus (Bonn, 1829) (Osteichthyes, lower Actinopterygii; Lower Permian, SW Germany). *PalZ*.


WARREN, A., CURRIE, B. P., BURROW, C. J. and TURNER, S. 2000. A


**Figure and table captions**

**FIGURE 1.** Map showing English/Scottish border (A) with the detailed location of the study area at Burnmouth (B). Modified from Kearsey et al. 2016.

**FIGURE 2.** Sedimentary log of the tetrapod-bearing beds at Burnmouth. The 250 cm height log represents the exposure in the foreshore, while the detailed log covers part of the sampled metre square. Fossil symbols illustrated on the metre square relate to those identified in hand specimen and from microfossil samples.

**FIGURE 3.** Tetrapod-bearing bed exposure at Burnmouth. A: Photograph of the cliff section, with position of the *Ossirarus, Aytonerpeton* and microfossil sample beds (1-3). B: The *Ossirarus*-bed exposed in the foreshore at Burnmouth. 1-3: Thin section scans of beds that were processed for microfossils; a palaeosol (1), sandy siltstone (2) and laminated grey siltstone (3). Scale bars 10 cm (A), 5 cm (B) and 5 mm (1-3).

**Table 1.** Summary list of vertebrate taxa from the TIM at Burnmouth. Taxa with material that has been described elsewhere are listed with the appropriate reference.

**FIGURE 4.** Selected chondrichthyan bones. A: UMZC 2017.2.583, menaspiform holocephalan mandibular spine; B: UMZC 2017.2.526, menaspiform holocephalan mandibular spine in multiple views, with lighting changed to make morphology more easily visible; C: UMZC 2017.2.481, fin spine showing insertion area and tubercles on ridges; D: UMZC 2017.2.579, gyracanth spine showing ridges; E: UMZC 2017.2.584, partial scapulocoracoid; F: UMZC 2017.2.582, partial scapulocoracoid All scale bars 1cm.

**FIGURE 5.** Lungfish toothplates with reconstructions. A: UMZC 2017.2.588, vomerine toothplate resembling that of *Uronemus splendens*; B: reconstruction of *Uronemus* dorsal dentition modified from Smith *et al.* 1987; C: CT scan of toothplate assigned to *Ctenodus roberti*; D: reconstruction of *C. roberti* dorsal dentition from Smithson *et al.* 2015; E-F: CT scans of toothplates assigned to *Ballagadus castrimi*; G: reconstruction of *B. castrimi* dorsal dentition from Smithson *et al.* 2015; H: CT scan of partial toothplate assigned to *Xylognathus macrusterus* in multiple views; I: reconstruction of *X. macrusterus* dorsal and ventral dentition. CT scans are from UMZC 2015.46b. Scale bars are 0.8cm in B, 0.5cm in C, E, F, and H, and otherwise 1cm.

**FIGURE 6.** Fossils attributed to *Ballagadus castrimi*. A-B: uncatalogued skull bones from the *Ossirarus* bed; C-G: body fossil from the *Aytonerpeton* bed arranged anterior-posterior from C/D to G/H- upper row: UMZC 2015.46a, b, e; lower row: UMZC 2015.46d, c, e. All scale bars are 1cm.

**FIGURE 7.** Various rhizodont bones assigned to *Strepsodus*. A: UMZC 2017.2.578, vertebrae in articulation; B: UMZC 2017.2.520, scales; C: UMZC 2017.2.447, opercular or subopercular in internal and external views; D-H:
cleithra showing growth series- D: UMZC 2017.2.537; E: UMZC 2017.2.541; F: UMZC 2017.2.571; G: UMZC 2017.2.539; H: UMZC 2017.2.515a; I: UMZC 2017.2.672, smallest cleithrum in internal view; J: UMZC 2017.2.534, partial left clavicle showing dermal ornament; K: UMZC 2017.2.535 partial right clavicle; L: UMZC 2017.2.531, partial left clavicle; M: UMZC 2017.2.532, partial left clavicle; N: UMZC 2017.2.538, partial humeral head; O: UMZC 2017.2.538, radius. All scale bars 1cm.

FIGURE 8. Rhizodont teeth showing unusual striation pattern. A-B: UMZC 2017.2.465, vomer with tooth in medial (A) and occlusal (B) views; C: UMZC 2017.2.465, zoomed-in view of tooth showing striation pattern; B: UMZC 2017.2.458, teeth in ?medial view showing striation pattern; E-F: UMZC 2017.2.551, tooth from Bed 1 at Burnmouth, 383 meters in the section (approximately 45 meters above the TIM); G-H: UMZC 2017.12.2, tooth from the Coldstream locality showing striation pattern. Scale bars are 0.5cm in 8C and 8D, all others 1cm.

FIGURE 9. Selection of TIM 'whatcheeriid' material. A: UMZC 2016.8: paired frontals; B: CT scan of the UMZC 2016.8 frontals- the lower frontal in the CT scan is exposed in the specimen; C: UMZC2017.2.569: cleithrum; D: UMZC 2016.9: left maxilla in multiple views; E: UMZC 2017.2.611, radius in external and internal view; F: UMZC 2017.2.577, digit bone in dorsal and ventral view; G: UMZC 2017.3.576, intercentrum in external (left) and internal (right) views. All scale bars are 1cm.

FIGURE 10. Tetrapod material cf. Aytonerpeton. A: UMZC 2016.7, skull table exposed in interior view with mold of dermal ornament; B-C: CT scan of the same specimen in external (B) and internal (C) views; D-F: UMZC 2016.6b, parashenoid in ventral (D), dorsal (E), right lateral (F), posterior (G) and antero-dorsal (H) views. Anatomical abbreviations: BPP: basipterygoid process; CF: cultriform process; DS: dorsum sellae; FR: frontal; LL: lateral line; PAR: parietal; PF: pineal foramen; PFR: postfrontal; PO: postorbital. All scale bars are 1cm.

FIGURE 11. Overview of actinopterygian material. A: scale hash from 2015.46; B: UMZC 2017.2.719, isolated scale; C: UMZC 2017.2.718, isolated scale; D: UMZC 2017.2.717, dermal bone(s); E: UMZC 2017.2.668, partial skull roof, anterior is at the top of the image; F: interpretive drawing of 2017.2.668 at same scale. Anatomical abbreviations: FR: frontal; PAR: parietal; PPL: parietal pit lines; SOC: supraorbital canal. All scale bars 1cm.

FIGURE 12. CT scan of UMZC 2017.2.483 before preparation, showing the internal stratigraphy of the fossil-rich layer captured by the TIM. This specimen does not come from within the TIM itself, but is laterally adjacent to it. Note the two event beds marked by the fossil concentrations at the top and bottom of the specimen. Scale bar is 1cm.

FIGURE 13. Microfossil assemblages. Percentage counts of total assemblage microfossil counts for three samples: sample 1 (A, palaeosol, n = 777
specimens), sample 2 (B, sandy siltstone, n = 1470) and sample 3 (C, laminated grey siltstone, n = 262). The full data table of counts for all size fractions and microfossils per gram is presented in the supplementary information. Abbreviations: actin. = actinopterygian; indet. = indeterminate; rhizo. = rhizodont.


FIGURE 15. Correspondence analysis results, full dataset, raw diversity (A) and relative diversity (B).

FIGURE 16. Correspondence analysis results, nonmarine sites only, raw diversity (A) and relative diversity (B).

FIGURE 17. Non-parametric multidimensional scaling results, full dataset, raw diversity (A) and relative diversity (B).

FIGURE 18. Non-parametric multidimensional scaling results, nonmarine sites only, raw diversity (A) and relative diversity (B).

FIGURE 19. Reconstruction of the Burnmouth TIM fauna. From left to right: rhizodont, gyracanth, tetrapod, actinopterygian, myriapod (lower), and eurypterid (upper). Artwork by Yasmin Yonan (University of Leicester, 2015).
FIGURE 1. Map showing English/Scottish border (A) with the detailed location of the study area at Burnmouth (B). Modified from Kearsey et al. 2016.

145x265mm (300 x 300 DPI)
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symbols illustrated on the metre square relate to those identified in hand specimen and from microfossil
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238x342mm (300 x 300 DPI)
FIGURE 3. Tetrapod-bearing bed exposure at Burnmouth. A: Photograph of the cliff section, with position of the Ossirarus, Aytonerpeton and microfossil sample beds (1-3). B: The Ossirarus-bed exposed in the foreshore at Burnmouth. 1-3: Thin section scans of beds that were processed for microfossils; a palaeosol (1), sandy siltstone (2) and laminated grey siltstone (3). Scale bars 10 cm (A), 5 cm (B) and 5 mm (1-3).

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108x70mm (300 x 300 DPI)
FIGURE 4. Selected chondrichthyan bones. A: UMZC 2017.2.583, menaspiform holocephalan mandibular spine; B: UMZC 2017.2.526, menaspiform holocephalan mandibular spine in multiple views, with lighting changed to make morphology more easily visible; C: UMZC 2017.2.481, fin spine showing insertion area and tubercles on ridges; D: UMZC 2017.2.579, gyracanth spine showing ridges; E: UMZC 2017.2.584, partial scapulocoracoid; F: UMZC 2017.2.582, partial scapulocoracoid. All scale bars 1 cm.

121x183mm (300 x 300 DPI)
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71x30mm (300 x 300 DPI)
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47x13mm (300 x 300 DPI)
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166x166mm (300 x 300 DPI)
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166x254mm (300 x 300 DPI)
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127x147mm (300 x 300 DPI)
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68x42mm (300 x 300 DPI)
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177x393mm (300 x 300 DPI)
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42x22mm (300 x 300 DPI)
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242x353mm (300 x 300 DPI)
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242x354mm (300 x 300 DPI)
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233x327mm (300 x 300 DPI)
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219x149mm (300 x 300 DPI)
Class ACANTHODII
Family GYRACANTHIDAE
Gyracanthidae indet.

Class CHONDRICHTHYES
Chondrichthyes indet.
Superorder HOLOCEPHALI
Order MENASPIFORMES
aff. Menaspiiformes

Class OSTEICHTHYES
Subclass SARCOPTERYGII
Order RHIZODONTIDA
cf. Strepsodus sauroides
aff. Archichthys portlocki

Order DIPNOI
Dipnoi indet.
Uronemus splendens
Ctenodus roberti
Xylognathus macrastenus
Ballagadus rossi (Smithson et al. 2015)

Superclass TETRAPODA
Aytonerpeton microps (Otoo, Clack, and Smithson in Clack et al., 2016)
Diploradus austiumensis (Clack and Smithson in Clack et al., 2016)
aff. Pederpes and Whatcheeria
Tetrapoda indet. (at least two taxa, Clack et al., 2016)

Class ACTINOPTERYGII Cope 1887
Actinopterygii indet.