



Since January 2020 Elsevier has created a COVID-19 resource centre with free information in English and Mandarin on the novel coronavirus COVID-19. The COVID-19 resource centre is hosted on Elsevier Connect, the company's public news and information website.

Elsevier hereby grants permission to make all its COVID-19-related research that is available on the COVID-19 resource centre - including this research content - immediately available in PubMed Central and other publicly funded repositories, such as the WHO COVID database with rights for unrestricted research re-use and analyses in any form or by any means with acknowledgement of the original source. These permissions are granted for free by Elsevier for as long as the COVID-19 resource centre remains active.

Factors determining human-to-human transmissibility of zoonotic pathogens via contact

Mathilde Richard¹, Sascha Knaut², Philip Lawrence^{3,4},
Alison E Mather⁵, Vincent J Munster⁶, Marcel A Müller⁷,
Derek Smith⁸ and Thijs Kuiken¹



The pandemic potential of zoonotic pathogens lies in their ability to become efficiently transmissible amongst humans. Here, we focus on contact-transmitted pathogens and discuss the factors, at the pathogen, host and environmental levels that promote or hinder their human-to-human transmissibility via the following modes of contact transmission: skin contact, sexual contact, respiratory contact and multiple route contact. Factors common to several modes of transmission were immune evasion, high viral load, low infectious dose, crowding, promiscuity, and co-infections; other factors were specific for a pathogen or mode of contact transmission. The identification of such factors will lead to a better understanding of the requirements for human-to-human spread of pathogens, as well as improving risk assessment of newly emerging pathogens.

Addresses

¹ Department of Viroscience, Postgraduate School Molecular Medicine, Erasmus MC, Rotterdam, The Netherlands

² Work Group Neglected Tropical Diseases, German Primate Center, Leibniz-Institute for Primate Research, Göttingen, Germany

³ Université de Lyon, UMR5149, Laboratoire de Biologie Générale, Université Catholique de Lyon – EPHE, Lyon 69288, France

⁴ Molecular Basis of Viral Pathogenicity, International Centre for Research in Infectiology (CIRI), INSERM U1111 – CNRS UMR5308, Université Lyon 1, Ecole Normale Supérieure de Lyon, Lyon 69007, France

⁵ Department of Veterinary Medicine, University of Cambridge, Cambridge, UK

⁶ Virus Ecology Unit, Laboratory of Virology, Division of Intramural Research, National Institute of Allergy and Infectious Diseases, National Institutes of Health, Rocky Mountain Laboratories, Hamilton, MT, USA

⁷ Institute of Virology, University of Bonn Medical Center, Bonn, Germany

⁸ Department of Zoology, University of Cambridge, Cambridge, UK

Corresponding author: Kuiken, Thijs (t.kuiken@erasmusmc.nl)

Current Opinion in Virology 2017, 22:7–12

This review comes from a themed issue on **Emerging viruses: intraspecies transmission**

Edited by **Ron A.M Fouchier** and **Lin-Fa Wang**

For a complete overview see the [Issue](#) and the [Editorial](#)

Available online 28th November 2016

<http://dx.doi.org/10.1016/j.coviro.2016.11.004>

1879-6257/© 2016 The Author(s). Published by Elsevier B.V. This is an open access article under the CC BY license (<http://creativecommons.org/licenses/by/4.0/>).

Introduction

Infectious diseases are the second leading cause of death worldwide after cardiovascular diseases [1]. More than half of the known pathogens that are able to infect humans are of zoonotic origin [2]. Once a zoonotic pathogen has crossed the species barrier by infecting humans, its success in the human population will depend on whether or not it can acquire the ability of sustained human-to-human (H2H) transmissibility. A better understanding of the factors that determine this ability would help to prevent the emergence or re-emergence of infectious diseases in the human population.

Transmission of infectious pathogens amongst humans can occur via multiple routes: airborne (aerosols and respiratory droplets) route, faecal-oral route, contact route or vector-borne route. In this review, we focused on pathogens that are transmitted via direct or indirect contact as their main or substantial routes of transmission. Pathogens that are mainly transmitted via the faecal-oral and food-borne routes — which also are types of contact transmission — were excluded because they are discussed elsewhere in this issue. Direct contact transmission requires physical contact between an infected person and a susceptible person and the transfer of pathogens via touching, sexual contact, or contact with bodily fluids or lesions. Indirect contact refers to the infection of a susceptible person via a contaminated surface. We divided contact transmission into four modes: skin, sexual, respiratory and multiple. We used the following examples to illustrate these four modes of contact transmission: *Treponema pallidum pertenuis* (TPE) for skin contact transmission, human immunodeficiency virus type 1 (HIV-1) for sexual contact transmission, coronaviruses (CoV) for respiratory contact transmission and Ebola virus for contact transmission via multiple routes. For each of these pathogens and their specific mode of transmission, we identified the factors, at the level of the pathogen, host or environment that promoted or hindered their ability of sustained H2H transmissibility.

Skin contact transmission

The spirochete bacterium *Treponema pallidum* (ssp. *pertenuis*; TPE) causes yaws. Another subspecies (ssp. *pallidum*) that causes syphilis is not further discussed here. Yaws primarily affects the skin, bones and cartilages of children in hot and humid areas of Africa and Asia and the

Pacific region. The main sources of infection are direct contact with skin ulcers.

Although TPE is traditionally considered to exclusively infect humans, it has recently been identified in African nonhuman primates. The fact that human and simian TPE strains share a high degree of genetic and functional similarity suggests that African nonhuman primates may serve as a reservoir for human infection and highlights the potential for zoonotic transmission [3].

TPE has obviously acquired the ability of sustained H2H transmissibility. There are several pathogen factors that may contribute to this ability. The spirochete evades the immune response by antigenic variation and abrogation of opsonizing antibodies [4–6], allowing it to survive permanently in the infected host. It reaches high loads in skin ulcers and the infectious dose is low. Besides entering a new host via cuts or abrasions, the closely related *T. pallidum pallidum* is known to use peptides of the outer membrane to attach to host surface proteins [7] and is reported to penetrate healthy mucous membranes [8]. Host factors favouring TPE transmission include crowded living conditions. High humidity and temperature are environmental factors that increase TPE survival outside the host. Finally, lack of surveillance and inadequate health care favour the persistence and spread of human yaws in affected countries.

Sexual transmission

The retrovirus HIV-1 is the archetypal example of a sexually transmitted human pathogen. Although HIV-1 can be contracted by sexual, percutaneous and perinatal routes, nearly 70% of infections worldwide result from heterosexual intercourse [9]. HIV-1 is the causative agent of Acquired Immune Deficiency Syndrome (AIDS) in humans, characterised by severe depletion of memory CD4⁺ T-lymphocytes early following infection, leading ultimately to immunodeficiency and death due to opportunistic infections and rare diseases [10]. Sexual transmission involves the transfer of virus particles or infected cells present in contaminated genital secretions or blood from an infected person to the mucosa of a susceptible host [11^{*}]. Following transmission, the successfully transmitted founder virus population is established in CD4⁺ cells in mucosa/submucosa, draining lymphatics, gut-associated lymphoid tissue and systemic lymphatic tissues. Viraemia follows and increases exponentially as a result of massive virus replication in gut associated and other peripheral lymphoid tissue [11^{*}].

HIV-1 likely originated from nonhuman primates at some time in the twentieth century. The most genetically similar and related primate lentivirus described to date is the simian immunodeficiency virus (SIV) found in chimpanzees in central Africa (SIVcpz) [12]. The majority of nonhuman primate species appear afflicted with a

single strain of SIV that is mostly non-pathogenic in its natural host.

Important restriction factors by which infected hosts control lentiviral infection are tetherin [13], APOBEC3G [14] and TRIM5 α [15]. The ability of the most prevalent strain of HIV-1, the M strain, to overcome such restriction factors is believed to have been critical to establish an infection in humans and to allow sustained H2H transmission, leading to the current global pandemic [16]. In comparison, the reduced abilities of other HIV-1 strains (N, O and P) and HIV-2 to counteract these restriction factors [17–19] may partly explain why they were not able to spread so effectively within the human population. Other pathogen factors that have contributed to the ‘success’ of HIV-1 M strain as a human pathogen, despite its relatively low infectivity (risk estimate of 1 in 1000 exposures for heterosexual transmission; [9]), include its extraordinary propensity to evolve its genome through recombination and low-fidelity replication, allowing immune and therapeutic escape [20], the nature of its long, ‘latent’, often sub-clinical infection, during which patients can transmit the virus [21], and high viral load. Host factors that favour transmission are the presence of other sexually transmitted diseases [9], as well as promiscuous sexual behaviour.

Respiratory contact transmission

Of the six known human (CoV), severe acute respiratory syndrome CoV (SARS-CoV) [22] and Middle East respiratory syndrome CoV (MERS-CoV) [23] are responsible for high morbidity and mortality in infected individuals. The other four human CoV (HCoV-229E, NL63, OC43, HKU1) have low pathogenicity and are associated with seasonal common colds [24].

The zoonotic origin of four out of six human CoV has been elucidated. HCoV-229E, SARS-CoV and MERS-CoV originate from bats and HCoV-OC43 from bovines, whereas animal ancestors for HCoV-HKU1 and NL63 are still to be found [25^{*}]. The common cold CoV likely emerged a long time ago in the human population, as reflected by a global distribution and a high prevalence in humans [26]. In contrast, intermediate host species such as Himalayan palm civets [27] and dromedaries [28^{**}] likely played a role in the recent introduction of SARS-CoV and MERS-CoV, respectively, in the human population.

SARS-CoV and MERS-CoV are predominantly transmitted via direct H2H contact, droplets and fomites [29–31], and have not (yet) established long-term and sustained H2H transmission. Virus replication occurs mainly in the lower respiratory tract (LRT) in type II pneumocytes and alveolar macrophages [32,33^{**},34]. Replication in the LRT may be explained by the protein expression profile of the respective receptors, the exopeptidases angiotensin-converting

enzyme 2 (ACE2) in case of SARS-CoV [35] and dipeptidyl peptidase 4 (DPP4) for MERS-CoV [36,37]. In addition, antiviral immunity of the epithelium may reduce viral replication in the upper respiratory tract (URT) [38]. For contact transmission of SARS-CoV and MERS-CoV between humans, the quantity of infectious particles seems to be an important factor as high viral loads in patients facilitated H2H transmission [39,40]. Pronounced stability of infectious CoV on surfaces for up to several days [41] could also explain fomite-related transmissions, a phenomenon that may contribute to superspreading events [39,42].

In contrast to SARS-CoV and MERS-CoV, the four common cold CoV are predominantly droplet-transmitted [43] and efficiently H2H transmissible. Virus replication occurs mainly in the central and upper parts of the respiratory tract. For HCoV-229E, this may be explained in part by the abundant expression of its entry receptor, aminopeptidase N, on non-ciliated cells of the bronchial epithelium [44]. However, although HCoV-NL63 uses the same entry receptor as SARS-CoV (ACE2) [45], it replicates mainly in the URT, perhaps because it uses additional attachment factors like heparan sulfate proteoglycans [46], or because of other as yet unknown viral replication-related or immune-related factors.

Comparison of these two groups of CoV suggests that URT replication and droplet transmission (common cold CoV) is more advantageous for sustainable H2H transmission than LRT replication and direct contact transmission (SARS-CoV and MERS-CoV). Replication in the URT, as well as transmission via the respiratory route, are also factors that favour the efficient H2H transmission of human influenza viruses, as compared to zoonotic avian influenza viruses [47].

In conclusion, high expression levels of a suitable receptor molecule in the URT combined with efficient and probably well-balanced viral countermeasures against local immunity may be major pathogen factors for zoonotic CoV to attain successful and sustained H2H transmission, as exemplified by the common cold CoV.

Multiroute transmission

Of the five known Ebola virus species, four are known to cause disease in humans [48]. Infection with Ebola virus causes Ebola virus disease (EVD), which is an acute systemic illness with a high case fatality rate [49].

Of all potential transmission routes, direct contact with patients or bodily fluids from these patients, as well as contact with contaminated surfaces or materials, is considered the most important [50,51]. Ebola virus has indeed been isolated from several bodily fluids such as blood, breast milk and semen of infected patients. In addition, Ebola virus RNA has been detected in sweat, tears, stool, on skin and from vaginal and rectal samples

[52]. During the 2014 outbreak, several researchers speculated about the potential for airborne transmission of Ebola virus [53]. However, the majority of EVD patients in previous outbreaks were infected by contact transmission and all EVD outbreaks, including the 2014 epidemic, have been contained without measures against airborne transmission in the general population. This suggests that extensive airborne transmission is unlikely and of limited epidemiological importance. The 2014 EVD outbreak revealed the potential for Ebola virus to be sexually transmitted. Infectious virus and Ebola virus RNA have been detected in semen from male EVD survivors as long as 70 and 270 days, respectively, after recovery from the initial infection [54]. This detection of infectious Ebola virus and viral RNA months after recovery from EVD highlights the potential for Ebola virus to seed new outbreaks after patients with clinical EVD are no longer present and an area is declared Ebola virus free.

The putative animal reservoirs for Ebola viruses are bats, and zoonotic transmission is thought to occur either by direct contact with bats, or via indirect transmission by contact with bats, or via indirect transmission by contact with other infected wildlife species, such as gorillas, chimpanzees or duikers, which — like humans — are affected by Ebola virus [55].

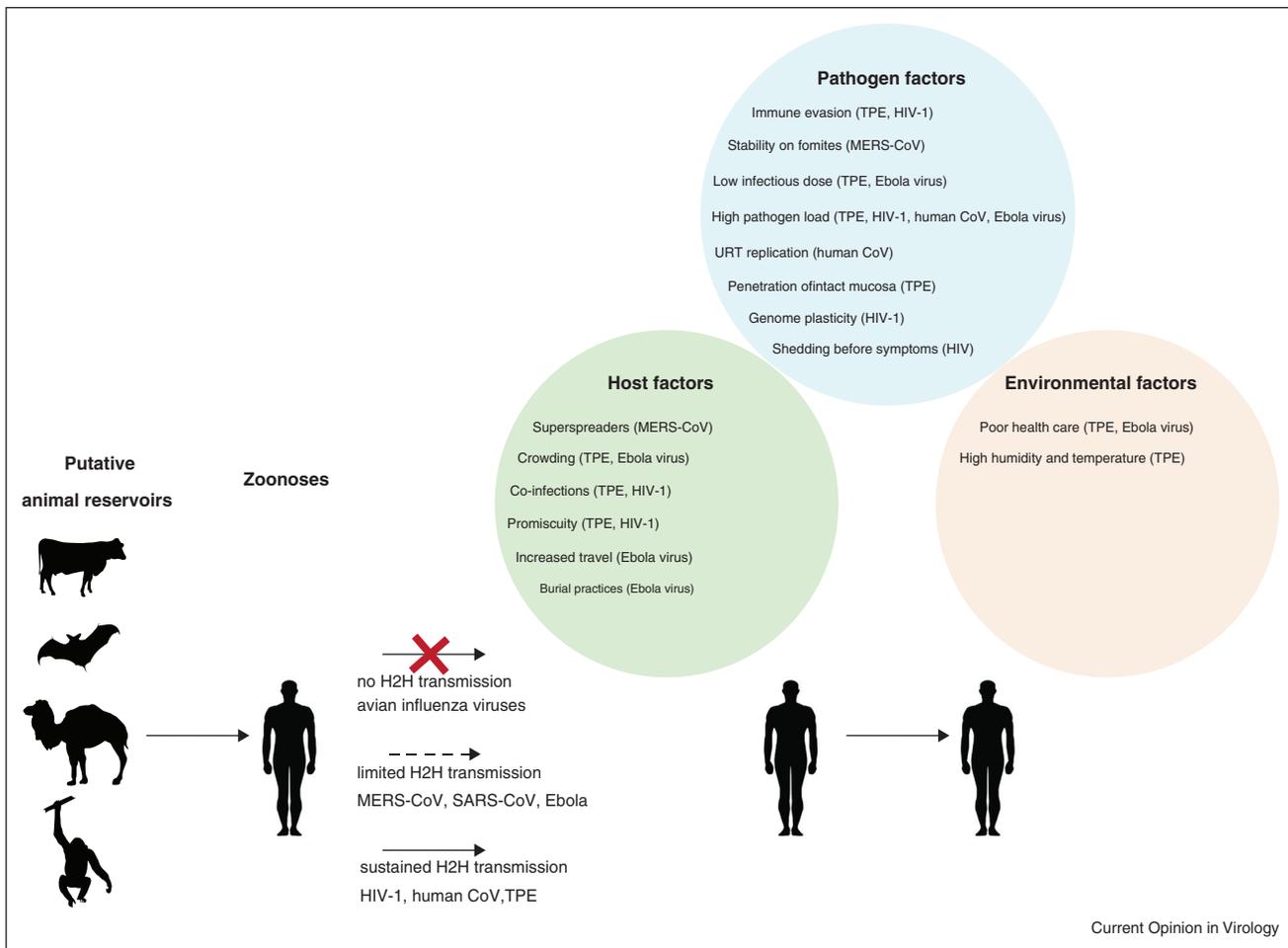
Amongst the pathogen factors that promote H2H transmission of Ebola virus is the high virus load in secreted bodily fluids combined with a very low infectious dose, as low as 10 plaque forming units as measured in experimental infection studies in nonhuman primates [56]. Amongst host factors, ancestral funeral and burial practices of deceased EVD patients, in which levels of Ebola virus remain high after death, have been identified as a major source of human infection [57]. Moreover, the 2014 outbreak of EVD in West Africa, caused by the Zaire Ebola virus, has shown, for the first time, the ability of Ebola virus to cause a long-term large-scale epidemic with sustained H2H transmission [58**]. In addition to EVD cases in Guinea, Sierra Leone and Liberia, travel-associated cases with subsequent nosocomial transmission have been reported in Mali, Nigeria and the United States. The 2014 Ebola virus strains were relatively closely related to viral strains from the previous two Zaire Ebola virus outbreaks in Democratic Republic of Congo, and, although the evolution rate of the genome of the Ebola virus during the 2014 outbreak was higher than the between-outbreak rate, the virus did not change substantially [58**,59**]. The clinical course, for example, incubation time, symptoms and development of the disease, as well as the transmissibility of the virus (R_0 , basic reproductive number) were not different from those in past outbreaks of Ebola virus. Most likely, the unprecedented epidemic of Ebola in 2014 was the result of a combination of human behavioural and societal factors [60]. Firstly, West African countries never had

experienced an EVD outbreak before, other than a single case of Taï Forest ebolavirus infection in the 1980s in Ivory Coast. In addition, Guinea, Sierra Leone and Liberia are amongst the poorest countries in the world, with impaired public health infrastructures. Moreover, compared to previous outbreaks of Ebola virus, the virus was not confined to remote and rural areas and the outbreak spread into large population centres, such as Monrovia, Conakry and Freetown. The spatial connectivity provided by roads and a travelling population allowed for the rapid dissemination of Ebola virus over these three countries, before a targeted international response was initiated.

Concluding remarks

Despite being categorised under one heading, contact-transmitted pathogens may differ substantially in their specific modes of transmission: via skin, via genital mucosa, via respiratory mucosa, or via several of these modes. Nonetheless, several factors were identified that were common amongst at least two modes of transmission. Therefore, it is important to identify both factors promoting H2H transmission that are common amongst contact-transmitted pathogens and factors that are specific for each mode of contact transmission (Figure 1). Common pathogen factors were immune evasion, high viral load, and low infectious dose. Common host factors were

Figure 1



Factors, at the pathogen, host and environmental levels, that promote human-to-human contact transmission of human pathogens of zoonotic or putative zoonotic origin. The transmissibility of pathogens of zoonotic origin determines their pandemic potential. Common factors, as well as specific factors, that promoted the transmissibility via contact amongst humans of the following pathogens via the following routes are described and categorised under pathogen, host and environmental factors: *Treponema pallidum pertenu* for skin contact transmission, human immunodeficiency virus type 1 for sexual contact transmission, coronaviruses for respiratory contact transmission and Ebola virus for contact transmission via multiple routes. The pathogen to which these factors refer is indicated between brackets. **Abbreviations:** H2H: human-to-human; TPE: *Treponema pallidum pertenu*; CoV: coronavirus; MERS-CoV: Middle East respiratory syndrome CoV; SARS-CoV: severe acute respiratory syndrome CoV; HIV-1: human immunodeficiency virus type 1.

crowding, promiscuity, and co-infections. Other factors were specific to one of the modes of transmission and the pathogen described. Specific factors that may be critical for efficient H2H transmission are high viral load in skin lesions for skin contact transmission, promiscuous sexual behaviour for sexual contact transmission, URT replication and a switch from contact to aerosol transmission for respiratory contact transmission and burial practices for the transmission of Ebola virus. Identification of these factors is critical to assess the risk of contact-transmitted zoonotic pathogens gaining efficient H2H transmissibility, and to implement mitigation measures and large scale prevention campaigns in case of outbreaks.

Acknowledgements

The expert workshop was financially supported by European FP7 programme ANTIGONE (ANTicipating the Global Onset of Novel Epidemics, project number 278976). MR's research is partly supported by NIAID/NIH contract HHSN272201400008C. SK's research is partly supported by grants of the German Research Foundation (DFG): KN1097/3-1 and KN1097/4-1. AEM is supported by a Biotechnology and Biological Sciences Research Council (BBSRC) grant BB/M014088/1. VJM is supported by the Division of Intramural Research of the NIAID/NIH. Marcel A Müller is supported by the Zoonoses Anticipation and Preparedness Initiative (ZAPI project; IMI Grant Agreement n° 115760 granted to Christian Drosten).

References and recommended reading

Papers of particular interest, published within the period of review, have been highlighted as:

- of special interest
- of outstanding interest

1. WHO: *The top 10 causes of death*. URL: <http://www.who.int/mediacentre/factsheets/fs310/en/>
2. Jones KE, Patel NG, Levy MA, Storeygard A, Balk D, Gittleman JL, Daszak P: **Global trends in emerging infectious diseases**. *Nature* 2008, **451**:990-993.
3. Knauf S, Liu H, Harper KN: **Treponemal infection in nonhuman primates as possible reservoir for human yaws**. *Emerg Infect Dis* 2013, **19**:2058-2060.
4. LaFond RE, Molini BJ, Van Voorhis WC, Lukehart SA: **Antigenic variation of TrpK V regions abrogates specific antibody binding in syphilis**. *Infect Immun* 2006, **74**:6244-6251.
5. Alderete JF, Baseman JB: **Surface-associated host proteins on virulent *Treponema pallidum***. *Infect Immun* 1979, **26**:1048-1056.
6. Cox DL, Chang P, McDowall AW, Radolf JD: **The outer membrane, not a coat of host proteins, limits antigenicity of virulent *Treponema pallidum***. *Infect Immun* 1992, **60**:1076-1083.
7. Baseman JB, Hayes EC: **Molecular characterization of receptor binding proteins and immunogens of virulent *Treponema pallidum***. *J Exp Med* 1980, **151**:573-586.
8. Riviere GR, Thomas DD, Cobb CM: **In vitro model of *Treponema pallidum* invasiveness**. *Infect Immun* 1989, **57**:2267-2271.
9. Shaw GM, Hunter E: **HIV transmission**. *Cold Spring Harb Perspect Med* 2012:2.
10. Williams KC, Burdo TH: **HIV and SIV infection: the role of cellular restriction and immune responses in viral replication and pathogenesis**. *APMIS* 2009, **117**:400-412.
11. Ronen K, Sharma A, Overbaugh J: **HIV transmission biology: translation for HIV prevention**. *AIDS* 2015, **29**:2219-2227.
In this very complete review, several aspects of the biology of HIV transmission and their implications for existing prevention and control measures are reviewed.
12. Sharp PM, Hahn BH: **Origins of HIV and the AIDS pandemic**. *Cold Spring Harb Perspect Med* 2011, **1** a006841.
13. Neil SJ, Zang T, Bieniasz PD: **Tetherin inhibits retrovirus release and is antagonized by HIV-1 Vpu**. *Nature* 2008, **451**:425-430.
14. Sheehy AM, Gaddis NC, Choi JD, Malim MH: **Isolation of a human gene that inhibits HIV-1 infection and is suppressed by the viral Vif protein**. *Nature* 2002, **418**:646-650.
15. Stremlau M, Owens CM, Perron MJ, Kiessling M, Autissier P, Sodroski J: **The cytoplasmic body component TRIM5alpha restricts HIV-1 infection in Old World monkeys**. *Nature* 2004, **427**:848-853.
16. Sauter D, Schindler M, Specht A, Landford WN, Munch J, Kim KA, Votteler J, Schubert U, Bibollet-Ruche F, Keele BF et al.: **Tetherin-driven adaptation of Vpu and Nef function and the evolution of pandemic and nonpandemic HIV-1 strains**. *Cell Host Microbe* 2009, **6**:409-421.
17. Bush S, Tebit DM: **HIV-1 Group O origin, evolution, pathogenesis, and treatment: unraveling the complexity of an outlier 25 years later**. *AIDS Rev* 2015, **17**:147-158.
18. de Silva TI, Cotten M, Rowland-Jones SL: **HIV-2: the forgotten AIDS virus**. *Trends Microbiol* 2008, **16**:588-595.
19. Sauter D, Hue S, Petit SJ, Plantier JC, Towers GJ, Kirchhoff F, Gupta RK: **HIV-1 Group P is unable to antagonize human tetherin by Vpu, Env or Nef**. *Retrovirology* 2011, **8**:103.
20. Boutwell CL, Rolland MM, Herbeck JT, Mullins JI, Allen TM: **Viral evolution and escape during acute HIV-1 infection**. *J Infect Dis* 2010, **202**(Suppl. 2):S309-S314.
21. Morse SS: **Factors in the emergence of infectious diseases**. *Emerg Infect Dis* 1995, **1**:7-15.
22. Drosten C, Gunther S, Preiser W, van der Werf S, Brodt HR, Becker S, Rabenau H, Panning M, Kolesnikova L, Fouchier RA et al.: **Identification of a novel coronavirus in patients with severe acute respiratory syndrome**. *N Engl J Med* 2003, **348**:1967-1976.
23. Zaki AM, van Boheemen S, Bestebroer TM, Osterhaus AD, Fouchier RA: **Isolation of a novel coronavirus from a man with pneumonia in Saudi Arabia**. *N Engl J Med* 2012, **367**:1814-1820.
24. Lau SK, Woo PC, Yip CC, Tse H, Tsoi HW, Cheng VC, Lee P, Tang BS, Cheung CH, Lee RA et al.: **Coronavirus HKU1 and other coronavirus infections in Hong Kong**. *J Clin Microbiol* 2006, **44**:2063-2071.
25. Drexler JF, Corman VM, Drosten C: **Ecology, evolution and classification of bat coronaviruses in the aftermath of SARS**. *Antiviral Res* 2014, **101**:45-56.
Drexler et al. provide phylogenetic evidence for a zoonotic origin of four out of six human coronaviruses: HCoV-229E, SARS-CoV and MERS-CoV originate from bats, and HCoV-OC43 from bovines.
26. Perlman S, Netland J: **Coronaviruses post-SARS: update on replication and pathogenesis**. *Nat Rev Microbiol* 2009, **7**:439-450.
27. Ge XY, Li JL, Yang XL, Chmura AA, Zhu G, Epstein JH, Mazet JK, Hu B, Zhang W, Peng C et al.: **Isolation and characterization of a bat SARS-like coronavirus that uses the ACE2 receptor**. *Nature* 2013, **503**:535-538.
28. Haagmans BL, Al Dhahiry SH, Reusken CB, Raj VS, Galiano M, Myers R, Godeke GJ, Jonges M, Farag E, Diab A et al.: **Middle East respiratory syndrome coronavirus in dromedary camels: an outbreak investigation**. *Lancet Infect Dis* 2014, **14**:140-145.
This study provides virological and serological evidence of MERS-CoV in camels. The sequences of the viruses obtained from nasal swabs of camels were similar to that of two human isolates from the same farm, suggesting the possibility of a recent outbreak affecting both humans and camels.
29. WHO: *First data on stability and resistance of SARS coronavirus compiled by members of WHO laboratory network*. URL: http://www.who.int/csr/sars/survival_2003_05_04/en/
30. WHO: *SARS (Severe Acute Respiratory Syndrome)*. URL: <http://www.who.int/ith/diseases/sars/en/>

12 Emerging viruses: intraspecies transmission

31. WHO: *Middle East respiratory syndrome coronavirus (MERS-CoV)*. URL: <http://www.who.int/mediacentre/factsheets/mers-cov/en/>
32. Hamming I, Timens W, Bulthuis ML, Lely AT, Navis G, van Goor H: **Tissue distribution of ACE2 protein, the functional receptor for SARS coronavirus. A first step in understanding SARS pathogenesis.** *J Pathol* 2004, **203**:631-637.
33. Meyerholz DK, Lambertz AM, McCray PB Jr: **Dipeptidyl Peptidase 4 distribution in the human respiratory tract: implications for the Middle East Respiratory Syndrome.** *Am J Pathol* 2016, **186**:78-86.
- In this study, the authors studied the localization of the receptor of the MERS-CoV, DPP4, in the human respiratory tract. DPP4 was found preferentially in alveolar regions, in several different cells types. Moreover, authors found that patients with respiratory co-morbidities exhibited an increased expression of DPP4, explaining why pre-existing pulmonary diseases could predispose people to MERS-CoV morbidity and mortality.
34. Ng DL, Al Hosani F, Keating MK, Gerber SI, Jones TL, Metcalfe MG, Tong S, Tao Y, Alami NN, Haynes LM *et al.*: **Clinicopathologic, immunohistochemical, and ultrastructural findings of a fatal case of Middle East respiratory syndrome coronavirus infection in the United Arab Emirates, April 2014.** *Am J Pathol* 2016, **186**:652-658.
35. Li W, Moore MJ, Vasilieva N, Sui J, Wong SK, Berne MA, Somasundaran M, Sullivan JL, Luzuriaga K, Greenough TC *et al.*: **Angiotensin-converting enzyme 2 is a functional receptor for the SARS coronavirus.** *Nature* 2003, **426**:450-454.
36. Raj VS, Mou H, Smits SL, Dekkers DH, Muller MA, Dijkman R, Muth D, Demmers JA, Zaki A, Fouchier RA *et al.*: **Dipeptidyl peptidase 4 is a functional receptor for the emerging human coronavirus-EMC.** *Nature* 2013, **495**:251-254.
37. Bertram S, Glowacka I, Muller MA, Lavender H, Gnirss K, Nehlmeier I, Niemeyer D, He Y, Simmons G, Drosten C *et al.*: **Cleavage and activation of the severe acute respiratory syndrome coronavirus spike protein by human airway trypsin-like protease.** *J Virol* 2011, **85**:13363-13372.
38. Kindler E, Jonsdottir HR, Muth D, Hamming OJ, Hartmann R, Rodriguez R, Geffers R, Fouchier RA, Drosten C, Muller MA *et al.*: **Efficient replication of the novel human betacoronavirus EMC on primary human epithelium highlights its zoonotic potential.** *MBio* 2013, **4**:e00611-e00612.
39. Korea Centers for Disease C: **Prevention Middle East respiratory syndrome coronavirus outbreak in the Republic of Korea, 2015.** *Osong Public Health Res Perspect* 2015, **6**:269-278.
40. Corman VM, Albarak AM, Omrani AS, Albarak MM, Farah ME, Almasri M, Muth D, Sieberg A, Meyer B, Assiri AM *et al.*: **Viral shedding and antibody response in 37 patients with Middle East respiratory syndrome coronavirus infection.** *Clin Infect Dis* 2016, **62**:477-483.
41. van Doremalen N, Bushmaker T, Munster VJ: **Stability of Middle East respiratory syndrome coronavirus (MERS-CoV) under different environmental conditions.** *Euro Surveill* 2013:18.
42. Lee SS, Wong NS: **Probable transmission chains of Middle East respiratory syndrome coronavirus and the multiple generations of secondary infection in South Korea.** *Int J Infect Dis* 2015, **38**:65-67.
43. Callow KA, Parry HF, Sergeant M, Tyrrell DA: **The time course of the immune response to experimental coronavirus infection of man.** *Epidemiol Infect* 1990, **105**:435-446.
44. van der Velden VH, Wierenga-Wolf AF, Adriaansen-Soeting PW, Overbeek SE, Moller GM, Hoogsteden HC, Versnel MA: **Expression of aminopeptidase N and dipeptidyl peptidase IV in the healthy and asthmatic bronchus.** *Clin Exp Allergy* 1998, **28**:110-120.
45. Hofmann H, Pyrc K, van der Hoek L, Geier M, Berkhout B, Pohlmann S: **Human coronavirus NL63 employs the severe acute respiratory syndrome coronavirus receptor for cellular entry.** *Proc Natl Acad Sci U S A* 2005, **102**:7988-7993.
46. Milewska A, Zarebski M, Nowak P, Stozek K, Potempa J, Pyrc K: **Human coronavirus NL63 utilizes heparan sulfate proteoglycans for attachment to target cells.** *J Virol* 2014, **88**:13221-13230.
47. Richard M, Fouchier RA: **Influenza A virus transmission via respiratory aerosols or droplets as it relates to pandemic potential.** *FEMS Microbiol Rev* 2016, **40**:68-85.
48. Kuhn JH, Becker S, Ebihara H, Geisbert TW, Johnson KM, Kawaoka Y, Lipkin WI, Negrodo AI, Netesov SV, Nichol ST *et al.*: **Proposal for a revised taxonomy of the family Filoviridae: classification, names of taxa and viruses, and virus abbreviations.** *Arch Virol* 2010, **155**:2083-2103.
49. Feldmann H, Geisbert TW: **Ebola haemorrhagic fever.** *Lancet* 2011, **377**:849-862.
50. Dowell SF, Mukunu R, Ksiazek TG, Khan AS, Rollin PE, Peters CJ: **Transmission of Ebola hemorrhagic fever: a study of risk factors in family members, Kikwit, Democratic Republic of the Congo, 1995.** *Commission de Lutte contre les Epidemies a Kikwit. J Infect Dis* 1999, **179**(Suppl. 1):S87-S91.
51. Judson S, Prescott J, Munster V: **Understanding ebola virus transmission.** *Viruses* 2015, **7**:511-521.
52. Bausch DG, Towner JS, Dowell SF, Kaducu F, Lukwiya M, Sanchez A, Nichol ST, Ksiazek TG, Rollin PE: **Assessment of the risk of Ebola virus transmission from bodily fluids and fomites.** *J Infect Dis* 2007, **196**(Suppl. 2):S142-S147.
53. Osterholm MT, Moore KA, Kelley NS, Brosseau LM, Wong G, Murphy FA, Peters CJ, LeDuc JW, Russell PK, Van Herp M *et al.*: **Transmission of Ebola viruses: what we know and what we do not know.** *MBio* 2015, **6**:e00137.
54. Uyeki TM, Erickson BR, Brown S, McElroy AK, Cannon D, Gibbons A, Sealy T, Kainulainen MH, Schuh AJ, Kraft CS *et al.*: **Ebola virus persistence in semen of male survivors.** *Clin Infect Dis* 2016, **62**:1552-1555.
- In this study, the semen of five male Ebola virus disease survivors was tested for virus presence after recovery. Viral RNA could be detected in the semen up to 290 days after the onset of the symptoms and virus culture from semen sample was possible until 72 days after illness onset.
55. Olival KJ, Hayman DT: **Filoviruses in bats: current knowledge and future directions.** *Viruses* 2014, **6**:1759-1788.
56. Reed DS, Lackemeyer MG, Garza NL, Sullivan LJ, Nichols DK: **Aerosol exposure to Zaire ebolavirus in three nonhuman primate species: differences in disease course and clinical pathology.** *Microbes Infect* 2011, **13**:930-936.
57. Curran KG, Gibson JJ, Marke D, Caulker V, Bomeh J, Redd JT, Bunga S, Brunkard J, Kilmarx PH: **Cluster of Ebola Virus Disease Linked to a Single Funeral — Moyamba District, Sierra Leone, 2014.** *Morb Mortal Wkly Rep* 2016, **65**:202-205.
58. Holmes EC, Dudas G, Rambaut A, Andersen KG: **The evolution of Ebola virus: insights from the 2013–2016 epidemic.** *Nature* 2016, **538**:193-200.
- This review addresses very well several aspects of Ebola virus biology during the last 2013–2016 outbreak, such as the controversy about evolution rates, the lack of changing in route of transmission or the sexual transmission of the virus.
59. Carroll MW, Matthews DA, Hiscox JA, Elmore MJ, Pollakis G, Rambaut A, Hewson R, Garcia-Dorival I, Bore JA, Koundouno R *et al.*: **Temporal and spatial analysis of the 2014–2015 Ebola virus outbreak in West Africa.** *Nature* 2015, **524**:97-101.
- Carroll *et al.* retraced the genetic evolution of the Ebola virus during the last 2014 outbreak in West Africa by sequencing 179 full genomes of viruses isolated from patients using next generation sequencing. They traced back the origin and time of transmission of the virus in the different affected countries and found that the outbreak has resulted in multiple lineages.
60. WHO: **Factors that contributed to undetected spread of the Ebola virus and impeded rapid containment.** URL: <http://www.who.int/csr/disease/ebola/one-year-report/factors/en/>.