REVIEW ARTICLE

Open Access

The role of noncoding RNAs in epithelial cancer

Massimiliano Agostini¹, Carlo Ganini¹, Eleonora Candi^{1,2} and Gerry Melino^{1,3}

Abstract

Regulatory noncoding RNAs (ncRNAs) are a class of RNAs transcribed by regions of the human genome that do not encode for proteins. The three main members of this class, named microRNA, long noncoding RNA, and circular RNA play a key role in the regulation of gene expression, eventually shaping critical cellular processes. Compelling experimental evidence shows that ncRNAs function either as tumor suppressors or oncogenes by participating in the regulation of one or several cancer hallmarks, including evading cell death, and their expression is frequently deregulated during cancer onset, progression, and dissemination. More recently, preclinical and clinical studies indicate that ncRNAs are potential biomarkers for monitoring cancer progression, relapse, and response to cancer therapy. Here, we will discuss the role of noncoding RNAs in regulating cancer cell death, focusing on those ncRNAs with a potential clinical relevance.

Facts

- Many of the genetic and epigenetics alterations in human cancers are found in noncoding regions of the DNA.
- miRNAs and lncRNAs have been largely described in their role in controlling epithelial tissues homeostasis, both in physiological and pathological conditions such as cancer.
- The clinical application of ncRNAs is already under evaluation in ongoing clinical trials exploring their role as biomarkers for patient survival, metastasis development prediction, or therapy response.

Open questions

- What is the role of regulatory ncRNA in pyroptosis, ferroptosis, and autophagy?
- What is the functional interplay between miRNA, lncRNA, and circRNA in cancer biology?

Correspondence: Massimiliano Agostini (m.agostini@med.uniroma2.it) or Eleonora Candi (candi@uniroma2.it) or Gerry Melino (gm614@mrc-tox.cam.ac.uk) ¹Department of Experimental Medicine, TOR, University of Rome "Tor Vergata", 00133 Rome, Italy

²IDI-IRCCS, Via Monti di Creta 106, 00166 Rome, Italy Full list of author information is available at the end of the article. Edited by I. Amelio

How can we integrate preclinical findings to select the best regulatory ncRNA for clinical studies?

Introduction

Protein-coding genes undoubtfully play an established role in cancer transformation and progression, however, recent compelling evidence is highlighting a role for noncoding RNAs. There is no question that loss or mutation of crucial genes are frequent events in cancer biology: the mutation of the tumor suppressor gene p53 is observed in almost 50% of human cancers¹⁻⁷, and the amplification of the oncogene MYC⁸ or the translocation of BCL-2 gene^{9–12} are crucial drivers in various cancer contexts^{13–16}. However, many of the recurrent genetic and epigenetic alterations are found in genes that do not codify for proteins but that codify for an entire class of molecules called noncoding RNAs (ncRNAs), which play a key role in the regulation of many cellular activities.

The non-protein-coding regions of the human genome are transcribed into molecules of RNA classified as regulatory noncoding RNAs¹⁷. ncRNAs are mainly transcribed by RNA polymerase II and share several characteristics with messenger RNAs (mRNAs). Indeed, they have a cap structure at the 5' end, a poly(A) tail at the 3' end and their expression is controlled by canonical promoter elements and transcription factors. Conventionally, regulatory ncRNAs are classified either as

© The Author(s) 2020

Open Access This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons license, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons license, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons license and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this license, visit http://creativecommons.org/licenses/by/4.0/.



Table 1 Pro-apoptotic and anti-apoptotic ncRNAs.

ncRNA	PRO-apoptotic role	Cellular/animal model	Relevant references (original papers)
Let-7	Inhibition of BCL2L1 Upregulation of BAK and BAX, and downregulation of BCL-xL	Colorectal cancer Leukemia	Mizuno et al. ⁴⁶ Huang et al. ⁴⁷
miR-15/16	• Repression of BCL-2 and BMI1	Chronic lymphatic leukemiaMantle cell lymphoma	Cimmino et al. ⁵² Teshima et al. ⁵³
miR-34	Regulation of proteins involved in cell death: BCL-2, BIRC5 (Survivin), CREB, and YY1	Pancreatic cancerMyeloid leukemiaNeuroblastomaLaryngeal squamous cell cancer	Chang et al. ⁶⁷ Pigazzi et al. ⁶⁸ Chen et al. ⁶⁹ Shen et al. ⁷⁰
miR-29	Repression of MCL-1 expression	 Acute myeloid leukemia 	Garzon et al. ⁷⁶
GAS5	 Induction of apoptosis in a mouse model 	Brest cancer (mouse)	Mourtada-Maarabouni et al. ¹⁰³
MEG3	• Induction of p53	Colorectal cancer	Zhou et al. ¹⁰⁸
NIKLA	 Activation of apoptosis in tumor-specific CTLs 	Breast cancer and lung cancer	Huang et al. ¹¹³
NEAT1 ^a	Enhanced apoptosis after DNA damage	Chronic lymphocytic leukemia	Blume et al. ¹¹⁶
circFOXO3	Increased PUMA expression	Breast cancer (mouse)	Du et al. ¹⁴⁷
ncRNA	ANTI-apoptotic role	Cellular/animal model	Relevant references (original papers)
miR-21	expression of pro-apoptotic genes: APAF11, PDCD4, RHOB, and FASLG	Non-small cell lung cancer	Hatley et al. ⁸⁴
miR-155	• Repression of SHIP-1	B lymphocytesHematopoietic cells	Costinean et al. ⁹¹ O'Connell et al. ⁹²
miR-221	 Increased apoptosis and cell-cycle arrest 	 Hepatocellular carcinoma 	Park et al. ⁹⁸
CCAT	 Interaction with p63 and SOX2 	 Squamous cell carcinoma 	Jiang et al. ¹²³
FAL1	• Interaction with BMI1	Pan-cancer database	Hu et al. ¹²⁸
PVT1	 Downregulation of Caspase-7, Caspase-9, and PARP 	Nasopharyngeal carcinoma	He et al. ¹³²

^aControversial role since NEAT1 has also been correlated to an oncogenic activity in some tumor types.

small ncRNAs, if they are shorter than 200 ribonucleotides or as long noncoding RNAs (lncRNAs), longer than 200 ribonucleotides. Small ncRNAs include microRNAs (miRNAs), which mediates post-transcriptional RNA silencing, piwiRNAs, which regulate chromatin modifications and transposons repression, as well as the more recent circular RNAs (circRNAs)¹⁸.

In this review, we shall discuss the role of ncRNAs in regulating cancer cell death (Table 1)¹⁹. In particular, we will describe those regulatory ncRNAs that have been largely investigated, on the basis of both in vitro and in vivo evidence as clinical data.

Epithelial ncRNAs

Recently, miRNAs and lncRNAs have gained significant attention for their role in controlling epithelial tissue homeostasis in normal and pathological conditions, being key regulators of epithelial progenitor stem cells proliferation, somatic lineage specification, and differentiation^{20,21}.

Distinct miRNAs control epidermal development, epidermal differentiation, and adult stem cells maintenance (for reviews see refs. ^{22,23}). These include the miR-20 family, miR-24, the miR-200 family, miR-205, and miR-203^{24–28}. The latter has been described in his mechanistic role in the control of epithelial progenitor cells proliferation and in the inhibition of cellular senescence, having as a target one of the epithelial master genes, the transcription factor TP63^{24,29}. Interestingly, these

epithelial-specific miRNAs have also been implicated in controlling key regulators genes of the pathogenesis of many adenocarcinomas and squamous cell carcinomas (SCC)³⁰. As an example, the miR-200 family, controlling the expression of ZEB1 and ZEB2, have been described in the epithelial-to-mesenchymal transition (EMT) in ovarian cancer and SCCs. Moreover, miR-203, targeting ABL1, SOCS3, and ZEB3, controls EMT in epithelial cancers^{28,31}. miRNAs also play a key role in epithelial cells upon different environmental stressors, such us UV radiations and inflammation^{32–36}.

Only a few lncRNAs have been studied so far in the regulation of epithelial development, such as ANCR, an anti-differentiation lncRNA, and TINCR, a terminal differentiation-induced lncRNA; however, the mechanism through which they maintain the cell of the epidermis in a undifferentiated state has not been identified yet^{37,38}. Similarly, LIN00941 has also been implicated in the control of epidermal homeostasis repressing the expression of pro-differentiation genes through a not-yet identified mechanism³⁹. Moreover, the pro-differentiation lncRNA, uc.291, has been shown acting as a pro-differentiation transcript by facilitating the activation/binding of the chromatin remodeling complex SWI/SNF (BAF) in proximity to the epidermal differentiation complex (EDC) genes to allow their expression⁴⁰.

While several studies investigated the specific contributions of miRNAs in epithelial cancer development,

the role of lncRNAs in this context has not been investigated thoroughly yet.

microRNAs with pro-apoptotic functions let-7

let-7 has been one of the first miRNAs isolated and characterized in the nematode C. elegans. The human genome contains 13 let-7 family member genes, which are distributed in different genomic loci and codify for 9 mature miRNAs⁴¹. Let-7 expression is downregulated in many cancers, such as lung, breast, pancreatic cancer, and melanoma and in many cancer-associated clinical conditions like cholestasis⁴². Moreover, downregulation of let-7 expression correlates with poor survival in lung cancer⁴³; ovarian cancer⁴⁴, and head and neck SCC patients⁴⁵. Functionally, let-7 acts as a tumor suppressor partially through the downregulation of several genes involved in cell death, as shown by the ectopic expression of let-7 resulting in the inhibition of BCL2L1 in colorectal cancer⁴⁶ or by the induction of apoptosis via upregulation of BAK and BAX and downregulation of BCL-XL protein levels⁴⁷. Although in vivo delivery of let-7 in mouse models of lung cancer demonstrated its therapeutic potential, its tumor suppressor functions might be associated with repression of cell proliferation and elimination of cancer cells mainly through a non-apoptotic mechanism⁴⁸.

miR-15/16

miR-15 and miR-16 are localized on chromosome 13q14. This chromosomic region is frequently deleted in B-cell chronic lymphocytic leukemia (B-CLL)⁴⁹. miR-15 and miR-16 are found in a 30-kb region, which is lost in CLL, and both genes are either deleted or downregulated in most of CLL cases (~68%)⁵⁰. However, several studies suggested that miR-15/16 could be also downregulated by additional mechanisms, such as defective DROSHA processing or epigenetic alterations⁵¹. The tumor suppressor action of miR-15/16 is the result of the induction of cell death through the repression of the anti-apoptotic proteins BCL-2 and BMI1 (Fig. 1a)⁵²⁻⁵⁷. More recently, the tumor suppressor function of miR-15/16 has also been observed in solid tumors such as mesothelioma⁵⁸ or chondrosarcomas⁵⁹, where its expression is suppressed in order to promote tumor neo-angiogenesis. Moreover, the miR-15/16 knockout mouse model supports the in vivo tumor suppressor activity of miR-15/16. Indeed, deletion of miR-15/16 gene results in B cells proliferation and in the development of lymphoid malignancies^{60,61}.

miR-34

The miR-34 family consists of three different transcripts miR-34a, miR-34b, and miR-34c with high sequence homology.

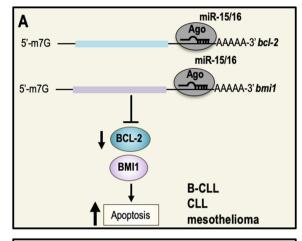
In humans, the miR-34a gene is located on the chromosomal region 1p36.22, which is frequently deleted in many human cancers, including neuroblastoma, glioma, breast cancer, lung cancer, colorectal cancer, and melanoma⁶²⁻⁶⁴. The miR-34b and miR-34c genes are both encoded on chromosome 11q23.1, and rearrangements of this region have been observed in several solid tumors and in hematological malignancies⁶⁵. Both in vitro and in vivo studies have shown that the miR-34 family genes (in particular miR-34a) act as tumor suppressors: when overexpressed they repress several oncogenes, resulting in an increased cancer cell death and in an inhibition of metastasis development. miR-34a has also been shown to play a role in the regulation of NF-κB in CD8 + T cells, promoting their cytotoxic activity⁶⁶. In the context of cell death, miR-34 family acts on proteins such as BCL-2, BIRC5 (Survivin), CREB, and YY1, which are involved in apoptosis regulation (Fig. 1b)^{67–71}.

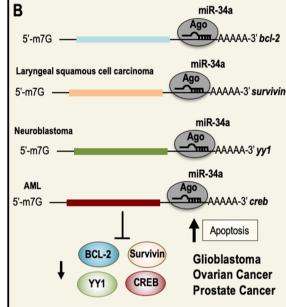
Nevertheless, it should be noted that miR-34-deficient animals are not showing increased susceptibility to either spontaneous or to irradiation-induced nor MYC-initiated tumorigenesis.

Strikingly in 2013, a miR-34 mimic (MRX34) became the first microRNA tested in a phase 1 clinical trial (NCT002862145). In particular, a liposomal miR-34 mimetic was administered intravenously to patients with unresectable liver cancer or metastatic cancer refractory to standard treatment, with or without liver involvement. From these studies, MRX34 treatment was associated with antitumor activity and acceptable safety, but subsequent monitoring showed immune-related toxicities. To date, it is not clear whether further clinical trials will be started⁷².

miR-29

The miR-29 family is composed of three isoforms. miR-29b-1 and miR-29a form one cluster on chromosome 7q32, while miR-29b-2 and miR-29c form a second cluster on chromosome 1q23. Of note that the first cluster region is frequently deleted in myelodysplastic syndromes and therapy-related acute myeloid leukemia (AML)^{73,74}. In addition, several experimental observations showed that miR-29 family is downregulated in chronic lymphocytic leukemia, lung cancer, invasive breast cancer, and cholangiocarcinoma. miR-29 functions as a tumor suppressor gene in several cancer types. Indeed, ectopic expression of miR-29b induced apoptosis in cholangiocarcinoma cell lines and reduced tumorigenicity in a xenograft model of lung cancer, rhabdomyosarcoma, and AML⁷⁵. Mechanistically, miR-29 induces cell death by directly repressing the expression of the anti-apoptotic BCL-2 family member, MCL-1 (Fig. 1c)⁷⁶. In addition, modulation of miR-29 expression results in the upregulation of some pro-apoptotic genes, such as BIM and the tumor suppressor programmed cell death-4 (PDCD4) in





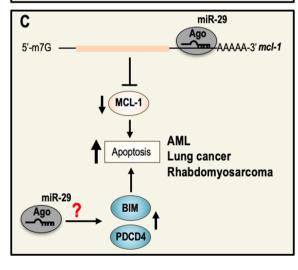


Fig. 1 microRNAs with pro-apoptotic functions and relative inhibited molecular pathways. a miR-15/16 expression is deregulated in hematological malignancies and induces cell death by inhibiting the expression of the anti-apoptotic protein BCL-2 and BMI1. **b** miR-34a acts as a tumor suppressor gene by directly repressing the expression of several proteins involved in the regulation of cell survival. **c** miR-29a regulates the critical anti-apoptotic genes MCL-1. MCL-1 is an anti-apoptotic protein that promotes cancer cell survival and proliferation, and it is frequently overexpressed in AML. It has been also shown that miR-29 can induce cell death by inducing the expression of pro-apoptotic proteins, including BIM and PDCD4. BCL-2 B-cell lymphoma 2 gene, BMI1 polycomb complex protein BMI1, MCL-1 induced myeloid leukemia cell differentiation protein, BIM BCL-2-like protein 11, PDCD4 programmed cell death protein 4, AML acute myeloid leukemia.

AML, and also inhibits the expression of DNMT3B in hepatocarcinoma cell lines⁷⁷. Interestingly, MCL-1 mRNA inversely correlated with miR-29a or miR-29b expression in 45 primary AML samples⁷⁸.

microRNAs with anti-apoptotic functions miR-21

The human miR-21 gene is located in the fragile site FRA17B on chromosome 17q23.2⁷⁹. Expression profile studies on tumor samples, including lung, breast, stomach, prostate, colon, pancreatic tumors, and B-cell lymphomas showed that miR-21 is the most commonly upregulated miRNA in solid tumors and hematological malignancies, indicating miR-21 as an "oncomiR"80-82. Indeed, functional studies in several cancer cell lines demonstrated that knockdown of miR-21 activates caspases leading to apoptotic cell death. Moreover, in vitro evidence also suggested a positive regulatory role in pyroptosis through the activation of the NLRP3 inflammasome⁸³. At a molecular level, miR-21 suppresses the expression of pro-apoptotic genes, such as APAF11, PDCD4, RHOB, and FASLG (Fig. 2a)⁸⁴. The oncogenic role of miR-21 was also confirmed in vivo by generating conditionally expressing miR-21 mice⁸⁴. This mouse model demonstrated that miR-21 is capable of initiation, maintenance, and prolonged survival of tumors in vivo and demonstrated the importance of miR-21 in hematological malignancies⁸⁵.

miR-155

The human gene miR-155 is localized on chromosome 21 within an exon of a noncoding RNA, from the B-cell Integration Cluster (BIC)⁸⁶. miR-155 is mainly expressed in lymphoid tissues, including thymus and spleen, where it regulates several physiological functions of these organs, such as antibodies and cytokines production⁸⁷. In addition, several observations have shown that miR-155 is accumulated in non-Hodgkin lymphomas, as in the

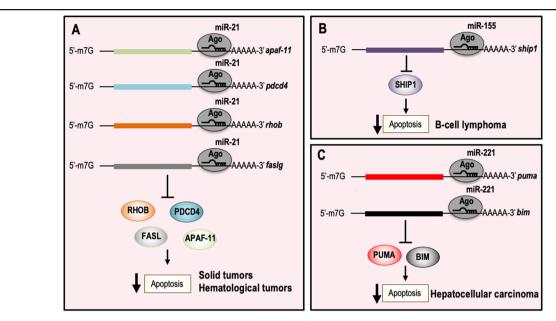


Fig. 2 microRNAs with anti-apoptotic functions and relative inhibited molecular pathways. a miR-21 is the most upregulated oncomiR in solid tumors and hematological malignancies. The oncogenic activity of miR-21 is associated with the direct negative regulation of pro-apoptotic genes, such as APAF11, PDCD4, RHOB, and FASLG. **b** miR-155 plays a key role in the regulation of the immune response. miR-155 is upregulated in B-cell lymphomas and by targeting SHIP-1 supports cell survival. **c** In liver cancer, miR-221 exerts oncogenic activity by repressing the expression of the apoptotic proteins PUMA and BIM. APAF11 apoptotic protease activating factor 1, PDCD4 programmed cell death protein 4, RHOB Ras Homolog Family Member B, FASLG Fas ligand, SHIP-1 Src homology 2-SH2 domain containing inositol polyphosphate 5-phosphatase 1, PUMA p53 upregulated modulator of apoptosis, BIM BCL-2-like protein 11.

diffuse large B-cell lymphomas, and Burkitt lymphomas and in Hodgkin lymphomas^{88–90}. The oncogenic role of miR-155 was confirmed by the generation of genetically modified mice, developing B-cell lymphoma upon its overexpression. Among the molecular mechanisms underlying its oncogenic properties, the direct repression of SH2-containing inositol phosphatase (SHIP-1) (Fig. 2b)^{91,92}, a positive regulator of apoptosis, seems to play a pivotal role. Apart from hematologic malignancies, miR-155 is also overexpressed in several solid tumors, such as breast, colon, pancreatic, and lung cancer, and has also been described in the regulation of autophagy in an experimental mouse model of pancreatitis⁹³. More recently, the therapeutic efficacy of an anti-miR-155 was tested in vivo by using a novel delivery system targeting the acidic tumor microenvironment: a nucleic acid antimiR-155 linked to a peptide with a low pH-induced transmembrane structure (pHLIP) effectively inhibited miR-155 expression after intravenous administration, leading to cancer regression in a mouse model of lymphoma⁹⁴.

miR-221

miR-221/222 is a microRNA with oncogenic properties, highly conserved in vertebrates, located on the X chromosome in humans, mice, and rats. Overexpression of miR-221/222 has been observed in several human

malignancies, including hepatocellular carcinoma, breast, prostate, pancreatic cancer, and glioblastoma^{95,96}. The oncogenic activities of miR-221 were also confirmed in vivo as the overexpression of miR-221 in p53^{-/-}; Myc liver progenitors stimulates tumor onset and progression⁹⁷. The oncogenic activity of miR-221 might be partially explained by the repression of PUMA and BIM proteins (Fig. 2c). Interestingly, in vivo preclinical studies showed that a cholesterol-modified isoform of anti-miR-221 significantly reduced miR-221 levels in livers after injection, reducing tumor cell proliferation, and increasing the expression of apoptosis and cell-cycle arrest markers, eventually prolonging mice survival⁹⁸. Although miR-221 is classified as an "oncomiR", it should be noted that his role as tumor suppressor gene has also been observed⁹⁹.

Long noncoding RNAs

Long noncoding RNAs are classified as endogenous ncRNAs longer than 200 nucleotides. They are transcribed by RNA polymerase II from an independent promoter and processed as coding RNAs. Indeed, they are capped, spliced, and polyadenylated, however, lncRNAs lack a significant open-reading frame. Compared with protein-coding transcripts, lncRNAs are overall expressed at lower levels and they are not highly evolutionarily conserved, with only 5–6% of lncRNAs harboring conserved sequences¹⁰⁰.

Several studies have described a range of molecular mechanisms by which lncRNAs may exert their functions. In particular, lncRNAs have been described as molecular scaffolds or architectural RNAs in a variety of cellular processes, among which epigenetics modifications, alternative splicing, mRNA translation, and maintenance, and acting as decoys or "sponges" for miRNAs or transcription factors.

IncRNAs with pro-apoptotic function *GAS5*

The growth arrest-specific transcript 5 (GAS5) is located at 1q25, and the gene transcribed contains 12 exons that do not encode for functional proteins¹⁰¹. GAS5 is a tumor suppressor gene, and is one of the most expressed lncRNAs in all human tissues¹⁰². The expression of GAS5 in cancers is significantly reduced. Reduced expression has been observed in breast, prostate, head and neck, gastric, colorectal, pancreatic, and cervical cancer, and more importantly, its expression is negatively correlated with clinicalpathological characteristics, such as tumor size, staging, or metastasis. Moreover, in vivo studies have shown inhibition of breast tumor growth by inducing cell-cycle arrest and apoptosis after the overexpression of GAS5 in breast cancer cell lines and their subsequent injection in nude mice¹⁰³. Several molecular mechanisms of action for GAS5 have been proposed, including inhibition of translation and decoy or miRNA sponge activity. The decoy function of GAS5 has been shown in HeLa and HepG2 cell lines, and seems to be its major mechanism of action. Indeed, GAS5 interacts with the DNA-responsive elements of glucocorticoids receptor preventing the binding of the receptor to the DNA, thereby blocking the activation of target genes transcription (Fig. 3a)¹⁰⁴. Moreover, GAS5 can also interact with other steroid hormone receptors including progesterone and androgen receptors, which play important roles in hormone-dependent cancers¹⁰².

MEG3

Maternally expressed gene 3 (MEG3) located on the human chromosome region 14q32.3 is expressed in many normal tissues¹⁰⁵. However, MEG3 expression is lost in several human tumors including osteosarcoma, hepatocellular cancer, gastric cancer, and non-small cell lung cancer (NSCLC) by promoter or intergenic differentially methylated region hypermethylation, suggesting that loss of MEG3 expression contributes to tumor development in several tissues 106,107. Importantly, patients with lower levels of MEG3 expression had a relatively poor prognosis. MEG3 functions as a tumor suppressor gene, and its reexpression inhibits cell proliferation and promotes apoptosis in human glioma and NSCLC cell lines. At a molecular level, MEG3 function is mediated, at least partially, by the activation of the tumor suppressor gene p53 by the downregulation of MDM2 expression ^{108–110}.

NKILA

The human gene that encodes NKILA, an NF-κBinteracting lncRNA, is located on chromosome 20. NKILA is downregulated in breast cancer, nasopharyngeal carcinoma, and melanoma¹¹¹. In addition, reduced NKILA expression is associated with breast cancer metastasis and poor patient prognosis 112. Mechanistically, the transcription factor NF-κB upregulates the expression of NKILA, which in turn binds to NF-κB/IκB, and directly masks the phosphorylation motifs of IkB, resulting in the inhibition of IKK-induced IkB phosphorylation, and NFκB activation, forming a negative regulatory loop (Fig. 3b). This regulatory mechanism is promoting tumorigenesis by inhibiting apoptosis and increasing invasion. More recently, NKILA has also been shown to regulate T-cells activation by inhibiting NF-kB activity, and ectopic expression of NKILA in tumor-specific CTLs and T_H1 cells correlated with apoptosis and shorter patient overall survival¹¹³.

NEAT1

The Homo sapiens nuclear paraspeckle assembly transcript 1 (NEAT1) sequence is a product of the NEAT1 gene, which is located on chromosome 11. NEAT1 has been identified as a p53 target gene that plays an important role in the formation of paraspeckles, and is indispensable for cell-cycle arrest and apoptosis in response to genotoxic stress¹¹⁴. Therefore, it has a key function in suppressing neoplastic transformation and cancer onset. Indeed, in a pancreatic cancer mouse model, NEAT1 deficiency increased transformation and contributed to the development of premalignant pancreatic intraepithelial neoplasia and cystic lesions through global changes in gene expression¹¹⁵. Moreover, NEAT1 expression is downregulated in several cancers, while increased NEAT1 levels are correlated with better overall survival in colorectal cancer patients. In addition, elevated NEAT1 levels have been associated with enhanced apoptosis after DNA damage by irradiation of chronic lymphocytic leukemia cells 116,117. However, NEAT1 role is controversial since several studies also characterized its oncogenic activity. As matter of fact, during tumorigenesis NEAT1 levels are increased, and high levels of NEAT1 were associated with worse prognosis in gastric adenocarcinoma and laryngeal SCC^{118,119}. Accordingly, modulation of NEAT1 expression promotes cell survival and/or proliferation of human cancer cell lines¹²⁰, and NEAT1 acts as an oncogene in a mouse model of skin carcinogenesis¹¹⁴. At a molecular level, one possible mechanism by which NEAT1 exerts its oncogenic role, at least in ovarian cancer, is the interaction with the tumor suppressor miRNA miR-34a-5p, acting as a sponge and negatively regulating miR-34a expression.

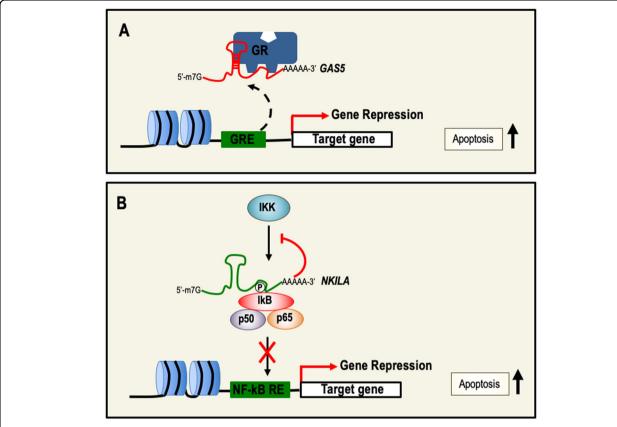


Fig. 3 IncRNAs as inducers of cell death and relative molecular mechanisms. a GAS5 prevents GR-dependent gene activation by binding to the glucocorticoids receptor (GR) functioning as a decoy. **b** NF-κB is a key transcription factor in the regulation of cell survival. NKILA binds IkB and prevents its phosphorylation resulting in the inhibition of NF-κB activation, thereby repressing NF-κB target genes involved in cell survival. GAS5 growth arrest-specific transcript 5, GR glucocorticoids receptors, NF-κB nuclear factor kappa-light-chain-enhancer of activated B cells, IkB inhibitor of kappa B.

Overall, NEAT1 can elicit a context-specific function in either promoting or suppressing neoplastic transformation.

IncRNAs with anti-apoptotic functions CCAT

Colon cancer-associated transcript (CCAT) is located within the 8q.24.21 genomic region that is frequently amplified in colorectal cancer (CRC)^{121,122}. Expression profile studies highlight that CCAT1 and CCAT2 genes are frequently overexpressed in CRC, and their expression is associated with shorter progression-free and overall survival.

The oncogenic function of CCAT is the result of the activation of genes involved in cell proliferation and in the inhibition of apoptosis. Indeed, CCAT regulates the overexpression of MYC, possibly through its physical interaction with TCF7L2, leading to genomic instability and promoting cell growth (Fig. 4a). In addition, a second possible molecular regulatory mechanism has been observed in SCC; in this context, CCAT regulates transcription of genes involved in cell proliferation and

survival by a physical interaction with both transcription factors p63 and $SOX2^{123}$. (Fig. 4b). Finally, overexpression of CCAT has been observed in other types of cancer, including gastric, breast, and lung cancer¹²⁴.

FAL1

The gene encodes for focally amplified LncRNA on chromosome 1 (FAL1), and is localized at the chromosomal region 1q21.2. FAL1 functions as an oncogene in several cancers, and is upregulated in ovarian, thyroid cancer and NSCLC^{125,126}. In addition, FAL1 expression correlates with some clinical and pathological characteristics of NSCLC and ovarian cancer patients. Both in vitro and in vivo studies demonstrated that the biological function of FAL1 is to regulate cell cycle, cell death, migration, and invasion¹²⁷. Mechanistically, FAL1 interacts with the epigenetic repressor BMI1 and modulates the transcription of a number of genes involved in cell-cycle arrest and apoptosis, such as CDKN1A, FAS, BTG2, TP53I3, FBXW7, and CYFIP2¹²⁸ (Fig. 4c).

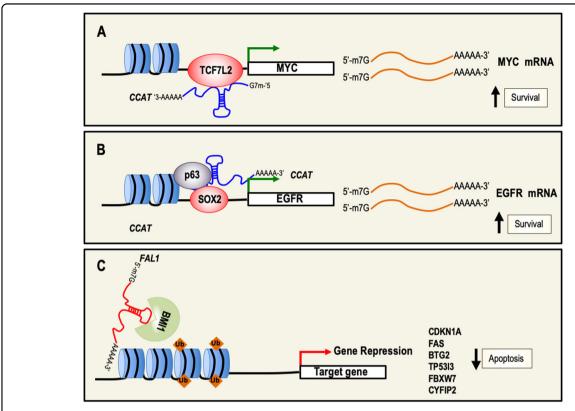


Fig. 4 IncRNAs with an anti-apoptotic activity and relative molecular mechanisms. a CCAT is an oncogene that protects from cell death by upregulating the expression of MYC. **b** p63, SOX2, and CCAT form a trimeric complex, which in turn binds the promoter region of EGFR and positively upregulates its expression. This results in sustaining cell survival. **c** FAL1 associates with the epigenetic repressor BMI1 and regulates its stability in order to modulate the transcription of a number of genes involved in cell death. CCAT colon cancer-associated transcript, MYC Myc proto-oncogene, SOX2 sex determining region box-2, EGFR epithelial growth factor receptor, FAL1 focally amplified LncRNA on chromosome 1, BMI1 polycomb complex protein BMI1.

PVT1

The human PVT1 gene is located in the human chromosome 8q24 region close to the well-established oncogene MYC¹²⁹. Interestingly, the amplification of 8q24 is a frequent event in a vast variety of cancers including CRC, where it is also associated with clinical findings of decreased overall survival 130,131. PVT1 functions as an oncogene by inhibiting the apoptosis of tumor cells, promoting cell proliferation, and affecting tumor invasion and metastasis generation. However, a detailed molecular mechanism underlying its anti-apoptotic activity is still missing. Recently, a possible mechanism by which PVT1 inhibits cell death has been described in nasopharyngeal carcinomas. In particular, PVT1 downregulates the expression of cleaved caspase-9, caspase-7, and PARP, inhibiting apoptosis and also promoting radiation resistance 132,133

Circular RNAs

Circular RNAs (circRNAs) were identified more than 20 years ago, and for many years have just been considered as

secondary products of aberrant splicing processes ¹³⁴. Only recently, with the advent of next-generation sequencing, a large number of circRNAs has been identified, several of them showing high and stable expression. Nearly 10% of genes transcribed in cells can produce circRNAs.

The research on circRNAs is still in his infancy and their specific functions in several physiological processes and in the pathogenesis of human diseases is still largely unknown. However, recently high-throughput sequencing

studies on clinical tumor samples showed a deregulation of circRNAs expression in tumors 138,139. In particular, their downregulation has been described in proliferative cells across different tumor types, indicating that some circRNAs may have tumor suppressive roles. Global transcriptome analysis on 144 samples of localized prostate cancer, identifying 76,311 circRNAs, showed a correlation of circRNAs expression with tumor aggressivity, suggesting that 171 circRNAs are essential to prostate cancer cell proliferation 140.

Overall, studies on clinical samples together with in vitro experiments showed that circRNAs play a role in several hallmarks of cancer having either tumor suppressive or oncogenic functions 141-143. To our knowledge, no circRNAs from the literature have a clear and defined role in cancer biology proven by in vitro or in vivo experiments or by clinical data. However, we have decided to describe one of the more studied circRNAs that has a role in cell death regulation.

circFOXO3

circFOXO3 is a circular transcript of 1435 nucleotides derived from the tumor suppressor gene FOXO3¹⁴⁴. Several expression profile studies indicated that circFOXO3 is downregulated in several tumors such as breast and NSCLC, suggesting a possible role as a tumor suppressor gene^{145,146}. Both gain and loss of function experiments showed that circFOXO3 function is associated with the induction of apoptosis and with the inhibition of cell-cycle progression and angiogenesis. At a molecular level, ectopic expression of circFOXO3 decreases the interaction between FOXO3 and MDM2, releasing FOXO3 from MDM2-dependent degradation, therefore increasing FOXO3 activity, promoting PUMA expression, and enhancing cell death¹⁴⁷.

Perspectives and conclusions

During the last decades, we have witnessed considerable developments in understanding the role of the regulatory ncRNAs in the regulation of physiological processes as well as in the pathogenesis of several diseases. Deregulation of ncRNAs expression has been largely documented during the initiation, progression, and dissemination of cancer, and preclinical studies have shown that they can be used as diagnostic and prognostic biomarkers. More importantly, it has been documented that ncRNAs can be released into the extracellular space and detected in body fluids (blood or urine) as circulating ncRNAs, potentially showing their application in liquid biopsies 148. The clinical application of ncRNAs is already under evaluation in ongoing clinical trials trying to describe their role as biomarkers for patient survival, metastasis development prediction, or therapy response (Table 2).

Current clinical trials involving ncRNAs. 7

ncRNA	Study title	Condition or disease	Interventions	Primary purpose	Clinical trails identifier	Status
miR-16	MesomiR 1: A Phase I Study of TargomiRs as 2nd or 3rd Line Treatment for Mesothelioma Patients with Recurrent MPM and NSCLC laters with Recurrent MPM and NSCLC	Mesothelioma Non-small cell Iung cancer	Drug: TargomiRs	Treatment	NCT02369198	Completed
miR-34	Critical Role of MicroRNA-34a and MicroRNA-194 in Acute Myeloid Leukemia With CEBPA Mutations	Leukemia	Genetic: RNA analysis	Not Reported	NCT01057199	Completed
miR-29	Phase 1: safety, tolerability and pharmacokinetic study of MRG-201 in healthy volunteers	Healthy volunteers	Drug: MRG-201	Treatment	NCT02603224	Completed
miR-155	The Potential Role Of microRNA-155 And Telomerase Reverse Transcriptase In Diagnosis Of Non-Muscle Invasive Bladder Cancer And Their Pathological Correlation	Bladder cancer	Diagnostic test	Diagnostic	NCT03591367	Completed
miR-221	Clinical Significance of Hepatic and Circulating microRNAs miR-221 and miR-222 in Hepatocellular Carcinoma	Hepatocellular carcinoma	Hepatocellular carcinoma Analysis of microRNA expression	Not Reported	NCT02928627	Unknown
miRNA/ IncRNA	Noncoding RNA in the exosome of the epithelial ovarian cancer	Ovarian cancer	Sequencing miRNA/IncRNA		NCT03738319	Recruiting
miRNA	Circulating miRNAs as biomarkers of hormone sensitivity in breast cancer Breast cancer	Breast cancer	Drugs: tamoxifen, letrozole, anastrozole, exemestane	Treatment	NCT01612871	Completed
miRNA	Circulating microRNA as disease markers in pediatric cancer	Leukemia lymphoma	Not reported	Observational	NCT01541800	Recruiting
Source: ClinicalTrials.gov.	alTrials.gov.					

As from above, regulatory ncRNAs play a key role in driving or preventing the process of tumorigenesis and preclinical studies indicate that modulation of their expression by mimetic or inhibitory oligonucleotides might be a novel therapeutic approach for cancer therapy. Indeed, a limited number of clinical trials using drugs based on ncRNAs, especially on microRNAs, are evaluating clinical safety, tolerability, and efficacy of this approach (Table 2). Although therapeutic applications are still at an early phase, a new chapter in the pharmacological treatment of human diseases has started. The recent approval of the RNA-targeting oligonucleotides drug Spinraza for the treatment of spinal muscular atrophy by FDA¹⁴⁹ makes reasonable to believe that more ncRNAs-based drugs will be soon available on the patient's bedside.

Acknowledgements

We apologize for those whose contributions could not be cited due to space constraints. This work has been supported by the Medical Research Council (to G.M.), UK Associazione Italiana per la Ricerca contro il Cancro (AIRC) to G.M. IG #20473 (2018–2022), to E.C. (AIRC IG #22206; 2019–2023). This work has been also supported by Regione Lazio through Laziolnnova Progetto Gruppo di Ricerca n 85–2017–14986 to G.M. and by Fondazione Luigi Maria Monti IDI-IRCCS (R.C. to E.C.), Ministry of Health & MAECI Italy-China Science and Technology Cooperation (#PGR00961).

Author details

¹Department of Experimental Medicine, TOR, University of Rome "Tor Vergata", 00133 Rome, Italy. ²IDI-IRCCS, Via Monti di Creta 106, 00166 Rome, Italy. ³MRC Toxicology Unit, University of Cambridge, Department of Pathology, Tennis Court Road, Cambridge CB2 1QP, UK

Conflict of interest

The authors declare that they have no conflict of interest.

Publisher's note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Received: 27 January 2020 Revised: 11 February 2020 Accepted: 17 February 2020

Published online: 12 March 2020

References

- Kastenhuber, E. R. & Lowe, S. W. Putting p53 in context. Cell 170, 1062–1078 (2017).
- Aubrey, B. J., Kelly, G. L., Janic, A., Herold, M. J. & Strasser, A. How does p53 induce apoptosis and how does this relate to p53-mediated tumour suppression? *Cell Death Differ.* 25, 104–113 (2018).
- Baugh, E. H., Ke, H., Levine, A. J., Bonneau, R. A. & Chan, C. S. Why are there hotspot mutations in the TP53 gene in human cancers? *Cell Death Differ.* 25, 154–160 (2018).
- Kaiser, A. M. & Attardi, L. D. Deconstructing networks of p53-mediated tumor suppression in vivo. Cell Death Differ. 25, 93–103 (2018).
- Kim, M. P. & Lozano, G. Mutant p53 partners in crime. Cell Death Differ. 25, 161–168 (2018).
- Parrales, A., Thoenen, E. & Iwakuma, T. The interplay between mutant p53 and the mevalonate pathway. Cell Death Differ. 25, 460–470 (2018).
- Nemajerova, A. et al. Non-oncogenic roles of TAp73: from multiciliogenesis to metabolism. Cell Death Differ. 25, 144–153 (2018).
- Gabay, M., Li, Y. & Felsher, D. W. MYC activation is a hallmark of cancer initiation and maintenance. *Cold Spring. Harb. Perspect. Med.* 4, https://doi. org/10.1101/cshperspecta014241 (2014).

- Tsujimoto, Y., Finger, L. R., Yunis, J., Nowell, P. C. & Croce, C. M. Cloning of the chromosome breakpoint of neoplastic B cells with the t(14;18) chromosome translocation. Science 226, 1097–1099 (1984).
- Adams, J. M. & Cony, S. The BCL-2 arbiters of apoptosis and their growing role as cancer targets. Cell Death Differ. 25, 27–36 (2018).
- Gross, A. & Katz, S. G. Non-apoptotic functions of BCL-2 family proteins. Cell Death Differ. 24, 1348–1358 (2017).
- 12. Strasser, A. & Vaux, D. L. Viewing BCL2 and cell death control from an evolutionary perspective. *Cell Death Differ.* **25**, 13–20 (2018).
- Charni, M., Aloni-Grinstein, R., Molchadsky, A. & Rotter, V. p53 on the crossroad between regeneration and cancer. Cell Death Differ. 24, 8–14 (2017).
- Engeland, K. Cell cycle arrest through indirect transcriptional repression by p53: I have a DREAM. Cell Death Differ. 25, 114–132 (2018).
- Furth, N., Aylon, Y. & Oren, M. p53 shades of Hippo. Cell Death Differ. 25, 81–92 (2018).
- Furth, N. & Aylon, Y. The LATS1 and LATS2 tumor suppressors: beyond the Hippo pathway. Cell Death Differ. 24, 1488–1501 (2017).
- Huttenhofer, A., Schattner, P. & Polacek, N. Non-coding RNAs: hope or hype? Trends Genet. 21, 289–297 (2005).
- Chen, L. L. The biogenesis and emerging roles of circular RNAs. Nat. Rev. Mol. Cell Biol. 17, 205–211 (2016).
- Galluzzi, L. et al. Molecular mechanisms of cell death: recommendations of the Nomenclature Committee on Cell Death 2018. Cell Death Differ. 25, 486–541 (2018).
- Flynn, R. A. & Chang, H. Y. Long noncoding RNAs in cell-fate programming and reprogramming. Cell Stem Cell 14, 752–761 (2014).
- Moran, V. A., Perera, R. J. & Khalil, A. M. Emerging functional and mechanistic paradigms of mammalian long non-coding RNAs. *Nucleic Acids Res.* 40, 6391–6400 (2012).
- Aberdam, D., Candi, E., Knight, R. A. & Melino, G. miRNAs, 'stemness' and skin. Trends Biochem. Sci. 33, 583–591 (2008).
- 23. Mancini, M. et al. MicroRNAs in human skin ageing. *Ageing Res. Rev.* 17, 9–15 (2014).
- Yi, R., Poy, M. N., Stoffel, M. & Fuchs, E. A skin microRNA promotes differentiation by repressing 'stemness'. *Nature* 452, 225–229 (2008).
- Amelio, I. et al. miR-24 triggers epidermal differentiation by controlling actin adhesion and cell migration. J. Cell Biol. 199, 347–363 (2012).
- Viticchie, G. et al. MicroRNA-203 contributes to skin re-epithelialization. Cell Death Dis. 3, e435 (2012).
- Tucci, P. et al. Loss of p63 and its microRNA-205 target results in enhanced cell migration and metastasis in prostate cancer. Proc. Natl Acad. Sci. USA 109, 15312–15317 (2012).
- Gregory, P. A. et al. The miR-200 family and miR-205 regulate epithelial to mesenchymal transition by targeting ZEB1 and SIP1. Nat. Cell Biol. 10, 593–601 (2008).
- Lena, A. M. et al. miR-203 represses 'stemness' by repressing DeltaNp63. Cell Death Differ. 15, 1187–1195 (2008).
- Bommer, G. T. et al. p53-mediated activation of miRNA34 candidate tumorsuppressor genes. Curr. Biol. 17, 1298–1307 (2007).
- Hurteau, G. J., Carlson, J. A., Spivack, S. D. & Brock, G. J. Overexpression of the microRNA hsa-miR-200c leads to reduced expression of transcription factor 8 and increased expression of E-cadherin. *Cancer Res.* 67, 7972–7976 (2007).
- de Pedro, I., Alonso-Lecue, P., Sanz-Gomez, N., Freije, A. & Gandarillas, A. Sublethal UV irradiation induces squamous differentiation via a p53independent, DNA damage-mitosis checkpoint. *Cell Death Dis.* 9, 1094 (2018)
- 33. Messenger, Z. J. et al. C/EBPbeta deletion in oncogenic Ras skin tumors is a synthetic lethal event. *Cell Death Dis.* **9**, 1054 (2018).
- Singh, T. P., Vieyra-Garcia, P. A., Wagner, K., Penninger, J. & Wolf, P. Cbl-b deficiency provides protection against UVB-induced skin damage by modulating inflammatory gene signature. *Cell Death Dis.* 9, 835 (2018).
- Bigas, J., Sevilla, L. M., Carceller, E., Boix, J. & Perez, P. Epidermal glucocorticoid and mineralocorticoid receptors act cooperatively to regulate epidermal development and counteract skin inflammation. *Cell Death Dis.* 9, 588 (2018).
- Xue, Z. et al. lincRNA-Cox2 regulates NLRP3 inflammasome and autophagy mediated neuroinflammation. Cell Death Differ. 26, 130–145 (2019).
- Kretz, M. et al. Suppression of progenitor differentiation requires the long noncoding RNA ANCR. Genes Dev. 26, 338–343 (2012).
- Kretz, M. et al. Control of somatic tissue differentiation by the long noncoding RNA TINCR. Nature 493, 231–235 (2013).

- Ziegler, C. et al. The long non-coding RNA LINC00941 and SPRR5 are novel regulators of human epidermal homeostasis. EMBO Rep. 20, https://doi.org/ 10.15252/embr.201846612 (2019).
- Panatta, E. et al. Long non-coding RNA uc.291 controls epithelial differentiation by interfering with the ACTL6A/BAF complex. EMBO Rep. 21, e46734 (2020).
- Roush, S. & Slack, F. J. The let-7 family of microRNAs. *Trends Cell Biol.* 18, 505–516 (2008).
- Zhang, L., Yang, Z., Huang, W. & Wu, J. H19 potentiates let-7 family expression through reducing PTBP1 binding to their precursors in cholestasis. *Cell Death Dis.* 10, 168 (2019).
- 43. Takamizawa, J. et al. Reduced expression of the let-7 microRNAs in human lung cancers in association with shortened postoperative survival. *Cancer Res.* **64**, 3753–3756 (2004).
- Shell, S. et al. Let-7 expression defines two differentiation stages of cancer. Proc. Natl Acad. Sci. USA 104, 11400–11405 (2007).
- Childs, G. et al. Low-level expression of microRNAs let-7d and miR-205 are prognostic markers of head and neck squamous cell carcinoma. Am. J. Pathol. 174, 736–745 (2009).
- Mizuno, R., Kawada, K. & Sakai, Y. The molecular basis and therapeutic potential of Let-7 microRNAs against colorectal cancer. *Can. J. Gastroenterol. Hepatol.* 2018, 5769591 (2018).
- Huang, Y., Hong, X., Hu, J. & Lu, Q. Targeted regulation of MiR-98 on E2F1 increases chemosensitivity of leukemia cells K562/A02. Onco Targets Ther. 10, 3233–3239 (2017).
- Trang, P. et al. Regression of murine lung tumors by the let-7 microRNA. Oncogene 29, 1580–1587 (2010).
- Pekarsky, Y., Balatti, V. & Croce, C. M. BCL2 and miR-15/16: from gene discovery to treatment. Cell Death Differ. 25, 21–26 (2018).
- Calin, G. A. et al. Frequent deletions and down-regulation of micro- RNA genes miR15 and miR16 at 13q14 in chronic lymphocytic leukemia. *Proc. Natl Acad. Sci. USA* 99, 15524–15529 (2002).
- Allegra, D. et al. Defective DROSHA processing contributes to downregulation of MiR-15/-16 in chronic lymphocytic leukemia. *Leukemia* 28, 98–107 (2014).
- Cimmino, A. et al. miR-15 and miR-16 induce apoptosis by targeting BCL2. Proc. Natl Acad. Sci. USA 102, 13944–13949 (2005).
- Teshima, K. et al. Dysregulation of BMI1 and microRNA-16 collaborate to enhance an anti-apoptotic potential in the side population of refractory mantle cell lymphoma. *Oncogene* 33, 2191–2203 (2014).
- Kalkavan, H. & Green, D. R. MOMP, cell suicide as a BCL-2 family business. Cell Death Differ. 25, 46–55 (2018).
- Montero, J. & Letai, A. Why do BCL-2 inhibitors work and where should we use them in the clinic? Cell Death Differ. 25, 56–64 (2018).
- Pihan, P., Carreras-Sureda, A. & Hetz, C. BCL-2 family: integrating stress responses at the ER to control cell demise. *Cell Death Differ.* 24, 1478–1487 (2017).
- 57. Opferman, J. T. & Kothari, A. Anti-apoptotic BCL-2 family members in development. *Cell Death Differ.* **25**, 37–45 (2018).
- Reid, G. et al. Restoring expression of miR-16: a novel approach to therapy for malignant pleural mesothelioma. Ann. Oncol. 24, 3128–3135 (2013).
- Chen, S. S. et al. Resistin facilitates VEGF-A-dependent angiogenesis by inhibiting miR-16-5p in human chondrosarcoma cells. *Cell Death Dis.* 10, 31 (2019)
- Klein, U. et al. The DLEU2/miR-15a/16-1 cluster controls B cell proliferation and its deletion leads to chronic lymphocytic leukemia. Cancer Cell 17, 28–40 (2010).
- Raveche, E. S. et al. Abnormal microRNA-16 locus with synteny to human 13q14 linked to CLL in NZB mice. Blood 109, 5079–5086 (2007).
- Henrich, K. O., Schwab, M. & Westermann, F. 1p36 tumor suppression-a matter of dosage? *Cancer Res.* 72, 6079–6088 (2012).
- Agostini, M. & Knight, R. A. miR-34: from bench to bedside. *Oncotarget* 5, 872–881 (2014).
- Li, F. et al. MALAT1 regulates miR-34a expression in melanoma cells. Cell Death Dis. 10, 389 (2019).
- Laake, K. et al. Loss of heterozygosity at 11q23.1 in breast carcinomas: indication for involvement of a gene distal and close to ATM. Genes Chromosomes Cancer 18, 175–180 (1997).
- Hart, M. et al. miR-34a: a new player in the regulation of T cell function by modulation of NF-kappaB signaling. *Cell Death Dis.* 10, 46 (2019).

- Chang, T. C. et al. Transactivation of miR-34a by p53 broadly influences gene expression and promotes apoptosis. Mol. Cell 26, 745–752 (2007).
- Pigazzi, M., Manara, E., Baron, E. & Basso, G. miR-34b targets cyclic AMPresponsive element binding protein in acute myeloid leukemia. *Cancer Res.* 69, 2471–2478 (2009).
- Chen, Q. R. et al. Systematic proteome analysis identifies transcription factor YY1 as a direct target of miR-34a. J. Proteome Res. 10, 479–487 (2011).
- Shen, Z. et al. MicroRNA-34a affects the occurrence of laryngeal squamous cell carcinoma by targeting the antiapoptotic gene survivin. *Med Oncol.* 29, 2473–2480 (2012).
- Kale, J., Osterlund, E. J. & Andrews, D. W. BCL-2 family proteins: changing partners in the dance towards death. *Cell Death Differ.* 25, 65–80 (2018).
- Beg, M. S. et al. Phase I study of MRX34, a liposomal miR-34a mimic, administered twice weekly in patients with advanced solid tumors. *Invest. N. Drugs* 35, 180–188 (2017).
- Le Beau, M. M. et al. Cytogenetic and molecular delineation of a region of chromosome 7 commonly deleted in malignant myeloid diseases. *Blood* 88, 1930–1935 (1996).
- 74. Kwon, J. J., Factora, T. D., Dey, S. & Kota, J. A systematic review of miR-29 in cancer. *Mol. Ther. Oncolytics* **12**. 173–194 (2019).
- Mott, J. L., Kobayashi, S., Bronk, S. F. & Gores, G. J. mir-29 regulates Mcl-1 protein expression and apoptosis. Oncogene 26, 6133–6140 (2007).
- Garzon, R. et al. MicroRNA 29b functions in acute myeloid leukemia. Blood 114, 5331–5341 (2009).
- Wu, H. et al. miR-29c-3p regulates DNMT3B and LATS1 methylation to inhibit tumor progression in hepatocellular carcinoma. Cell Death Dis. 10, 48 (2019).
- Ngankeu, A. et al. Discovery and functional implications of a miR-29b-1/miR-29a cluster polymorphism in acute myeloid leukemia. *Oncotarget* 9, 4354–4365 (2018).
- Calin, G. A. et al. Human microRNA genes are frequently located at fragile sites and genomic regions involved in cancers. Proc. Natl Acad. Sci. USA 101, 2999–3004 (2004).
- Iorio, M. V. et al. MicroRNA gene expression deregulation in human breast cancer. Cancer Res. 65, 7065–7070 (2005).
- 81. Seike, M. et al. MiR-21 is an EGFR-regulated anti-apoptotic factor in lung cancer in never-smokers. *Proc. Natl Acad. Sci. USA* **106**, 12085–12090 (2009).
- Volinia, S. et al. A microRNA expression signature of human solid tumors defines cancer gene targets. Proc. Natl Acad. Sci. USA 103, 2257–2261 (2006).
- Xue, Z. et al. miR-21 promotes NLRP3 inflammasome activation to mediate pyroptosis and endotoxic shock. *Cell Death Dis.* 10, 461 (2019).
- Hatley, M. E. et al. Modulation of K-Ras-dependent lung tumorigenesis by MicroRNA-21. Cancer Cell 18, 282–293 (2010).
- Medina, P. P., Nolde, M. & Slack, F. J. OncomiR addiction in an in vivo model of microRNA-21-induced pre-B-cell lymphoma. *Nature* 467, 86–90 (2010).
- Rodriguez, A. et al. Requirement of bic/microRNA-155 for normal immune function. Science 316, 608–611 (2007).
- Baltimore, D., Boldin, M. P., O'Connell, R. M., Rao, D. S. & Taganov, K. D. MicroRNAs: new regulators of immune cell development and function. *Nat. Immunol.* 9, 839–845 (2008).
- Eis, P. S. et al. Accumulation of miR-155 and BIC RNA in human B cell lymphomas. Proc. Natl Acad. Sci. USA 102, 3627–3632 (2005).
- Kluiver, J. et al. BIC and miR-155 are highly expressed in Hodgkin, primary mediastinal and diffuse large B cell lymphomas. J. Pathol. 207, 243–249 (2005).
- Kluiver, J. et al. Lack of BIC and microRNA miR-155 expression in primary cases of Burkitt lymphoma. Genes Chromosomes Cancer 45, 147–153 (2006).
- Costinean, S. et al. Src homology 2 domain-containing inositol-5phosphatase and CCAAT enhancer-binding protein beta are targeted by miR-155 in B cells of Emicro-MiR-155 transgenic mice. *Blood* 114, 1374–1382 (2009).
- O'Connell, R. M., Chaudhuri, A. A., Rao, D. S. & Baltimore, D. Inositol phosphatase SHIP1 is a primary target of miR-155. *Proc. Natl Acad. Sci. USA* 106, 7113–7118 (2009).
- 93. Wan, J. et al. Inhibition of miR-155 reduces impaired autophagy and improves prognosis in an experimental pancreatitis mouse model. *Cell Death Dis.* **10**, 303 (2019).
- Cheng, C. J. et al. MicroRNA silencing for cancer therapy targeted to the tumour microenvironment. Nature 518, 107–110 (2015).
- Lee, E. J. et al. Expression profiling identifies microRNA signature in pancreatic cancer. Int. J. Cancer 120, 1046–1054 (2007).

- Braconi, C. & Patel, T. MicroRNA expression profiling: a molecular tool for defining the phenotype of hepatocellular tumors. *Hepatology* 47, 1807–1809 (2008).
- Pineau, P. et al. miR-221 overexpression contributes to liver tumorigenesis. Proc. Natl Acad. Sci. USA 107, 264–269 (2010).
- 98. Park, J. K. et al. miR-221 silencing blocks hepatocellular carcinoma and promotes survival. *Cancer Res.* **71**, 7608–7616 (2011).
- Liu, X. et al. MicroRNA-222 regulates cell invasion by targeting matrix metalloproteinase 1 (MMP1) and manganese superoxide dismutase 2 (SOD2) in tongue squamous cell carcinoma cell lines. *Cancer Genomics Proteom.* 6, 131–139 (2009).
- Wilusz, J. E., Sunwoo, H. & Spector, D. L. Long noncoding RNAs: functional surprises from the RNA world. Genes Dev. 23, 1494–1504 (2009).
- Pickard, M. R. & Williams, G. T. Molecular and cellular mechanisms of action of tumour suppressor GAS5 LncRNA. Genes 6, 484–499 (2015).
- Hudson, W. H. et al. Conserved sequence-specific lincRNA-steroid receptor interactions drive transcriptional repression and direct cell fate. *Nat. Commun.* 5, 5395 (2014).
- Mourtada-Maarabouni, M., Pickard, M. R., Hedge, V. L., Farzaneh, F. & Williams, G. T. GASS, a non-protein-coding RNA, controls apoptosis and is downregulated in breast cancer. *Oncogene* 28, 195–208 (2009).
- Kino, T., Hurt, D. E., Ichijo, T., Nader, N. & Chrousos, G. P. Noncoding RNA gas5 is a growth arrest- and starvation-associated repressor of the glucocorticoid receptor. Sci. Signal 3, ra8 (2010).
- Zhou, Y., Zhang, X. & Klibanski, A. MEG3 noncoding RNA: a tumor suppressor.
 J. Mol. Endocrinol. 48, R45–R53 (2012).
- Lu, K. H. et al. Long non-coding RNA MEG3 inhibits NSCLC cells proliferation and induces apoptosis by affecting p53 expression. *BMC Cancer* 13, 461 (2013).
- Gailhouste, L. et al. MEG3-derived miR-493-5p overcomes the oncogenic feature of IGF2-miR-483 loss of imprinting in hepatic cancer cells. *Cell Death Dis.* 10, 553 (2019).
- Zhou, Y. et al. Activation of p53 by MEG3 non-coding RNA. J. Biol. Chem. 282, 24731–24742 (2007).
- Wu, D. & Prives, C. Relevance of the p53-MDM2 axis to aging. *Cell Death Differ*.
 169–179 (2018).
- Sullivan, K. D., Galbraith, M. D., Andrysik, Z. & Espinosa, J. M. Mechanisms of transcriptional regulation by p53. Cell Death Differ. 25, 133–143 (2018).
- Lu, Z. et al. Long non-coding RNA NKILA inhibits migration and invasion of non-small cell lung cancer via NF-kappaB/Snail pathway. J. Exp. Clin. Cancer Res. 36, 54 (2017).
- Liu, B. et al. A cytoplasmic NF-kappaB interacting long noncoding RNA blocks lkappaB phosphorylation and suppresses breast cancer metastasis. Cancer Cell 27, 370–381 (2015).
- Huang, D. et al. NKILA IncRNA promotes tumor immune evasion by sensitizing T cells to activation-induced cell death. *Nat. Immunol.* 19, 1112–1125 (2018).
- Adriaens, C. et al. p53 induces formation of NEAT1 IncRNA-containing paraspeckles that modulate replication stress response and chemosensitivity. Nat. Med. 22, 861–868 (2016).
- Mello, S. S. et al. Neat1 is a p53-inducible lincRNA essential for transformation suppression. Genes Dev. 31, 1095–1108 (2017).
- Blume, C. J. et al. p53-dependent non-coding RNA networks in chronic lymphocytic leukemia. *Leukemia* 29, 2015–2023 (2015).
- 117. Wu, Y. et al. Nuclear-enriched abundant transcript 1 as a diagnostic and prognostic biomarker in colorectal cancer. *Mol. Cancer* **14**, 191 (2015).
- Ma, Y. et al. Enhanced expression of long non-coding RNA NEAT1 is associated with the progression of gastric adenocarcinomas. World J. Surg. Oncol. 14, 41 (2016)
- Wang, P. et al. Long noncoding RNA NEAT1 promotes laryngeal squamous cell cancer through regulating miR-107/CDK6 pathway. J. Exp. Clin. Cancer Res. 35, 22 (2016).
- Shin, V. Y. et al. Long non-coding RNA NEAT1 confers oncogenic role in triple-negative breast cancer through modulating chemoresistance and cancer stemness. *Cell Death Dis.* 10, 270 (2019).
- Ozawa, T. et al. CCAT1 and CCAT2 long noncoding RNAs, located within the 8q.24.21 'gene desert', serve as important prognostic biomarkers in colorectal cancer. Ann. Oncol. 28, 1882–1888 (2017).

- Alaiyan, B. et al. Differential expression of colon cancer associated transcript1 (CCAT1) along the colonic adenoma-carcinoma sequence. BMC Cancer 13, 196 (2013).
- Jiang, Y. et al. Co-activation of super-enhancer-driven CCAT1 by TP63 and SOX2 promotes squamous cancer progression. Nat. Commun. 9, 3619 (2018).
- McCleland, M. L. et al. CCAT1 is an enhancer-templated RNA that predicts BET sensitivity in colorectal cancer. J. Clin. Invest. 126, 639–652 (2016).
- Zhong, X., Hu, X. & Zhang, L. Oncogenic long noncoding RNA FAL1 in human cancer. Mol. Cell Oncol. 2, e977154 (2015).
- Jeong, S. et al. Relationship of focally amplified long noncoding on chromosome 1 (FAL1) IncRNA with E2F transcription factors in thyroid cancer. Medicine 95, e2592 (2016).
- Pan, C. et al. Long noncoding RNA FAL1 promotes cell proliferation, invasion and epithelial-mesenchymal transition through the PTEN/AKT signaling axis in non-small cell lung cancer. Cell Physiol. Biochem. 43, 339–352 (2017).
- Hu, X. et al. A functional genomic approach identifies FAL1 as an oncogenic long noncoding RNA that associates with BMI1 and represses p21 expression in cancer. Cancer Cell 26, 344–357 (2014).
- Tseng, Y. Y. et al. PVT1 dependence in cancer with MYC copy-number increase. Nature 512, 82–86 (2014).
- Popescu, N. C. & Zimonjic, D. B. Chromosome-mediated alterations of the MYC gene in human cancer. J. Cell Mol. Med. 6, 151–159 (2002).
- Guan, Y. et al. Amplification of PVT1 contributes to the pathophysiology of ovarian and breast cancer. Clin. Cancer Res. 13, 5745–5755 (2007).
- He, Y. et al. Long non-coding RNA PVT1 predicts poor prognosis and induces radioresistance by regulating DNA repair and cell apoptosis in nasopharyngeal carcinoma. Cell Death Dis. 9, 235 (2018).
- Mukherjee, A. & Williams, D. W. More alive than dead: non-apoptotic roles for caspases in neuronal development, plasticity and disease. *Cell Death Differ.* 24. 1411–1421 (2017).
- Cocquerelle, C., Mascrez, B., Hetuin, D. & Bailleul, B. Mis-splicing yields circular RNA molecules. FASEB J. 7, 155–160 (1993).
- 135. Barrett, S. P., Wang, P. L. & Salzman, J. Circular RNA biogenesis can proceed through an exon-containing lariat precursor. *eLife* **4**, e07540 (2015).
- Hansen, T. B. et al. Natural RNA circles function as efficient microRNA sponges. Nature 495, 384–388 (2013).
- Memczak, S. et al. Circular RNAs are a large class of animal RNAs with regulatory potency. *Nature* 495, 333–338 (2013).
- 138. Vo, J. N. et al. The landscape of circular RNA in cancer. *Cell* **176**, 869–881 e813 (2019).
- Xiong, D. D. et al. High throughput circRNA sequencing analysis reveals novel insights into the mechanism of nitidine chloride against hepatocellular carcinoma. *Cell Death Dis.* 10, 658 (2019).
- Chen, S. et al. Widespread and functional RNA circularization in localized prostate cancer. Cell 176, 831–843 e822 (2019).
- Kristensen, L. S., Hansen, T. B., Veno, M. T. & Kjems, J. Circular RNAs in cancer: opportunities and challenges in the field. *Oncogene* 37, 555–565 (2018).
- Liu, Z. et al. CircRNA-5692 inhibits the progression of hepatocellular carcinoma by sponging miR-328-5p to enhance DAB2IP expression. *Cell Death Dis.* 10, 900 (2019).
- Gao, L. et al. CircCDR1as upregulates autophagy under hypoxia to promote tumor cell survival via AKT/ERK(1/2)/mTOR signaling pathways in oral squamous cell carcinomas. Cell Death Dis. 10, 745 (2019).
- 144. Yang, W., Du, W. W., Li, X., Yee, A. J. & Yang, B. B. Foxo3 activity promoted by non-coding effects of circular RNA and Foxo3 pseudogene in the inhibition of tumor growth and angiogenesis. *Oncogene* 35, 3919–3931 (2016)
- Zhang, Y., Zhao, H. & Zhang, L. Identification of the tumorsuppressive function of circular RNA FOXO3 in nonsmall cell lung cancer through sponging miR155. Mol. Med. Rep. 17, 7692–7700 (2018).
- Lu, W. Y. Roles of the circular RNA circ-Foxo3 in breast cancer progression. Cell Cycle 16, 589–590 (2017).
- Du, W. W. et al. Induction of tumor apoptosis through a circular RNA enhancing Foxo3 activity. Cell Death Differ. 24, 357–370 (2017).
- Anfossi, S., Babayan, A., Pantel, K. & Calin, G. A. Clinical utility of circulating non-coding RNAs - an update. Nat. Rev. Clin. Oncol. 15, 541–563 (2018).
- Stein, C. A. & Castanotto, D. FDA-approved oligonucleotide therapies in 2017.
 Mol. Ther. 25, 1069–1075 (2017).