Machine learning for target discovery in drug development

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Abstract

The discovery of macromolecular targets for bioactive agents is currently a bottleneck for the informed design of chemical probes and drug leads. Typically, activity profiling against genetically-manipulated cell lines or chemical proteomics is pursued to shed light on their biology and deconvolute drug-target networks. By taking advantage of the ever-growing wealth of publicly available bioactivity data, learning algorithms now provide an attractive means to generate statistically motivated research hypotheses and thereby prioritize biochemical screens. Here we highlight recent successes in machine intelligence for target identification and discuss challenges and opportunities for drug discovery.

Introduction

The future of molecular medicine relies on the identification and validation of small molecule effectors [1-4] despite the advent of biologics [2,5], such as antibody–drug conjugates. Indeed, specific drug-target binding and engagement remains a hallmark for modulation and treatment of diseases. Bioactive matter is rarely selective for a given target, but rather engages several other related or unrelated macromolecules [6,7], through what has been coined as polypharmacology or network pharmacology [8,9]. Although in some circumstances, such as cancer, it is desirable to harness the potential of therapeutics that modulate a plethora of targets – ideally belonging to distinct signalling pathways [10] – generally, polypharmacology is unwanted because it is responsible for adverse drug reactions [11,12]. Therefore, factual knowledge of on- and off-targets correlated to efficacy and liabilities of chemical matter is of utmost importance to maximize benefits and mitigate attrition in development pipelines.

The identification and unravelling of pharmacology networks for phenotypic screening hits – either of synthetic or natural origin – arguably remains a bottleneck of modern drug discovery [13-15]. This is a consequence of our current inability to streamline state-of-the-art technologies for target identification, *e.g.* chemical proteomics. Nonetheless, advances in machine-intelligence heuristics and hardware, and the increasing amount of publicly available chemical-biology data may afford untapped opportunities to speed up discovery programs that to date have stalled at the target identification phase [15]. In this Review, we discuss recent progress made in machine learning (for an overview on basic concepts we refer to Refs. [16] and [17]) as a research hypothesis generator and as a vital method in the chemical biology toolbox for research into drug-target recognition. In addition, we highlight advantages of machine learning based technologies and outline gaps in our knowledge to hopefully spur on future investigations and democratize its use among chemical biologists.

A winding road to drug target identification

Chemical proteomic approaches remain the gold-standard for the identification of macromolecular counterparts for small bioactive molecules [18-22]. These methods require the chemical modification of the ligand of interest by appending a “tag” moiety that will afterwards enable a pull down of the formed ligand–target complexes. Downstream identification of the bound proteins can then be made by mass spectrometry or bespoke analytical methods [20]. By application of this concept, Cravatt and co-workers were able to identify the mitochondrial carnitine–acylcarnitine translocase SLC25A20 as a functional target of diterpenoid ester ingerol mebutate (**1**), which is a first-in-class treatment for actinic keratosis (Figure 1a) [23]. Other prominent examples include the identification of carbamates as arylacetamide deacetylase-like 1 regulators [24], and a clinical-stage imidazole as a promiscuous lipid hydrolases’ inhibitor [25]. Although these studies clearly suggest that chemical proteomics lends itself to scrutinizing drug–target relationships, it becomes apparent that chemical manipulation of the native ligand can disrupt the binding affinity towards relevant on-

and off-targets. Moreover, chemical proteomics rarely enables the identification of membrane proteins due to their instability in solution and low copy number [26].

As an alternative, screening of the native ligand against a battery of genetically defined cell lines, may offer a solution to identify drug targets based on the observed activity signatures. Indeed, this method, has enabled the identification of transient receptor potential canonical channels 4 and 5 as targets for anti-cancer sesquiterpene (–)-englerin A (**2**) by correlating gene expression with phenotypic changes (Figure 1b) [26,27]. However, like chemical proteomics, this approach is laborious, time consuming and expensive, which has prompted the search for viable alternatives, such as machine learning.

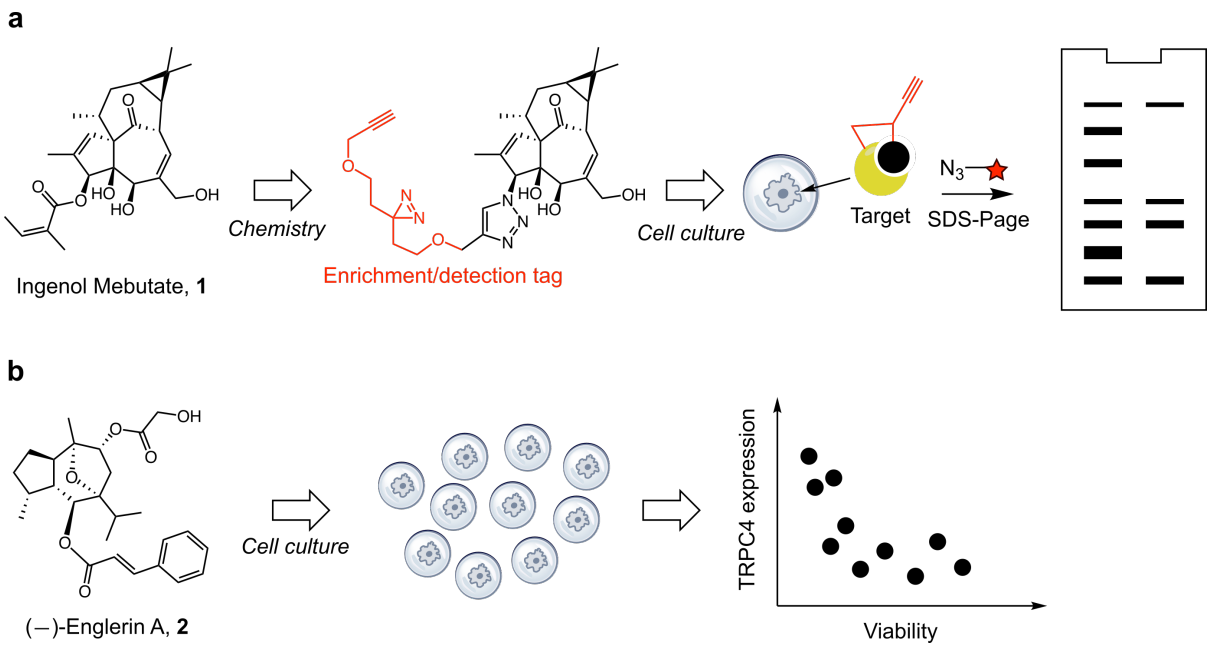

Figure 1. Schematics of two different methods for target identification. a) Identification of targets for ingenol mebutate (**1**) through a chemical proteomics approach. b) Identification of targets for (–)-englerin A (**2**) by means of a phenotypic screen approach.

Machine learning for target identification

With the advent of high-throughput experimentation, a wealth of chemical and biological data has been generated [16,28,29]. Thus, it became impossible for researchers to efficiently analyse all available information and became reasonable to assume that computer algorithms could be employed to sieve through this data to identify latent patterns, which expert human researchers may struggle to identify [30,31]. Indeed, machine learning has recently seen a range of applications in biology, medicine [32–38], chemistry [39–41], and materials science [42–44], which suggests that it can be used to speed up development pipelines and augment human

perception. For example, deep-learning algorithms have been devised to predict retrosynthetic pathways to molecules of interest [45] and design new chemical entities that can be scrutinized as hits/drug leads [46–48]. Also, different learning heuristics that leverage chemical structure data have been implemented to identify drug–target interactions that can be exploited in pre-clinical studies [49,50] and to design experiments [51,52].

The self-organizing maps (SOM)-based prediction of drug equivalence relationships (SPiDER) software uses a neural network-inspired algorithm to discretise the input feature vector onto a so-called feature map in an unsupervised fashion [6]. In practice, this means that drug–target relationships are inferred based on descriptor similarity to reference ligands in the same neuron without explicitly considering the target identity of those reference ligands for the purpose of training or heuristics' validation studies. The method employs a set of topological pharmacophore descriptors (CATS2 [53]) to categorize non-hydrogen atoms. From the autocorrelation of those topological feature pairs in a molecule, a SOM is built. A second, independent SOM is built by using physicochemical descriptors prior to generating a consensus prediction from both SOMs, which is supported by background statistics. Taken together, the prime goals are to employ descriptors that represent molecules in a sufficiently fuzzy manner, and analyse data from disparate vantage points to allow generalization of the method to previously unseen test cases, i.e. molecules of potential biological interest. The software tool has been extensively applied to *de novo* designed entities and, especially, natural products of high biological value. Prominent use cases of SPiDER include identification of farnesoid X receptor (EC_{50} = 0.2 μM), peroxisome proliferator-activated receptor gamma (EC_{50} = 8 μM), 5-lipoxygenase (EC_{50} = 11 μM) and microsomal prostaglandin E synthase-1 (EC_{50} = 8 μM) as drug targets for macrolide archazolid A (**3**, Figure 2). Importantly, given the structural difference between **3** and the small molecules used as reference structures by SPiDER, target inference was successfully achieved by exploiting synthetically motivated fragments of **3** as bioactivity blueprints [54]. This appears to be a reasonable approach as an identical strategy was followed to identify the prostanoid 3 receptor (EC_{50} = 6 nM) and the voltage-gated $Ca_v1.2$ channel (IC_{50} = 6 μM) as targets for doliculide, **4**, and **2**, respectively [55,56]. Similarly, fragment-like alkaloids graveolinine (**5**), isomacroin (**6**), and piperlongumine (**7**) could be de-orphanized with SPiDER as serotonin 2B receptor (IC_{50} = 12 μM), platelet-derived growth factor receptor alpha (IC_{50} = 25 μM), and transient receptor potential channel vanilloid 2 (EC_{50} = 5 μM) modulators, respectively, which opens new avenues for molecular optimization in hit-to-lead programs [49,57]. More recently, SPiDER has been applied to prioritize *de novo* designed chemical entities fitting a predefined pharmacological profile for synthesis and experimental validation [46,58]. Given that SPiDER is built from two-ddimensional descriptors, one may expect a higher rate of false positive predictions for molecules with stereogenic centres relative to achiral molecules, as this information is not taken into account. To better predict the chiral nature of molecular recognition, the extended three-

dimensional fingerprint was recently developed and applied to synthetic molecules [59], but remains to be validated with complex natural products.

The target inference generator (TIGER) method offers target predictions based on SOMs — similar to SPiDER — but differs in the scoring function and the use of a restricted number of topological pharmacophore atom types [60,61]. For flat and neutral natural products, such as resveratrol (**8**), and (±)-marinopyrrole A (**9**), the method was able to confidently predict relationships with targets from different families, pinpointing the polypharmacology that underlies these structures [61,62]. For example, **8** was confirmed as an estrogen receptor ligand (K_i = 0.4–4 μM), whereas **9** was successfully associated to orexin (K_B = 0.3–0.6 μM) and glucocorticoid (K_B = 0.7 μM) receptors, and trypsin (IC_{50} = 3 μM).

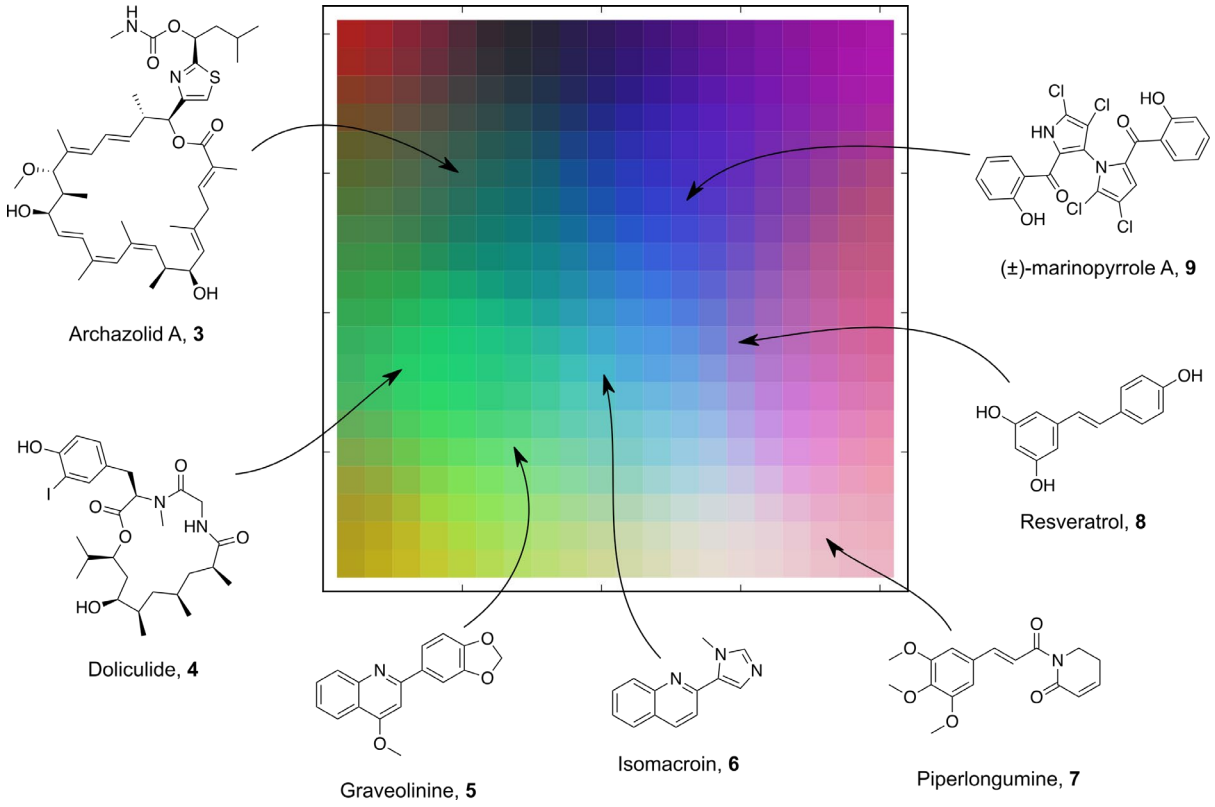

Figure 2. Application of self-organizing maps (SOM) for target deconvolution of phenotypic screen hits. SPiDER and TIGER use the SOM technology to tessellate chemical space and infer targets for bioactive molecules. SPiDER was used to identify targets for archazolid A, doliculide, graveolinine, isomacroin and piperlongumine, whereas TIGER was used for resveratrol and (±)-marinopyrrole A.

The heuristics learn the data structure in an unsupervised way to aggregate molecules with similar feature patterns, yet without knowing if the resulting association is correct in respect to drug target associations. This is frequently assessed through cross-validation studies, hold out test data [63] and adversarial control models [64], as a

means of ascertaining the relevance of the obtained models and soundness of the exploited descriptors. Regression-based methods can circumvent some of the limitations of unsupervised and classification methods, by affording a predicted affinity value. This however comes at the expense of needing a larger volume of training data, which is often not in hand or is expensive to collect. The molecular ant algorithm (MAntA) [65-67] is a small molecule generator workflow that implements Gaussian process regression to predict affinity values for query ligands and model binding uncertainties towards a range of ChEMBL targets. Thus, the method is well suited to not only prioritize *de novo* designed entities [66], but also repurpose molecules [68] or de-risk pre-clinical development [69]. The application of other conceptually distinct regression algorithms is viable [50,51] and one can also expect the growing utility of deep-learning architectures in the target prediction space in the near future [70–72].

The DEcRyPT method has been recently reported [73,74] for the prediction of network pharmacology, either as a standalone tool or in combination with SPiDER, by employing random forest technology and CATS2 descriptors. In short, this method harnesses a user-defined number of predictors (decision trees) that independently analyse different portions of the training data, prior to aggregation of the resulting predicted outputs. When applied to beta-lapachone (**10**), the workflow confidently predicted 5-lipoxygenase as a target. Follow-up studies revealed that **10** was a reversible, allosteric modulator of 5-lipoxygenase (IC_{50} = 240 nM) and also a selective relative to other lipoxygenases and metalloenzymes. Activity of **10** could only be observed in cell-free and cell-based assays in the presence of a reducing agent. The observation suggests that beta-lapachone is reduced *in situ* to its hydroquinone form prior to modulation of 5-lipoxygenase. Importantly, modulation of 5-lipoxygenase by **10** was crucial for the anti-proliferative activity of **10** in an acute myeloid leukemia cell line [73]. More recently, a second use case unveiled modulation of cannabinoid receptors by celastrol (**11**), which are also implicated in cancer progression [74].

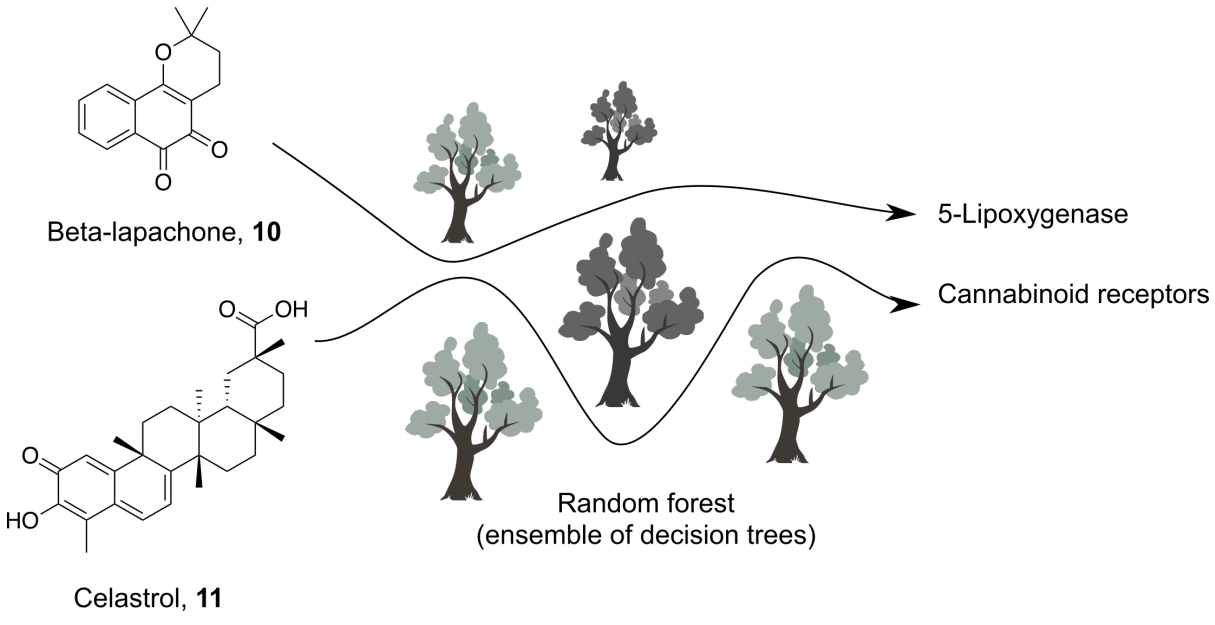

Figure 3. Schematics of target identification for beta-lapachone (**10**), and celastrol (**11**) by using the random forest technology implemented in DEcRyPT.

Outlook

Machine intelligence has recently seen an upsurge of applications in chemical sciences, in particular referring to the design of new chemical entities. The deconvolution of targets for phenotypic screen hits has been a necessary but laborious task [13], which enables the rational design and optimization of chemical matter towards drug leads. Over the years, the large volume of data collected [75] now allows scientists working at the interface of chemistry and biology, and equipped with machine learning tools to identify latent patterns worthy of further research. Here, we have shown recent successes in the identification of targets, leveraged by statistical learning algorithms. Naturally, each technique has its own strengths and limitations, and domain of applicability, all of which require careful consideration. For example, none of the discussed methods can identify unreported proteins as targets, as these methods are based on the principle that similar ligands (irrespective of the employed descriptor) exert similar biological effects. Similarly, the identification of DNA/RNA binders, modulators of protein–lipid interactions, among others ought to be possible provided that sufficient data is available; this is however not easily accessible, contributing to a preferential exploration of protein targets. One must also bear in mind that no new biology can be uncovered through machine learning, unlike wet laboratory experiments that are able to provide the ground truth. Even for known proteins, there is shortage of ligand data in some cases, which can limit the scope of a new method. Erroneous predictions are also common, especially for poorly studied targets or targets with noisy data available. However, negative, yet validated results offer opportunities to build better-informed machine intelligence, and thus should still be reported and not discarded. Alongside healthy scepticism, machine learning for target identification entails an important set of tools to aid decision-making. By filling a gap within the chemical biologists toolbox, we expect machine intelligence to speed up some tasks in drug discovery towards the development of life-changing therapeutics.

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** Reference work for a database annotating ligand-receptor relationships, which can be curated and used for building bespoke machine learning methods.