

Identification of novel susceptibility loci and genes for breast cancer risk: A transcriptome-wide association study of 229,000 women of European descent

Lang Wu^{1,160}, Wei Shi^{2,160}, Jirong Long¹, Xingyi Guo¹, Kyriaki Michailidou^{3,4}, Jonathan Beesley², Manjeet K. Bolla³, Xiao-Ou Shu¹, Yingchang Lu¹, Qiuyin Cai¹, Fares Al-Ejeh², Esdy Rozali², Qin Wang³, Joe Dennis³, Bingshan Li¹⁵¹, Chenjie Zeng¹, Helian Feng^{5,6}, Alexander Gusev^{153, 154, 155}, Richard T. Barfield⁵, Irene L. Andrulis^{7,8}, Hoda Anton-Culver⁹, Volker Arndt¹⁰, Kristan J. Aronson¹¹, Paul L. Auer^{12,13}, Myrto Barrdahl¹⁴, Caroline Baynes¹⁵, Matthias W. Beckmann¹⁶, Javier Benitez^{17,18}, Marina Bermisheva^{19,20}, Carl Blomqvist^{21,159}, Natalia V. Bogdanova^{20,22,23}, Stig E. Bojesen²⁴⁻²⁶, Hiltrud Brauch²⁷⁻²⁹, Hermann Brenner^{10,29,30}, Louise Brinton³¹, Per Broberg³², Sara Y. Brucker³³, Barbara Burwinkel^{34,35}, Trinidad Caldés³⁶, Federico Canzian³⁷, Brian D. Carter³⁸, J. Esteban Castelao³⁹, Jenny Chang-Claude^{14,40}, Xiaoqing Chen², Ting-Yuan David Cheng⁴¹, Hans Christiansen²², Christine L. Clarke⁴², NBCS Collaborators^{43-120,44-124,45}, Margriet Collée⁴⁶, Sten Cornelissen⁴⁷, Fergus J. Couch⁴⁸, David Cox^{49,50}, Angela Cox⁵¹, Simon S. Cross⁵², Julie M. Cunningham⁴⁸, Kamila Czene⁵³, Mary B. Daly⁵⁴, Peter Devilee^{55,56}, Kimberly F. Doheny⁵⁷, Thilo Dörk²⁰, Isabel dos-Santos-Silva⁵⁸, Martine Dumont⁵⁹, Miriam Dwek⁶⁰, Diana M. Eccles⁶¹, Ursula Eilber¹⁴, A. Heather Eliassen^{6,62}, Christoph Engel⁶³, Mikael Eriksson⁵³, Laura Fachal¹⁵, Peter A. Fasching^{16,64}, Jonine Figueroa^{31,65}, Dieter Flesch-Janys^{66,67}, Olivia Fletcher⁶⁸, Henrik Flyger⁶⁹, Lin Fritschi⁷⁰, Marike Gabrielson⁵³, Manuela Gago-Dominguez^{71,72}, Susan M. Gapstur³⁸, Montserrat García-Closas³¹, Mia M. Gaudet³⁸, Maya Ghoussaini¹⁵, Graham G. Giles^{73,74}, Mark S. Goldberg^{75,76}, David E. Goldgar⁷⁷, Anna González-Neira¹⁷, Pascal Guénel⁷⁸, Eric Hahnen⁷⁹⁻⁸¹, Christopher A. Haiman⁸², Niclas Håkansson⁸³, Per Hall⁵³, Emily Hallberg⁸⁴, Ute Hamann⁸⁵, Patricia Harrington¹⁵, Alexander Hein¹⁶, Belynda Hicks⁸⁶, Peter Hillemanns²⁰, Antoinette Hollestelle⁸⁷, Robert N. Hoover³¹, John L. Hopper⁷⁴, Guanmengqian Huang⁸⁵, Keith Humphreys⁵³, David J. Hunter^{6,158}, Anna Jakubowska⁸⁸, Wolfgang Janni⁸⁹, Esther M. John⁹⁰⁻⁹², Nichola Johnson⁶⁸, Kristine Jones⁸⁶, Michael E. Jones⁹³, Audrey Jung¹⁴, Rudolf Kaaks¹⁴, Michael J. Kerin⁹⁴, Elza Khusnutdinova^{19,95}, Veli-Matti Kosma⁹⁶⁻⁹⁸, Vessela N. Kristensen⁹⁹⁻¹⁰¹, Diether Lambrechts^{102,103}, Loic Le Marchand¹⁰⁴, Jingmei Li¹⁵⁷, Sara Lindström^{5,105}, Jolanta Lissowska¹⁰⁶, Wing-Yee Lo^{27,28}, Sibylle Loibl¹⁰⁷, Jan Lubinski⁸⁸, Craig Luccarini¹⁵, Michael P. Lux¹⁶, Robert J. MacInnis^{73,74}, Tom Maishman^{61,108}, Ivana Maleva Kostovska^{20,109}, Arto Mannermaa⁹⁶⁻⁹⁸, JoAnn E. Manson^{6,110}, Sara Margolin¹¹¹, Dimitrios Mavroudis¹¹², Hanne Meijers-Heijboer¹⁵², Alfons Meindl¹¹³, Usha Menon¹¹⁴, Jeffery Meyer⁴⁸, Anna Marie Mulligan^{115,116}, Susan L. Neuhausen¹¹⁷, Heli Nevanlinna¹¹⁸, Patrick Neven¹¹⁹, Sune F. Nielsen^{24,25}, Børge G. Nordestgaard²⁴⁻²⁶, Olufunmilayo I. Olopade¹²⁰, Janet E. Olson⁸⁴, Håkan Olsson³², Paolo Peterlongo¹²¹, Julian Peto⁵⁸, Dijana Plaseska-Karanfilska¹⁰⁹, Ross Prentice¹², Nadege Presneau⁶⁰, Katri Pyrkäs^{122,123}, Brigitte Rack⁸⁹, Paolo Radice¹²⁵, Nazneen Rahman¹²⁶, Gad Rennert¹²⁷, Hedy S. Rennert¹²⁷, Valerie Rhenius¹⁵, Atocha Romero^{36,128}, Jane Romm⁵⁷, Anja Rudolph¹⁴, Emmanouil Saloustros¹²⁹, Dale P. Sandler¹³⁰, Elinor J. Sawyer¹³¹, Marjanka K. Schmidt^{47,132}, Rita K. Schmutzler⁷⁹⁻⁸¹, Andreas Schneeweiss^{34,133}, Rodney J. Scott^{134,135}, Christopher Scott⁸⁴, Sheila Seal¹²⁶, Mitul Shah¹⁵, Martha J. Shrubsole¹, Ann Smeets¹¹⁹, Melissa C. Southey¹³⁶, John J. Spinelli^{137,138}, Jennifer Stone^{139,140}, Harald Surowy^{34,35}, Anthony J. Swerdlow^{93,141}, Rulla M. Tamimi^{5,6,62}, William Tapper⁶¹, Jack A. Taylor^{130,142}, Mary Beth Terry¹⁴³, Daniel C. Tessier¹⁴⁴, Abigail Thomas⁸⁴, Kathrin Thöne⁶⁷, Rob A.E.M. Tollenaar¹⁴⁵, Diana Torres^{85,146}, Thérèse Truong⁷⁸, Michael Untch¹⁴⁷, Celine Vachon⁸⁴, David Van Den Berg⁸², Daniel Vincent¹⁴⁴, Quinten Waisfisz¹⁵², Clarice R. Weinberg¹⁴⁸, Camilla Wendt¹¹¹, Alice S. Whittemore^{91,92}, Hans Wildiers¹¹⁹, Walter C.

Willett^{6,62,156}, Robert Winqvist^{122,123}, Alicja Wolk⁸³, Lucy Xia⁸², Xiaohong R. Yang³¹, Argýrios Ziogas⁹, Elad Ziv¹⁴⁹, kConFab/AOCS Investigators¹⁵⁰, Alison M. Dunning¹⁵, Paul D.P. Pharoah^{3,15}, Jacques Simard⁵⁹, Roger L. Milne^{73,74}, Stacey L. Edwards², Peter Kraft^{5,6}, Douglas F. Easton^{3,15}, Georgia Chenevix-Trench^{2*}, Wei Zheng^{1*}

***Corresponding Authors:** Wei Zheng, MD, PhD, Division of Epidemiology, Department of Medicine, Vanderbilt Epidemiology Center, Vanderbilt-Ingram Cancer Center, Vanderbilt University Medical Center, 2525 West End Ave, Suite 800, Nashville, Tennessee, 37203, USA. Email: wei.zheng@vanderbilt.edu and Georgia Chenevix-Trench, PhD, Cancer Division, QIMR Berghofer Medical Research Institute, 300 Herston Road, Herston 4006, Australia. Email: Georgia.Trench@qimrberghofer.edu.au

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1. Division of Epidemiology, Department of Medicine, Vanderbilt Epidemiology Center, Vanderbilt-Ingram Cancer Center, Vanderbilt University School of Medicine, Nashville, TN, USA.
2. Cancer Division, QIMR Berghofer Medical Research Institute, Brisbane, Australia.
3. Centre for Cancer Genetic Epidemiology, Department of Public Health and Primary Care, University of Cambridge, Cambridge, UK.
4. Department of Electron Microscopy/Molecular Pathology, The Cyprus Institute of Neurology and Genetics, Nicosia, Cyprus.
5. Program in Genetic Epidemiology and Statistical Genetics, Harvard T.H. Chan School of Public Health, Boston, MA, USA.
6. Department of Epidemiology, Harvard T.H. Chan School of Public Health, Boston, MA, USA.
7. Fred A. Litwin Center for Cancer Genetics, Lunenfeld-Tanenbaum Research Institute of Mount Sinai Hospital, Toronto, ON, Canada.
8. Department of Molecular Genetics, University of Toronto, Toronto, ON, Canada.
9. Department of Epidemiology, University of California Irvine, Irvine, CA, USA.
10. Division of Clinical Epidemiology and Aging Research, German Cancer Research Center (DKFZ), Heidelberg, Germany.
11. Department of Public Health Sciences, and Cancer Research Institute, Queen's University, Kingston, ON, Canada.
12. Cancer Prevention Program, Fred Hutchinson Cancer Research Center, Seattle, WA, USA.
13. Zilber School of Public Health, University of Wisconsin-Milwaukee, Milwaukee, WI, USA.
14. Division of Cancer Epidemiology, German Cancer Research Center (DKFZ), Heidelberg, Germany.
15. Centre for Cancer Genetic Epidemiology, Department of Oncology, University of Cambridge, Cambridge, UK.
16. Department of Gynaecology and Obstetrics, University Hospital Erlangen, Friedrich-Alexander University Erlangen-Nuremberg, Comprehensive Cancer Center Erlangen-EMN, Erlangen, Germany.

- 93 17. Human Cancer Genetics Program, Spanish National Cancer Research Centre, Madrid,
94 Spain.
- 95 18. Centro de Investigación en Red de Enfermedades Raras (CIBERER), Valencia, Spain.
- 96 19. Institute of Biochemistry and Genetics, Ufa Scientific Center of Russian Academy of
97 Sciences, Ufa, Russia.
- 98 20. Gynaecology Research Unit, Hannover Medical School, Hannover, Germany.
- 99 21. Department of Oncology, Helsinki University Hospital, University of Helsinki, Helsinki,
100 Finland.
- 101 22. Department of Radiation Oncology, Hannover Medical School, Hannover, Germany.
- 102 23. N.N. Alexandrov Research Institute of Oncology and Medical Radiology, Minsk,
103 Belarus.
- 104 24. Copenhagen General Population Study, Herlev and Gentofte Hospital, Copenhagen
105 University Hospital, Herlev, Denmark.
- 106 25. Department of Clinical Biochemistry, Herlev and Gentofte Hospital, Copenhagen
107 University Hospital, Herlev, Denmark.
- 108 26. Faculty of Health and Medical Sciences, University of Copenhagen, Copenhagen,
109 Denmark.
- 110 27. Dr. Margarete Fischer-Bosch-Institute of Clinical Pharmacology, Stuttgart, Germany.
- 111 28. University of Tübingen, Tübingen, Germany.
- 112 29. German Cancer Consortium (DKTK), German Cancer Research Center (DKFZ),
113 Heidelberg, Germany.
- 114 30. Division of Preventive Oncology, German Cancer Research Center (DKFZ) and National
115 Center for Tumor Diseases (NCT), Heidelberg, Germany.
- 116 31. Division of Cancer Epidemiology and Genetics, National Cancer Institute, Rockville,
117 MD, USA.
- 118 32. Department of Cancer Epidemiology, Clinical Sciences, Lund University, Lund, Sweden.
- 119 33. Department of Gynecology and Obstetrics, University of Tübingen, Tübingen, Germany.
- 120 34. Department of Obstetrics and Gynecology, University of Heidelberg, Heidelberg,
121 Germany.
- 122 35. Molecular Epidemiology Group, C080, German Cancer Research Center (DKFZ),
123 Heidelberg, Germany.
- 124 36. Medical Oncology Department, CIBERONC Hospital Clínico San Carlos, Madrid, Spain.
- 125 37. Genomic Epidemiology Group, German Cancer Research Center (DKFZ), Heidelberg,
126 Germany.
- 127 38. Epidemiology Research Program, American Cancer Society, Atlanta, GA, USA.
- 128 39. Oncology and Genetics Unit, Instituto de Investigacion Biomedica (IBI) Orense-
129 Pontevedra-Vigo, Xerencia de Xestion Integrada de Vigo-SERGAS, Vigo, Spain.
- 130 40. University Cancer Center Hamburg (UCCH), University Medical Center Hamburg-
131 Eppendorf, Hamburg, Germany.
- 132 41. Department of Epidemiology, University of Florida, Gainesville, FL, USA.
- 133 42. Westmead Institute for Medical Research, University of Sydney, Sydney, Australia.
- 134 43. Department of Oncology, Haukeland University Hospital, Bergen, Norway.
- 135 44. National Advisory Unit on Late Effects after Cancer Treatment, Oslo University Hospital
136 Radiumhospitalet, Oslo, Norway.
- 137 45. Oslo University Hospital, Oslo, Norway.

- 138 46. Department of Clinical Genetics, Erasmus University Medical Center, Rotterdam, The
139 Netherlands.
- 140 47. Division of Molecular Pathology, The Netherlands Cancer Institute - Antoni van
141 Leeuwenhoek Hospital, Amsterdam, The Netherlands.
- 142 48. Department of Laboratory Medicine and Pathology, Mayo Clinic, Rochester, MN, USA.
- 143 49. Department of Epidemiology and Biostatistics, School of Public Health, Imperial College
144 London, London, UK.
- 145 50. INSERM U1052, Cancer Research Center of Lyon, Lyon, France.
- 146 51. Sheffield Institute for Nucleic Acids, Department of Oncology and Metabolism,
147 University of Sheffield, Sheffield, UK.
- 148 52. Academic Unit of Pathology, Department of Neuroscience, University of Sheffield,
149 Sheffield, UK.
- 150 53. Department of Medical Epidemiology and Biostatistics, Karolinska Institutet, Stockholm,
151 Sweden.
- 152 54. Department of Clinical Genetics, Fox Chase Cancer Center, Philadelphia, PA, USA.
- 153 55. Department of Pathology, Leiden University Medical Center, Leiden, The Netherlands.
- 154 56. Department of Human Genetics, Leiden University Medical Center, Leiden, The
155 Netherlands.
- 156 57. Center for Inherited Disease Research (CIDR), Institute of Genetic Medicine, Johns
157 Hopkins University School of Medicine, Baltimore, MD, USA.
- 158 58. Department of Non-Communicable Disease Epidemiology, London School of Hygiene
159 and Tropical Medicine, London, UK.
- 160 59. Genomics Center, Centre Hospitalier Universitaire de Québec Research Center, Laval
161 University, Québec City, QC, Canada.
- 162 60. Department of Biomedical Sciences, Faculty of Science and Technology, University of
163 Westminster, London, UK.
- 164 61. Cancer Sciences Academic Unit, Faculty of Medicine, University of Southampton,
165 Southampton, UK.
- 166 62. Channing Division of Network Medicine, Department of Medicine, Brigham and
167 Women's Hospital, Harvard Medical School, Boston, MA, USA.
- 168 63. Institute for Medical Informatics, Statistics and Epidemiology, University of Leipzig,
169 Leipzig, Germany.
- 170 64. David Geffen School of Medicine, Department of Medicine Division of Hematology and
171 Oncology, University of California at Los Angeles, Los Angeles, CA, USA.
- 172 65. Usher Institute of Population Health Sciences and Informatics, The University of
173 Edinburgh Medical School, Edinburgh, UK.
- 174 66. Institute for Medical Biometrics and Epidemiology, University Medical Center Hamburg-
175 Eppendorf, Hamburg, Germany.
- 176 67. Department of Cancer Epidemiology, Clinical Cancer Registry, University Medical
177 Center Hamburg-Eppendorf, Hamburg, Germany.
- 178 68. The Breast Cancer Now Toby Robins Research Centre, The Institute of Cancer Research,
179 London, UK.
- 180 69. Department of Breast Surgery, Herlev and Gentofte Hospital, Copenhagen University
181 Hospital, Herlev, Denmark.
- 182 70. School of Public Health, Curtin University, Perth, Australia.

- 183 71. Genomic Medicine Group, Galician Foundation of Genomic Medicine, Instituto de
184 Investigación Sanitaria de Santiago de Compostela (IDIS), Complejo Hospitalario
185 Universitario de Santiago, SERGAS, Santiago De Compostela, Spain.
- 186 72. Moores Cancer Center, University of California San Diego, La Jolla, CA, USA.
- 187 73. Cancer Epidemiology & Intelligence Division, Cancer Council Victoria, Melbourne,
188 Australia.
- 189 74. Centre for Epidemiology and Biostatistics, Melbourne School of Population and Global
190 Health, The University of Melbourne, Melbourne, Australia.
- 191 75. Department of Medicine, McGill University, Montréal, QC, Canada.
- 192 76. Division of Clinical Epidemiology, Royal Victoria Hospital, McGill University,
193 Montréal, QC, Canada.
- 194 77. Department of Dermatology, Huntsman Cancer Institute, University of Utah School of
195 Medicine, Salt Lake City, UT, USA.
- 196 78. Cancer & Environment Group, Center for Research in Epidemiology and Population
197 Health (CESP), INSERM, University Paris-Sud, University Paris-Saclay, Villejuif,
198 France.
- 199 79. Center for Hereditary Breast and Ovarian Cancer, University Hospital of Cologne,
200 Cologne, Germany.
- 201 80. Center for Integrated Oncology (CIO), University Hospital of Cologne, Cologne,
202 Germany.
- 203 81. Center for Molecular Medicine Cologne (CMMC), University of Cologne, Cologne,
204 Germany.
- 205 82. Department of Preventive Medicine, Keck School of Medicine, University of Southern
206 California, Los Angeles, CA, USA.
- 207 83. Institute of Environmental Medicine, Karolinska Institutet, Stockholm, Sweden.
- 208 84. Department of Health Sciences Research, Mayo Clinic, Rochester, MN, USA.
- 209 85. Molecular Genetics of Breast Cancer, German Cancer Research Center (DKFZ),
210 Heidelberg, Germany.
- 211 86. Cancer Genomics Research Laboratory, Leidos Biomedical Research, Frederick National
212 Laboratory for Cancer Research, Frederick, MD, USA.
- 213 87. Department of Medical Oncology, Family Cancer Clinic, Erasmus MC Cancer Institute,
214 Rotterdam, The Netherlands.
- 215 88. Department of Genetics and Pathology, Pomeranian Medical University, Szczecin,
216 Poland.
- 217 89. Department of Gynecology and Obstetrics, University Hospital Ulm, Ulm, Germany.
- 218 90. Department of Epidemiology, Cancer Prevention Institute of California, Fremont, CA,
219 USA.
- 220 91. Department of Health Research and Policy - Epidemiology, Stanford University School
221 of Medicine, Stanford, CA, USA.
- 222 92. Stanford Cancer Institute, Stanford University School of Medicine, Stanford, CA, USA.
- 223 93. Division of Genetics and Epidemiology, The Institute of Cancer Research, London, UK.
- 224 94. School of Medicine, National University of Ireland, Galway, Ireland.
- 225 95. Department of Genetics and Fundamental Medicine, Bashkir State University, Ufa,
226 Russia.
- 227 96. Translational Cancer Research Area, University of Eastern Finland, Kuopio, Finland.

- 228 97. Institute of Clinical Medicine, Pathology and Forensic Medicine, University of Eastern
229 Finland, Kuopio, Finland.
- 230 98. Imaging Center, Department of Clinical Pathology, Kuopio University Hospital, Kuopio,
231 Finland.
- 232 99. Department of Cancer Genetics, Institute for Cancer Research, Oslo University Hospital
233 Radiumhospitalet, Oslo, Norway.
- 234 100. Institute of Clinical Medicine, Faculty of Medicine, University of Oslo, Oslo, Norway.
- 235 101. Department of Clinical Molecular Biology, Oslo University Hospital, University of Oslo,
236 Oslo, Norway.
- 237 102. VIB KULeuven Center for Cancer Biology, VIB, Leuven, Belgium.
- 238 103. Laboratory for Translational Genetics, Department of Human Genetics, KU Leuven,
239 Leuven, Belgium.
- 240 104. Epidemiology Program, University of Hawaii Cancer Center, Honolulu, HI, USA.
- 241 105. Department of Epidemiology, University of Washington School of Public Health, Seattle,
242 WA, USA.
- 243 106. Department of Cancer Epidemiology and Prevention, M. Sklodowska-Curie Institute -
244 Oncology Center, Warsaw, Poland.
- 245 107. German Breast Group, GmbH, Neu Isenburg, Germany.
- 246 108. Southampton Clinical Trials Unit, Faculty of Medicine , University of Southampton,
247 Southampton, UK.
- 248 109. Research Centre for Genetic Engineering and Biotechnology "Georgi D. Efremov" ,
249 Macedonian Academy of Sciences and Arts, Skopje, Republic of Macedonia.
- 250 110. Department of Medicine, Brigham and Women's Hospital, Harvard Medical School,
251 Boston, MA, USA.
- 252 111. Department of Oncology - Pathology, Karolinska Institutet, Stockholm, Sweden.
- 253 112. Department of Medical Oncology, University Hospital of Heraklion, Heraklion, Greece.
- 254 113. Division of Gynaecology and Obstetrics, Technische Universität München, Munich,
255 Germany.
- 256 114. Gynaecological Cancer Research Centre, Women's Cancer, Institute for Women's Health,
257 University College London, London, UK.
- 258 115. Department of Laboratory Medicine and Pathobiology, University of Toronto, Toronto,
259 ON, Canada.
- 260 116. Laboratory Medicine Program, University Health Network, Toronto, ON, Canada.
- 261 117. Department of Population Sciences, Beckman Research Institute of City of Hope, Duarte,
262 CA, USA.
- 263 118. Department of Obstetrics and Gynecology, Helsinki University Hospital, University of
264 Helsinki, Helsinki, Finland.
- 265 119. Leuven Multidisciplinary Breast Center, Department of Oncology, Leuven Cancer
266 Institute, University Hospitals Leuven, Leuven, Belgium.
- 267 120. Center for Clinical Cancer Genetics and Global Health, The University of Chicago,
268 Chicago, IL, USA.
- 269 121. IFOM, The FIRC (Italian Foundation for Cancer Research) Institute of Molecular
270 Oncology, Milan, Italy.
- 271 122. Laboratory of Cancer Genetics and Tumor Biology, Cancer and Translational Medicine
272 Research Unit, Biocenter Oulu, University of Oulu, Oulu, Finland.

- 273 123. Laboratory of Cancer Genetics and Tumor Biology, Northern Finland Laboratory Centre
274 Oulu, Oulu, Finland.
- 275 124. Department of Gynecology and Obstetrics, Ludwig-Maximilians University of Munich,
276 Munich, Germany.
- 277 125. Unit of Molecular Bases of Genetic Risk and Genetic Testing, Department of Preventive
278 and Predictive Medicine, Fondazione IRCCS (Istituto Di Ricovero e Cura a Carattere
279 Scientifico) Istituto Nazionale dei Tumori (INT), Milan, Italy.
- 280 126. Section of Cancer Genetics, The Institute of Cancer Research, London, UK.
- 281 127. Clalit National Cancer Control Center, Haifa, Israel.
- 282 128. Medical Oncology Department, Hospital Universitario Puerta de Hierro, Madrid, Spain.
- 283 129. Hereditary Cancer Clinic, University Hospital of Heraklion, Heraklion, Greece.
- 284 130. Epidemiology Branch, National Institute of Environmental Health Sciences, NIH,
285 Research Triangle Park, NC, USA.
- 286 131. Research Oncology, Guy's Hospital, King's College London, London, UK.
- 287 132. Division of Psychosocial Research and Epidemiology, The Netherlands Cancer Institute -
288 Antoni van Leeuwenhoek hospital, Amsterdam, The Netherlands.
- 289 133. National Center for Tumor Diseases, University of Heidelberg, Heidelberg, Germany.
- 290 134. Division of Molecular Medicine, Pathology North, John Hunter Hospital, Newcastle,
291 Australia.
- 292 135. Discipline of Medical Genetics, School of Biomedical Sciences and Pharmacy, Faculty of
293 Health, University of Newcastle, Callaghan, Australia.
- 294 136. Department of Pathology, The University of Melbourne, Melbourne, Australia.
- 295 137. Cancer Control Research, BC Cancer Agency, Vancouver, BC, Canada.
- 296 138. School of Population and Public Health, University of British Columbia, Vancouver, BC,
297 Canada.
- 298 139. The Curtin UWA Centre for Genetic Origins of Health and Disease, Curtin University
299 and University of Western Australia, Perth, Australia.
- 300 140. Department of Obstetrics and Gynaecology, University of Melbourne and the Royal
301 Women's Hospital, Melbourne, Australia.
- 302 141. Division of Breast Cancer Research, The Institute of Cancer Research, London, UK.
- 303 142. Epigenetic and Stem Cell Biology Laboratory, National Institute of Environmental
304 Health Sciences, NIH, Research Triangle Park, NC, USA.
- 305 143. Department of Epidemiology, Mailman School of Public Health, Columbia University,
306 New York, NY, USA.
- 307 144. McGill University and Génome Québec Innovation Centre, Montréal, QC, Canada.
- 308 145. Department of Surgery, Leiden University Medical Center, Leiden, The Netherlands.
- 309 146. Institute of Human Genetics, Pontificia Universidad Javeriana, Bogota, Colombia.
- 310 147. Department of Gynecology and Obstetrics, Helios Clinics Berlin-Buch, Berlin, Germany.
- 311 148. Biostatistics and Computational Biology Branch, National Institute of Environmental
312 Health Sciences, NIH, Research Triangle Park, NC, USA.
- 313 149. Department of Medicine, Institute for Human Genetics, UCSF Helen Diller Family
314 Comprehensive Cancer Center, University of California San Francisco, San Francisco,
315 CA, USA.
- 316 150. Peter MacCallum Cancer Center, Melbourne, Australia.
- 317 151. Department of Molecular Physiology & Biophysics, Vanderbilt Genetics Institute,
318 Vanderbilt University, Nashville, TN, USA.

152. Department of Clinical Genetics, VU University Medical Center, Amsterdam, The Netherlands.
153. Department of Medical Oncology, Dana Farber Cancer Institute, Boston, MA.
154. Department of Medicine, Harvard Medical School, Boston, MA.
155. Division of Genetics, Brigham and Women's Hospital, Boston, MA.
156. Department of Nutrition, Harvard T.H. Chan School of Public Health, Boston, MA.
157. Human Genetics, Genome Institute of Singapore, Singapore, Singapore.
158. Nuffield Department of Population Health, University of Oxford, Big Data Institute, Old Road Campus, Oxford OX3 7LF, UK.
159. Department of Oncology University of Örebro, Örebro, Sweden.
160. Lang Wu and Wei Shi are joint co-first authors.

Abstract:

Breast cancer risk variants identified in genome-wide association studies explain only a small fraction of familial relative risk, and genes responsible for these associations remain largely unknown. To identify novel risk loci and likely causal genes, we performed a transcriptome-wide association study evaluating associations of genetically predicted gene expression with breast cancer risk in 122,977 cases and 105,974 controls of European ancestry. We used data from subjects included in the Genotype-Tissue Expression Project to establish genetic models to predict gene expression in breast tissue and evaluated model performance using data from subjects included in The Cancer Genome Atlas. Of the 8,597 genes evaluated, significant associations were identified for 48 at a Bonferroni-corrected threshold of $P < 5.82 \times 10^{-6}$, including 14 genes at loci not yet reported for breast cancer risk. We silenced 13 genes and showed an effect for 11 on cell proliferation and/or colony forming efficiency. Our study provides new insights into breast cancer genetics and biology.

Breast cancer is the most commonly diagnosed malignancy among women in many countries¹. Genetic factors play an important role in breast cancer etiology. Multiple high- and moderate-penetrance genes, including *BRCA1*, *BRCA2*, *PALB2*, *CHEK2* and *ATM*, have been identified as contributors to familial breast cancer^{2,3}. However, deleterious germline mutations in these genes are rare, thus accounting for only a small fraction of breast cancer cases in the general population^{4,5}. Since 2007, genome-wide association studies (GWAS) have identified approximately 180 genetic loci harboring common, low-penetrance variants for breast cancer⁶⁻¹³, but these more common variants explain less than 20% of familial relative risk⁷.

A large proportion of disease-associated risk variants identified by GWAS are located in non-protein coding or intergenic regions and are not in linkage disequilibrium (LD) with any nonsynonymous coding single nucleotide polymorphisms (SNPs)¹⁴. Many of these susceptibility variants are located in gene regulatory elements^{15,16}, and it has therefore been hypothesized that most of the GWAS-identified associations may be driven by the regulatory function of risk variants on the expression levels of nearby genes. For breast cancer, recent studies have shown that GWAS-identified associations at 1p34, 1p36, 2q35, 5p12, 5p15.33, 5q11.2, 5q14, 6q25, 7q22, 9q31.2, 10q21.3, 10q26.13, 11p15, 11q13.3, 15q26.1, 19p13 and 19q13.31 are likely due to the effect of risk variants at these loci on regulating the expression of either nearby or more distal genes: *CITED4*, *KLHDC7A*, *IGFBP5*, *FGF10/MRPS30*, *TERT*, *MAP3K1*, *ATP6AP1L*, *RMND1*, *RASA4/PRKRIP1*, *KLF4*, *NRBF2*, *FGFR2*, *PIDD1*, *CCND1*, *RCCD1*, *ABHD8*, and *ZNF404*^{7,9,10,13,17-22}. However, for the large majority of the GWAS-identified breast cancer risk loci, the genes responsible for the associations remain unknown.

Several recent studies have reported that regulatory variants may account for a large proportion of disease heritability not yet discovered through GWAS²³⁻²⁵. Many of these variants may have a small effect size, and thus are difficult to identify in individual SNP-based GWAS studies, even with a very large sample size. Applying gene-based approaches that aggregate the effects of multiple variants into a single testing unit may increase study power to identify novel disease-associated loci. Transcriptome-wide association studies (TWAS) systematically investigate across the transcriptome the association of genetically predicted gene expression with disease risk, providing an effective approach to identify novel susceptibility genes²⁶⁻²⁹. Instead of testing millions of SNPs in GWAS, TWAS evaluate the association of predicted expression for selected genes, thus greatly reducing the burden of multiple comparisons in statistical inference. Recently, Hoffman et al performed a TWAS including 15,440 cases and 31,159 controls and reported significant associations for five genes with breast cancer risk³⁰. However, the sample size of that study was relatively small and several reported associations were not statistically significant after Bonferroni correction. Herein, we report results from a larger TWAS of breast cancer that used the MetaXcan method²⁶ to analyze summary statistics data from 122,977 cases and 105,974 controls of European descent from the Breast Cancer Association Consortium (BCAC).

Results

Gene expression prediction models

The overall study design is shown in **Supplementary Figure 1**. We used transcriptome and high-density genotyping data from 67 women of European descent included in the Genotype-Tissue Expression (GTEx) project to build genetic models to predict RNA expression levels for

each of the genes expressed in normal breast tissues, by applying the elastic net method ($\alpha=0.5$) with ten-fold cross-validation. Genetically regulated expression was estimated for each gene using variants within a 2 MB window flanking the respective gene boundaries, inclusive. SNPs with a minor allele frequency of at least 0.05 and included in the HapMap Phase 2 subset were used for model building. Of the models built for 12,696 genes, 9,109 showed a prediction performance (R^2) of at least 0.01 ($\geq 10\%$ correlation between predicted and observed expression). For genes for which the expression could not be predicted well using this approach, we built models using only SNPs located in the promoter or enhancer regions, as predicted using three breast cell lines in the Roadmap Epigenomics Project/Encyclopedia of DNA Elements Project. This approach leverages information from functional genomics and reduces the number of variants for variable selection, and therefore potentially improving statistical power. This enabled us to build genetic models for additional 3,715 genes with $R^2 \geq 0.01$. **Supplementary Table 1** provides detailed information regarding the performance threshold and types of models built in this study. Overall, genes that were predicted with $R^2 \geq 0.01$ in GTEx data were also predicted well in The Cancer Genome Atlas (TCGA) tumor-adjacent normal tissue data (correlation coefficient of 0.55 for R^2 in two datasets; **Supplementary Figure 2**). Based on model performance in GTEx and TCGA, we prioritized 8,597 genes for analyses of the associations between predicted gene expression and breast cancer risk using the following criteria: 1) genes with a model prediction R^2 of at least 0.01 in the GTEx set (10% correlation) and a Spearman's correlation coefficient of ≥ 0.1 in the external validation experiment using TCGA data, 2) genes with a prediction R^2 of at least 0.09 (30% correlation) in the GTEx set regardless of their performance in the TCGA set, 3) genes with a prediction R^2 of at least 0.01 in

the GTEx set (10% correlation) that could not be evaluated in the TCGA set because of a lack of data.

Association analyses of predicted gene expression with breast cancer risk

Using the MetaXcan method²⁶, we performed association analyses to evaluate predicted gene expression and breast cancer risk using the meta-analysis summary statistics of individual genetic variants generated for 122,977 breast cancer cases and 105,974 controls of European ancestry included in BCAC. For the majority of the tested genes, most of the SNPs selected for prediction models were used for the association analyses (e.g., $\geq 95\%$ predicting SNPs used for 83.8% of the tested genes, and $\geq 80\%$ predicting SNPs used for 95.6% of the tested genes). Lambda 1,000 ($\lambda_{1,000}$), a standardized estimate of the genomic inflation scaling to a study of 1,000 cases and 1,000 controls, was 1.004 in our study (Quantile-quantile (QQ) plot presented in **Supplementary Figure 3 (A)**). Of the 8,597 genes evaluated in this study, we identified 179 genes whose predicted expression was associated with breast cancer risk at $P < 1.05 \times 10^{-3}$, a FDR-corrected significance level (**Figure 1, Supplementary Table 2**). Of these, 48 showed a significant association at the Bonferroni-corrected threshold of $P \leq 5.82 \times 10^{-6}$ (**Figure 1, Tables 1-3**), including 14 genes located at 11 loci that are 500 kb away from any of the risk variants identified in previous GWAS of breast cancer risk (**Table 1**). An association between lower predicted expression and increased breast cancer risk was detected for *LRRC3B* (3p24.1), *SPATA18* (4q12), *UBD* (6p22.1), *MIR31HG* (9p21.3), *RIC8A* (11p15.5), *B3GNT1* (11q13.2), *GALNT16* (14q24.1) and *MAN2C1* and *CTD-2323K18.1* (15q24.2). Conversely, an association between higher predicted expression and increased breast cancer risk was identified for *ZSWIM5* (1p34.1), *KLHDC10* (7q32.2), *RP11-867G23.10* (11q13.2), *RP11-218M22.1* (12p13.33) and

PLEKHD1 (14q24.1). The remaining 34 significantly associated genes are all located at breast cancer susceptibility loci identified in previous GWAS (**Tables 2-3**). Among them, 23 have not yet been previously implicated as genes responsible for association signals with breast cancer risk identified at these loci through expression quantitative trait loci (eQTL) and/or functional studies, and do not harbor GWAS or fine-mapping identified risk variants (**Table 2**), while the other eleven (*KLHDC7A*⁷, *ALS2CR12*³¹, *CASP8*^{31,32}, *ATG10*⁹, *SNX32*³³, *STXBP4*^{34,35}, *ZNF404*⁸, *ATP6AP1L*⁹, *RMND1*¹⁷, *L3MBTL3*⁶, and *RCCD1*¹⁰) had been reported as potential causal genes at breast cancer susceptibility loci or harbor GWAS or fine-mapping identified risk variants (**Table 3**). Except for *RP11-73O6.3* and *L3MBTL3*, there was no evidence of heterogeneity in the gene-expression association ($I^2 < 0.2$) across the iCOGS, OncoArray, and GWAS datasets included in our analyses (**Supplementary Table 3**). Overall, through our agnostic search, we identified 37 novel susceptibility genes for breast cancer, including 21 protein-coding genes, 15 long non-coding RNAs (lncRNAs) and a processed transcript, and confirmed eleven genes known to potentially play a role in breast cancer susceptibility.

To determine whether the associations between predicted gene expression and breast cancer risk were independent of the association signals identified in previous GWAS, we performed conditional analyses adjusting for the GWAS-identified risk SNPs closest to the TWAS-identified gene (**Supplementary Table 4**)³⁶. We found that the associations for 11 genes (*LRRC3B*, *SPATA18*, *KLHDC10*, *MIR31HG*, *RIC8A*, *B3GNT1*, *RP11-218M22.1*, *MAN2C1*, *CTD-2323K18.1* (**Table 1**), *ALK*, *CTD-3051D23.1* (**Table 2**)) remained statistically significant at $P < 5.82 \times 10^{-6}$ (**Tables 1-3**). This suggests the expression of these genes may be associated with breast cancer risk independent of the GWAS-identified risk variant(s). For nine of the genes

(*SPATA18*, *KLHDC10*, *MIR31HG*, *RIC8A*, *RP11-218M22.1*, *MAN2C1*, *CTD-2323K18.1* (**Table 1**), *ALK*, and *CTD-3051D23.1* (**Table 2**)), the significance level of the association remained essentially unchanged, suggesting these associations may be entirely independent of GWAS-identified association signals.

Of the 131 genes showing a significant association at P values between 5.82×10^{-6} and 1.05×10^{-3} (significant after FDR-correction but not Bonferroni-correction), 38 are located at GWAS-identified breast cancer risk loci (± 500 kb of the index SNPs) (**Table 4**). Except for *RP11-400F19.8*, there was no evidence of heterogeneity in TWAS association ($I^2 < 0.2$) across the iCOGS, OncoArray, and GWAS studies (**Supplementary Table 3**). After adjusting for the index SNPs, breast cancer associations for *MTHFD1L*, *PVT1*, *RP11-123K19.1*, *FES*, *RP11-400F19.8*, *CTD-2538G9.5*, and *CTD-3216D2.5* remained significant at $p \leq 1.05 \times 10^{-3}$, again suggesting that the association of these genes with breast cancer risk may be independent of the GWAS-identified association signals (**Table 4**).

For 41 of the 48 associated genes that reached the Bonferroni-corrected significant level, we obtained individual-level data from subjects included in the iCOGS ($n=84,740$) and OncoArray ($n=112,133$) datasets, which was 86% of the subjects included in the analysis using summary statistics (**Supplementary Table 5**). The results from the analysis using individual-level data were very similar to those described above using MetaXcan analyses (Pearson correlation of z -scores was 0.991 for iCOGS data and 0.994 for OncoArray data), although not all associations reached the Bonferroni-corrected significant level, possibly due to a smaller sample size (**Supplementary Table 5**). Conditional analyses using individual level data also revealed

consistent results compared with analyses using summary data. We found that for several genes within the same genomic region, their predicted expression levels were correlated with each other (**Tables 1-3**). The associations between predicted expression of *PLEKHD1* and *ZSWIM5* and breast cancer risk were largely influenced by their corresponding closest risk variants identified in GWAS, although these risk variants are >500 kb away from these genes (**Table 1**). There were significant correlation of rs999737 and rs1707302 with genetically predicted expression of *PLEKHD1* ($r = -0.47$ in the OncoArray dataset and -0.48 in the iCOGS dataset) and *ZSWIM5* ($r = 0.50$ in the OncoArray dataset and 0.51 in the iCOGS dataset), respectively.

INQUISIT algorithm scores for the identified genes

For the 48 associated genes after Bonferroni correction, we assessed their integrated expression quantitative trait and *in silico* prediction of GWAS target (INQUISIT) scores⁷ to assess whether there are other lines of evidence beyond the scope of eQTL for supporting our TWAS-identified genes as candidate target genes at GWAS-identified loci. The detailed methodology for INQUISIT scores have been described elsewhere⁷. In brief, a score for each gene-SNP pair is calculated across categories representing potential regulatory mechanisms - distal or proximal gene regulation (promoter). Features contributing to the score are based on functionally important genomic annotations such as chromatin interactions, transcription factor binding, and eQTLs. Compared with evidence from eQTL only, INQUISIT scores incorporate additional lines of evidence, including distal regulations. The INQUISIT scores for our identified genes are shown in **Supplementary Table 6**. Except for *UBD* with a very low score in the distal regulation category (0.05), none of the genes at novel loci (**Table 1**) showed evidence to be potential target genes for any of the GWAS-identified breast cancer susceptibility loci. This is interesting and

within the expectation since these genes may represent novel association signals. There was evidence suggesting that *RP11-439A17.7*, *NUDT17*, *ANKRD34A*, *BTN3A2*, *AP006621.6*, *RPLP2*, *LRRC37A2*, *LRRC37A*, *KANSL1-AS1*, *CRHR1* and *HAPLN4* listed in Table 2, and all eleven genes listed in Table 3, may be target genes for risk variants identified in GWAS at these loci (**Supplementary Table 6**). For *NUDT17*, *ANKRD34A*, *RPLP2*, *LRRC37A2*, *LRRC37A*, *KANSL1-AS1*, *CRHR1*, *HAPLN4*, *KLHDC7A*, *ALS2CR12*, *CASP8*, *ATG10*, *ATP6AP1L*, *L3MBTL3*, *RMND1*, *SNX32*, *RCCD1*, *STXBP4* and *ZNF404*, the INQUISIT scores were not derived only from eQTL data, providing orthogonal support for these loci. For these loci, the associations of candidate causal SNPs with breast cancer risk may be mediated through these genes. This is in general consistent with the findings from the conditional analyses described above.

Pathway enrichment analyses

Ingenuity Pathway Analysis (IPA)³⁷ suggested potential enrichment of cancer-related functions for the significantly associated protein-coding genes identified in this study (**Supplementary Table 7**). The top canonical pathways identified in these analyses included apoptosis related pathways (Granzyme B signaling ($p=0.024$) and cytotoxic T lymphocyte-mediated apoptosis of target cells ($p=0.046$)), immune system pathway (inflammasome pathway ($p=0.030$)), and tumoricidal function of hepatic natural killer cells ($p=0.036$). The identified pathways are largely consistent with findings in previous studies⁷. For the significantly associated lncRNAs identified in this study, pathway analysis of their highly co-expressed protein-coding genes also revealed potential over-representation of cancer related functions (**Supplementary Table 7**).

Knockdown of predicted risk-associated genes in breast cells

To assess the function of genes whose high levels of predicted expression were associated with increased breast cancer risk, we selected 13 genes for knockdown experiments in breast cells: *ZSWIM5*, *KLHDC10*, *RP11-218M22.1* and *PLEKHD1* (**Table 1**), *UBLCP1*, *AP006621.6*, *RP11-467J12.4*, *CTD-3032H12.1* and *RP11-15A1.7* (**Table 2**), and *ALS2CR12*, *RMND1*, *STXBP4* and *ZNF404* (**Table 3**). As negative controls, we selected *B2M*, *ARHGDIA* and *ZAP70* using the following criteria: 1) at least 2 MB from any known breast cancer risk locus; 2) not an essential gene in breast cancer^{38,39}; and 3) not predicted to be a target gene in INQUISIT. In addition, as positive controls, we included in the experiments *PIDD1* (**Table 4**)⁷, *NRBF2*²⁰ and *ABHD8*²², which have been functionally validated as the target genes at breast cancer risk loci. We performed quantitative PCR (qPCR) on a panel of three ‘normal’ mammary epithelial and 15 breast cancer cell lines to analyze their expression level (**Supplementary Figure 4 and Supplementary Table 8**). All 19 genes were expressed in the normal mammary epithelial line 184A1⁴⁰ and the luminal breast cancer cell lines, MCF7 and T47D, so we used these cell lines for the proliferation assay, and MCF7 for the colony formation assay⁴¹. We also evaluated *SNX32*, *ALK* and *BTN3A2* by qPCR, but they were not expressed in T47D and MCF7 cells; therefore they were not evaluated further. It was difficult to design siRNAs against *RP11-867G23.1* and *RP11-53O19.1* because they both have multiple transcripts with limited, GC-rich regions in common. We did not include *RPLP2* because it is already known to be an essential gene for breast cancer survival⁴². Knockdown of the 19 tested genes was achieved by small short interfering RNA (siRNA) (**Supplementary Table 9**) and the knockdown efficiency was calculated in 184A1, MCF7 and T47D for each siRNA pair. Robust knockdown of the gene of

interests (GOI) was validated by qPCR with the majority of the siRNAs (**Supplementary Figure 5**).

To evaluate the survival and proliferation ability of cells following gene interruption, we used an IncuCyte to quantify cell proliferation in real time and quantified the corrected proliferation of cells with knocking down of GOI in comparison to that of cells with non-target control (NTC) siRNA). As expected, knockdown of the three negative control genes (*B2M*, *ARHGDIA* and *ZAP70*) did not significantly change cell proliferation in any of the three cell lines (**Figure 2A, Supplementary Figure 6**). However, with the exception of *UBLCP1*, *RMND1* and *STXBP4*, knockdown of all other genes (11 TWAS-identified genes along with two known genes, *ABHD8* and *NRBF2*) resulted in significantly decreased cell proliferation in 184A1 normal breast cells, with *KLHDC10*, *PLEKHD1*, *RP11-218M22.1*, *AP006621.6*, *ZNF404*, *RP11-467J12.4*, *CTD-3032H12.1* and *STXBP4* showing a similar effect in one or both cancer cell lines. Down-regulation of three lncRNAs (*RP11-218M22.1*, *RP11-467J12.4* and *CTD-3032H12.1*) resulted in significant reduction in cell proliferation in all three cell lines. We also evaluated the effect of inhibition of these genes on colony forming ability in MCF7 cells. Knockdown of the three negative control genes did not significantly affect colony forming efficiency (CFE). By contrast, knockdown of *PIDD1*, *RP11-15A1.7*, *RP11-218M22.1*, *AP006621.6*, *ZNF404*, *RP11-467J12.4* and *CTD-3032H12.1* resulted in significantly decreased colony forming efficiency in MCF7 cells compared to the NTC (**Figure 2B, Supplementary Figure 7**).

Discussion

This is the largest study to systematically evaluate associations of genetically predicted gene

expression across the human transcriptome with breast cancer risk. We identified 179 genes showing a significant association at the FDR-corrected significance level. Of these, 48 showed a significant association at the Bonferroni-corrected threshold, including 14 genes at genomic loci that have not previously been implicated for breast cancer risk. Of the 34 genes we identified that are located at known risk loci, 23 have not previously been shown to be the targets of GWAS-identified risk SNPs at corresponding loci and not harbor any risk SNPs. Our study provides substantial new information to improve the understanding of genetics and etiology for breast cancer, the most common malignancy among women in most countries.

It is possible that TWAS-identified genes may be associated with breast cancer risk through their correlation with disease causal genes. To determine the potential functional significance of TWAS-identified genes and provide evidence for causal inference, we knocked down 13 genes for which high predicted levels of expression were associated with an increased breast cancer risk, in one normal and two breast cancer cell lines, and measured the effect on proliferation and colony forming efficiency. Although there was some variation between cell lines, knockdown of 11 of the 13 genes showed an effect in at least one cell line, particularly on proliferation in 184A1 normal breast cells; the effects were strongest and most consistent for the lncRNAs, *RP11-218M22.1*, *RP11-467J12.4* and *CTD-3032H12.1*. The observation of a more consistent effect in the normal breast cell line compared with the cancer cell lines is not surprising as cancer cell lines have increased capacity to handle gene interference through mutations which enhance cell survival. Rewiring of pathways and compensatory mechanisms is a hallmark of cancer. Knockdown of *PIDD1*, *NRBF2* and *ABHD8*, for which breast cancer risk associated haplotypes have been shown to be associated with increased expression in reporter assays^{7,20,22}, affected

either proliferation or colony forming efficiency, supporting the results from this study. Knockdown of *UBLCP1* and *RMND1* did not affect proliferation or colony formation but they could mediate breast cancer risk through other mechanisms.

Some of the genes with strong functional evidence from our study have been reported to have important roles in carcinogenesis. For example, *RP11-467J12.4* (PR-lncRNA-1) is a p53-regulated lncRNA that modulates gene expression in response to DNA damage downstream of p53⁴³. *STXBP4* encodes Syntaxin binding protein 4, a scaffold protein that can stabilise and prevent degradation of an isoform of p63, a member of the p53 tumor suppressor family⁴⁴. *KLHDC10* encodes a member of the Kelch superfamily that can activate apoptosis signal-regulating kinase 1, contributing to oxidative stress-induced cell death⁴⁵. Notably, another member of this superfamily, *KLHDC7A*, has recently been identified as the target gene at the 1p36 breast cancer risk locus⁷.

SNX32, *ALK* and *BTN3A2* are also likely susceptibility genes for breast cancer risk. However, their low or absent expression in our chosen breast cell lines prevented further functional analysis. *SNX32* (Sorting Nexin 32) is not well characterized, but *ALK* (Anaplastic lymphoma kinase) copy number gain and overexpression have been reported in aggressive and metastatic breast cancers⁴⁶. Therapeutic targeting of ALK rearrangement has significantly improved survival in advanced ALK-positive lung cancer⁴⁷, making it an attractive target for breast and other cancers. *BTN3A2* is a member of the B7/butyrophilin-like group of Ig superfamily receptors modulating the function of T-lymphocytes. While the exact role of *BTN3A2* remains

unknown, over-expression of this gene in epithelial ovarian cancer is associated with higher infiltrating immune cells and a better prognosis⁴⁸.

Our analyses identified multiple genes with reduced expression levels associated with increased breast cancer risk. Among them, *LRRC3B* and *CASP8* are putative tumor suppressors in multiple cancers, including breast cancer. Leucine-rich repeat-containing 3B (*LRRC3B*) is a putative LRR-containing transmembrane protein, which is frequently inactivated via promoter hypermethylation leading to inhibition of cancer cell growth, proliferation, and invasion⁴⁹. *CASP8* encodes a member of the cysteine-aspartic acid protease family, which play a central role in cell apoptosis. Previous studies have suggested that caspase-8 may act as a tumor suppressor in certain types of lung cancer and neuroblastoma, although this function has not yet been demonstrated in breast cancer. Notably, several large association studies have identified SNPs at the 2q33/*CASP8* locus associated with increased breast cancer risk^{31,50}. Consistent with our data, eQTL analyses showed that the risk alleles for breast cancer were associated with reduced *CASP8* mRNA levels in both peripheral blood lymphocytes and normal breast tissue³¹.

For seven of the genes listed in Tables 1 and 2, we found some evidence from studies using tumor tissues, *in vitro* or *in vivo* experiments linking them to cancer risk (**Supplementary Table 10**), although their association with breast cancer has not been previously demonstrated in human studies. For five of them, including *LRRC3B*, *SPATA18*, *RIC8A*, *ALK* and *CRHR1*, previous *in vitro* and *in vivo* experiments and human tissue studies showed a consistent direction of the association as demonstrated in our studies. For two other genes (*UBD* and *MIR31HG*), however, results from previous studies were inconsistent, reporting both potential promoting and inhibiting

effects on breast cancer development. Future studies are needed to evaluate functions of these genes.

We included a large number of cases and controls in this study, providing strong statistical power for the association analysis. This large sample size enabled us to identify a large number of candidate breast cancer susceptibility genes, much larger than the number identified in a TWAS study with a sample size of about 20% of ours³⁰. The previous study included subjects of different races, which could affect the results as linkage disequilibrium (LD) patterns differ by races. Of the five genes reported in that smaller TWAS that showed a suggestive association with breast cancer risk, the association for the *RCCD1* gene was replicated in our study (**Table 3**).

The other four genes (*ANKLE1*, *DHODH*, *ACAP1* and *LRRC25*) were not evaluated in our study because of unsatisfactory performance of our breast specific models for these genes which were built using the GTEx reference dataset including only female European descendants. In our study, the expression prediction model for *ANKLE1* has a marginal performance in predicting gene expression ($R^2=0.013$ in the GTEx). The model, however, did not perform well in the TCGA data. For *ACAP1* and *LRRC25*, previous results for suggestive associations were based on blood tissue models.

A substantial proportion of SNPs included in the OncoArray and iCOGS were selected from breast cancer GWAS and fine-mapping analyses, and thus these arrays were enriched for association signals with breast cancer risk. As a result, the overall λ value for the BCAC association analyses of individual variants is 1.26 after adjusting for population stratifications (QQ plot in **Supplementary Figure 3 (B)**)⁷. The λ value for the associations of the ~257,000

SNPs included in the gene expression prediction models of the 8,597 genes tested in our association analysis is 1.40 (QQ plot in **Supplementary Figure 3 (C)**). This higher λ value is perhaps expected because of a potential further enrichment of breast cancer associated signals in the set of SNPs selected to predict gene expression. There could be additional gain of power (and thus a higher λ value) in TWAS as it aggregates the effect of multiple SNPs to predict gene expression and use genes as the unit for association analyses. The lambda (λ) for our associated analyses of 8,597 genes was 1.51 (QQ plot presented in **Supplementary Figure 3 (A)**) likely due to the potential enrichment and power gain discussed above as well as our large sample size, and the highly polygenic nature of the disease^{7,51}. Interestingly, high λ values were also found in recent large studies of other polygenic traits, such as body mass index (BMI) ($\lambda = 1.99$) and height ($\lambda = 2.7$)^{52,53}. The $\lambda_{1,000}$, a standardized estimate of the genomic inflation scaling to a study of 1,000 cases and 1,000 controls, is 1.004 in our study.

The statistical power of our study is very large to detect associations for genes with a relatively high cis-heritability (h^2) (**Supplementary Figure 8**). For example, our study has 80% statistical power to detect an association with breast cancer risk at $P < 5.82 \times 10^{-6}$ with an OR of 1.07 or higher per one standard deviation increase (or decrease) in the expression level of genes with an h^2 of 0.1 or higher. One limitation of our study is the small sample size for building gene expression prediction models, which may have affected the precision of model parameter estimates. The prediction performance (R^2) for several of the genes identified in our study was not optimal, and thus additional research is needed to confirm our findings. We expect that models built with a larger sample size (and thus with more stable estimates of model parameters) will identify additional association signals. We used samples from women of European origin in

model building, given differences in gene expression patterns between males and females and in genetic architecture across ethnicities⁵⁴. We also used gene expression data of tumor-adjacent normal tissue samples from European descendants in TCGA as an external validation step to prioritize genes for association analyses. Given potential somatic alterations in tumor-adjacent normal tissues, we retained all models showing a prediction performance (R^2) of at least 0.09 in GTEx, regardless of their performance in TCGA. Not all genes have a significant hereditary component in expression regulation, and thus these genes could not be investigated in our study. For example, previous studies have provided strong evidence to support a significant role of the *TERT*, *ESR1*, *CCND1*, *IGFBP5*, *TET2* and *MRPS30* genes in the etiology of breast cancer. However, expression of these genes cannot be predicted well using the data from female European descendants included in the GTEx and thus they were not included in our association analyses. **Supplementary Table 11** summarizes the performance of prediction models and association results for breast cancer target genes reported previously at GWAS-identified loci.

In summary, our study has identified multiple gene candidates that can be further functionally characterized. By evaluating the associations of predicted gene expression levels with breast cancer risk, we provided evidence for the direction of the association for the identified genes. The silencing experiments we performed suggest that many of the genes identified by TWAS are likely to mediate risk of breast cancer by affecting proliferation or colony forming efficiency, two of the hallmarks of cancer. Further investigation of genes identified in our study will provide additional insight into the biology and genetics of breast cancer.

Methods

Building of gene expression prediction models

We used transcriptome and high-density genotyping data from the Genotype-Tissue Expression (GTEx) study to establish prediction models for genes expressed in normal breast tissues. Details of the GTEx have been described elsewhere⁵⁵. Genomic DNA samples obtained from study subjects included in the GTEx were genotyped using Illumina OMNI 5M or 2.5M SNP Array and RNA samples from 51 tissue sites were sequenced to generate transcriptome profiling data. Genotype data were processed according to the GTEx protocol (<http://www.gtexportal.org/home/documentationPage>). SNPs with a call rate < 98%, with differential missingness between the two array experiments (5M/2.5M Arrays), with Hardy-Weinberg equilibrium p -value < 10^{-6} (among subjects of European ancestry), or showing batch effects were excluded. One Klinefelter individual, three related individuals, and a chromosome 17 trisomy individual were also excluded. The genotype data were imputed to the Haplotype Reference Consortium reference panel⁵⁶ using Minimac3 for imputation and SHAPEIT for prephasing^{57,58}. SNPs with high imputation quality ($r^2 \geq 0.8$), minor allele frequency (MAF) ≥ 0.05 , and included in the HapMap Phase 2 version, were used to build expression prediction models. For gene expression data, we used Reads Per Kilobase per Million (RPKM) units from RNA-SeQC⁵⁹. Genes with a median expression level of 0 RPKM across samples were removed, and the RPKM values of each gene were log2 transformed. We performed quantile normalization to bring the expression profile of each sample to the same scale, and performed inverse quantile normalization for each gene to map each set of expression values to a standard normal. We adjusted for the top ten principal components (PCs) derived from genotype data and the top 15 probabilistic estimation of expression residuals (PEER) factors to correct for batch effects and experimental confounders in model building⁶⁰. Genetic and transcriptome data from 67 female

subjects of European descent without a prior breast cancer diagnosis were used to build gene expression prediction models for this study.

We built an expression prediction model for each gene by using the elastic net method as implemented in the glmnet R package, with $\alpha=0.5$, as recommended by Gamazon et al²⁷. The genetically regulated expression for each gene was estimated by including variants within a 2 MB window flanking the respective gene boundaries, inclusive. Expression prediction models were built for protein coding genes, long non-coding RNAs (lncRNAs), microRNAs (miRNAs), processed transcripts, immunoglobulin genes, and T cell receptor genes, according to categories described in the Gencode V19 annotation file (<http://www.gencodegenes.org/releases/19.html>). Pseudogenes were not included in the present study because of potential concerns of inaccurate calling⁶¹. Ten-fold cross-validation was used to validate the models internally. Prediction R^2 values (the square of the correlation between predicted and observed expression) were generated to estimate the prediction performance of each of the gene prediction models established.

For genes that cannot be predicted well using the above approach, we built models using only SNPs located in predicted promoter or enhancer regions in breast cell lines. This approach reduces the number of variants for model building, and thus potentially improves model accuracy, by increasing the ratio of sample size to effective degrees of freedom. SNP-level annotation data in three breast cell lines, namely, Breast Myoepithelial Primary Cells (E027), Breast variant Human Mammary Epithelial Cells (vHMEC) (E028), and HMEC Mammary Epithelial Primary Cells (E119) in the Roadmap Epigenomics Project/Encyclopedia of DNA Elements Project¹⁶, were downloaded from

<http://archive.broadinstitute.org/mammals/haploreg/data/> (Version 4.0, assessed on December 6, 2016). SNPs in regions classified as promoters (TssA, TssAFlnk), enhancers (Enh, EnhG), or regions with both promoter and enhancer signatures (ExFlnk) according to the core 15 chromatin state model¹⁶ in at least one of the cell lines were retained as input SNPs for model building.

Evaluating performance of gene expression prediction models using The Cancer Genome Atlas (TCGA) data

To assess further the validity of the models, we performed external validation using data generated in tumor-adjacent normal breast tissue samples obtained from 86 European-ancestry female breast cancer patients included in the TCGA. Genotype data were imputed using the same approach as described for GTEx data. Expression data were processed and normalized using a similar approach as described above. The predicted expression level for each gene was calculated using the model established using GTEx data and then compared with the observed level of that gene using the Spearman's correlation.

Evaluating statistical power for association tests

We conducted a simulation analysis to assess the power of our TWAS analysis. Specifically, we set the number of cases and controls to be 122,977 and 105,974, respectively, and generated the gene expression levels from the empirical distribution of predicted gene expression levels in the BCAC. We calculated statistical power at $P < 5.82 \times 10^{-6}$ (the significance level used in our TWAS) according to cis-heritability (h^2) which we aim to capture using gene expression prediction models (R^2). The results based on 1000 replicates are summarized in **Supplementary Figure 8**. Based on the power calculation, our TWAS analysis has 80% power to detect a

minimum odds ratio of 1.11, 1.07, 1.05, 1.04, or 1.03 for breast cancer risk per one standard deviation increase (or decrease) in the expression level of a gene whose cis-heritability is 5%, 10%, 20%, 40%, or 60%, respectively.

Association analyses of predicted gene expression with breast cancer risk

We used the following criteria to select genes for the association analysis: 1) with a model prediction R^2 of ≥ 0.01 in GTEx and a Spearman's correlation coefficient of ≥ 0.1 in TCGA, 2) with a prediction R^2 of ≥ 0.09 in GTEx regardless of the performance in TCGA, 3) with a prediction R^2 of ≥ 0.01 in GTEx but unable to be evaluated in TCGA. The second group of genes was selected because some gene expression levels might have changed in TCGA tumor-adjacent normal tissues, and thus it is anticipated that some genes may show low prediction performance in TCGA data due to the influence of tumor growth^{62,63}. Overall, a total of 8,597 genes met the criteria and were evaluated for their expression-trait associations.

To identify novel breast cancer susceptibility loci and genes, the MetaXcan method, as described elsewhere, was used for the association analyses²⁶. Briefly, the formula:

$$Z_g \approx \sum_{l \in \text{Model}_g} w_{lg} \frac{\hat{\sigma}_l}{\hat{\sigma}_g} \frac{\hat{\beta}_l}{\text{se}(\hat{\beta}_l)}$$

was used to estimate the Z-score of the association between predicted expression and breast cancer risk. Here w_{lg} is the weight of SNP l for predicting the expression of gene g , $\hat{\beta}_l$ and $\text{se}(\hat{\beta}_l)$ are the GWAS association regression coefficient and its standard error for SNP l , and $\hat{\sigma}_l$ and $\hat{\sigma}_g$ are the estimated variances of SNP l and the predicted expression of gene g respectively. Therefore, the weights for predicting gene expression, GWAS summary statistics results, and

correlations between model predicting SNPs are the input variables for the MetaXcan analyses. For this study we estimated correlations between SNPs included in the prediction models using the phase 3, 1000 Genomes Project data focusing on European population.

For the association analysis, we used the summary statistics data of genetic variants associated with breast cancer risk generated in 122,977 breast cancer patients and 105,974 controls of European ancestry from the Breast Cancer Association Consortium (BCAC). The details of the BCAC have been described elsewhere^{7,9,13,64,65}. Briefly, 46,785 breast cancer cases and 42,892 controls of European ancestry were genotyped using a custom Illumina iSelect genotyping array (iCOGS) containing ~211,155 variants. A further 61,282 cases and 45,494 controls of European ancestry were genotyped using the OncoArray including 570,000 SNPs (<http://epi.grants.cancer.gov/oncoarray/>). Also included in this analysis were data from nine GWAS studies including 14,910 breast cancer cases and 17,588 controls of European ancestry. Genotype data from iCOGS, OncoArray and GWAS were imputed using the October 2014 release of the 1000 Genomes Project data as reference. Genetic association results for breast cancer risk were combined using inverse variance fixed effect meta-analyses⁷. For our study, only SNPs with imputation $r^2 \geq 0.3$ were used. All participating BCAC studies were approved by their appropriate ethics review boards. This study was approved by the BCAC Data Access Coordination Committee.

Lambda 1,000 ($\lambda_{1,000}$) was calculated to represent a standardized estimate of the genomic inflation scaling to a study of 1,000 cases and 1,000 controls, using the following formula:

$\lambda_{1,000} = 1 + (\lambda_{\text{obs}} - 1) \times (1/n_{\text{cases}} + 1/n_{\text{controls}}) / (1/1,000_{\text{cases}} + 1/1,000_{\text{controls}})$ ^{66,67}. We used a Bonferroni

corrected p threshold of 5.82×10^{-6} ($0.05/8,597$) to determine a statistically significant association for the primary analyses. To identify additional gene candidates at previously identified susceptibility loci, we also used a false discovery rate (FDR) corrected p threshold of 1.05×10^{-3} ($\text{FDR} \leq 0.05$) to determine a significant association. Associated genes with an expression of >0.1 RPKM in less than 10 individuals in GTEx data were excluded as the corresponding prediction models may not be stable.

To determine whether the predicted expression-trait associations were independent of the top signals identified in previous GWAS, we performed GCTA-COJO analyses developed by Yang et al³⁶ to calculate association betas and standard errors of variants with breast cancer risk after adjusting for the index SNPs of interest. We then re-ran the MetaXcan analyses using the association statistics after conditioning on the index SNPs. This information was used to determine whether the detected expression-trait associations remained significant after adjusting for the index SNPs.

For 41 identified associated genes at the Bonferroni-corrected threshold, we also performed analyses using individual level data in iCOGS ($n=84,740$) and OncoArray ($n=112,133$) datasets. We generated predicted gene expression using predicting SNPs, and then assessed the association between predicted gene expression and breast cancer risk adjusting for study and nine principal components in iCOGS dataset, and country and the first ten principal components in OncoArray dataset. Conditional analyses adjusting for index SNPs were performed to assess potential influence of reported index SNPs on the association between predicted gene expression and breast cancer risk. Furthermore, we evaluated whether the predicted expression levels of

genes within a same genomic region were correlated with each other by using the OncoArray data.

INQUISIT algorithm scores for TWAS-identified genes

To evaluate whether there are additional lines of evidence supporting the identified genes as putative target genes of GWAS identified risk SNPs beyond the scope of eQTL, we assessed their INQUISIT algorithm scores, which have been described elsewhere⁷. Briefly, this approach evaluates chromatin interactions between distal and proximal regulatory transcription-factor binding sites and the promoters at the risk regions using Hi-C data generated in HMECs⁶⁸ and Chromatin Interaction Analysis by Paired End Tag (ChiA-PET) in MCF7 cells. This could detect genome-wide interactions brought about by, or associated with, CCCTC-binding factor (CTCF), DNA polymerase II (POL2), and Estrogen Receptor (ER), all involved in transcriptional regulation⁶⁸. Annotation of predicted target genes used the Integrated Method for Predicting Enhancer Targets (IM-PET)⁶⁹, the Predicting Specific Tissue Interactions of Genes and Enhancers (PreSTIGE) algorithm⁷⁰, Hnisz⁷¹ and FANTOM⁷². Features contributing to the scores are based on functionally important genomic annotations such as chromatin interactions, transcription factor binding, and eQTLs. The detailed information for the INQUISIT pipeline and scoring strategy has been included in a previous publication⁷. In brief, besides assigning integral points according to different features, we also set up-weighting and down-weighting criteria according to breast cancer driver genes, topologically associated domain (TAD) boundaries, and gene expression levels in relevant breast cell lines. Scores in the distal regulation category range from 0-7, and in the promoter category from 0-4. A score of "none" represents that no evidence was found for regulation of the corresponding gene.

Functional enrichment analysis using Ingenuity Pathway Analysis (IPA)

We performed functional enrichment analysis for the identified protein-coding genes reaching Bonferroni corrected association threshold. To assess potential functionality of the identified lncRNAs, we examined their co-expressed protein-coding genes determined using expression data of normal breast tissue of European females in GTEx. Spearman's correlations between protein-coding genes and identified lncRNAs of ≥ 0.4 or ≤ -0.4 were used to indicate a high co-expression. Canonical pathways, top associated diseases and biofunctions, and top networks associated with genes of interest were estimated using IPA software³⁷.

Gene expression in breast cell lines

Total RNA was isolated from 18 cell lines (**Supplementary Table 8**) using the RNeasy Mini Kit (Qiagen). cDNA was synthesized using the SuperScript III (Invitrogen) and amplified using the Platinum SYBR Green qPCR SuperMix-UDG cocktail (Invitrogen). Two or three primer pairs were used for each gene and the mRNA levels for each sample was measured in technical triplicates for each primer set. The primer sequences are listed in **Supplementary Table 12**. Experiments were performed using an ABI ViiA(TM) 7 System (Applied Biosystems), and data processing was performed using ABI QuantStudio™ Software V1.1 (Applied Biosystems). The average of Ct from all the primer pairs for each gene was used to calculate ΔC_t . The relative quantitation of each mRNA normalizing to that in 184A1 was performed using the comparative Ct method ($\Delta\Delta C_t$) and summarized in **Supplementary Figure 4**.

Short interfering RNA (siRNA) silencing

MCF7 and T47D cells were reverse-transfected with siRNAs targeting genes of interest (GOI) or a non-targeting control siRNA (consi; Shanghai Genepharma) with RNAiMAX (Invitrogen) according to the manufacturer's protocol. Verification of siRNA knockdown of gene expression by qPCR was performed 36 hours after transfection.

Proliferation and colony formation assays

For proliferation assays, MCF7 and T47D cells were trypsinized at 16 hours post-transfection and seeded into 24 well plates to achieve ~10% confluency. Phase-contrast images were collected with IncuCyte ZOOM (Essen Bioscience) for seven days. Duplicate samples were assessed for each GOI siRNA transfected cells along with non-target control si (NTCsi) treated cells in the same plate. 184A1 cells were reverse-transfected in 96 well plates to achieve 50% confluence at 8 hours after transfection. Two independent experiments were carried out for all siRNAs in all three cell lines. Each cell proliferation time-course was normalized to the baseline confluency and analyzed in GraphPad Prism. The area under the curve was calculated for each concentration (n=4) and used to calculate corrected proliferation (Corrected proliferation % = $100 \pm (\text{relative proliferation in indicated siRNA} - \text{proliferation in NTC siRNA}) / \text{knockdown efficiency}$ ("+" if the GOI promotes proliferation and "-" if it inhibits proliferation)). For each gene, results from two siRNAs in two independent experiments were averaged and summarized in **Figure 2** and **Supplementary Figure 6**. For colony formation assays; the same number of GOI siRNA transfected MCF7 cells was seeded in 6 well plates at 16 hours after transfection to assay colony forming efficiency at two weeks. All siRNA-treated cells were seeded in duplicate. Colonies (defined to consist of at least 50 cells) were fixed with methanol, stained with crystal violet (0.5% w/v), scanned and counted using ImageJ as batch analysis by a self-defined plug-in

Macro. Correct CFE % = $100 \pm (\text{relative CFE in indicated siRNA} - \text{CFE in NTC siRNA}) /$
knockdown efficiency (“+” if the GOI promotes CF and “-” if it inhibits CF). For each gene,
results from two siRNAs in two independent experiments were averaged and summarized in
Figure 2 and Supplementary Figure 7.

Data availability

The GTEx data are publicly available via dbGaP (www.ncbi.nlm.nih.gov/gap; dbGaP Study
Accession: phs000424.v6.p1). TCGA data are publicly available via National Cancer Institute's
Genomic Data Commons Data Portal (<https://gdc.cancer.gov/>). Most of the BCAC data used in
this study are or will be publicly available via dbGAP. Data from some BCAC studies are not
publicly available due to restraints imposed by the ethics committees of individual studies;
requests for further data can be made to the BCAC (<http://bcac.ccge.medschl.cam.ac.uk/>) Data
Access Coordination Committee.

Code availability

The computer codes used in our study are available upon reasonable request.

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Author Contributions

W.Z. and J.L. conceived the study. L.W. contributed to the study design, and performed statistical analyses. L.W., W.Z. and G.C.-T. wrote the manuscript with significant contributions from W.S., J.L., X.G., and S.L.E.. W.S. performed the *in vitro* experiments. G.C.-T. directed the *in vitro* experiments. X.G. contributed to the model building and pathway analyses. J.B. contributed to the bioinformatics analyses. F.A.-E., E.R., and S.L.E. contributed to the *in vitro* experiments. Y. L. and C. Z. contributed to the model building. K.M., M.K.B., X.-O.S., Q.W., J.D., B.L., C.Z., H.F., A.G., R.T.B., A.M.D., P.D.P.P., J.S., R.L.M., P.K., and D.F.E, contributed to manuscript revision, statistical analyses and/or BCAC data management. I.L.A., H.A.-C., V.A., K.J.A., P.L.A., M. Barrdahl, C.B., M.W.B., J.B., M. Bermisheva, C.B., N.V.B., S.E.B., H. Brauch, H. Brenner, L.B., P.B., S.Y.B., B.B., Q.C., T.C., F.C., B.D.C., J.E.C., J.C.-C., X.C., T.-Y.D.C., H.C., C.L.C., NBCS Collaborators, M.C., S.C., F.J.C., D.C., A.C., S.S.C., J.M.C., K.C., M.B.D., P.D., K.F.D., T.D., I.d.S.S., M. Dumont, M. Dwek, D.M.E., U.E., H.E., C.E., M.E., L.F., P.A.F., J.F., D.F.-J., O.F., H.F., L.F., M. Gabrielson, M.G.-D., S.M.G., M.G.-C., M.M.G., M. Ghousaini, G.G.G., M.S.G., D.E.G., A.G.-N., P.G., E. Hahnen, C.A.H., N.H., P. Hall, E. Hallberg, U.H., P. Harrington, A. Hein, B.H., P. Hillemanns, A. Hollestelle, R.N.H., J.L.H., G.H., K.H., D.J.H., A.J., W.J., E.M.J., N.J., K.J., M.E.J., A. Jung, R.K., M.J.K., E.K., V.-M.K., V.N.K., D.L., L.L.M., J. Li, S.L., J. Lissowska, W.-Y.L., S.Loibl, J.L., C.L., M.P.L., R.J.M., T.M., I.M.K., A. Mannermaa, J.E.M., S.M., D.M., H.M.-H., A. Meindl, U.M., J.M., A.M.M., S.L.N., H.N., P.N., S.F.N., B.G.N., O.I.O., J.E.O., H.O., P.P., J.P., D.P.-K., R.P., N.P., K.P.,

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1009 **Competing financial interests**

1010 The authors declare no competing financial interests.

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Figure Legends

Figure 1. Manhattan plot of association results from the breast cancer transcriptome-wide

association study. The red line represents $P = 5.82 \times 10^{-6}$. The blue line represents $P = 1.00 \times 10^{-3}$.

Figure 2. Heat maps of proliferation and colony formation efficiency in breast cells. (A)

184A1, MCF7 or T47D cells were transfected with indicated siRNAs over seven days and phase-contrast images collected using an IncuCyte ZOOM. Each cell proliferation time-course was normalized to the baseline confluency and analyzed using GraphPad Prism. Corrected proliferation % = $100 \pm$ (relative proliferation in indicated siRNA - proliferation in control siRNA (consi))/knockdown efficiency. (B) MCF7 cells were transfected with indicated siRNAs, then reseeded after 16 hours for colony formation (CF) assay. At day 14, colonies were fixed with methanol, stained with crystal violet, scanned and batch analyzed by ImageJ. Corrected CF efficiency (CFE) % = $100 \pm$ (relative CFE in indicated siRNA - CFE in control siRNA (consi))/knockdown efficiency. Error bars, SD ($N=2$). P -values were determined by one-way ANOVA followed by Dunnett's multiple comparisons test: * P -value < 0.05. NTC: non-target control.

Table 1. Fourteen expression-trait associations for genes located at genomic loci at least 500 kb away from any GWAS-identified breast cancer risk variants

Region	Gene ^a	Type ^b	Z score	P value ^c	R ^{2c}	Closest risk SNP ^d	Distance to the closest risk SNP (kb)	P value after adjusting for adjacent risk SNPs ^e
1p34.1	ZSWIM5	Protein	5.26	1.43×10^{-7}	0.17	rs1707302	829	0.006
3p24.1	LRRC3B	Protein	-9.57	1.11×10^{-21}	0.17	rs653465	591	1.60×10^{-6}
4q12	SPATA18	Protein	-4.62	3.86×10^{-6}	0.11	rs6815814	14,101	3.98×10^{-6}
6p22.1	UBD	Protein	-4.87	1.10×10^{-6}	0.13	rs9257408	597	0.94
7q32.2	KLHDC10	Protein	5.21	1.92×10^{-7}	0.14	rs4593472	892	2.90×10^{-7}
9p21.3	MIR31HG	lncRNA	-5.02	5.22×10^{-7}	0.12	rs1011970	502	1.23×10^{-7}
11p15.5	RIC8A	Protein	-5.27	1.40×10^{-7}	0.15	rs6597981	588	4.95×10^{-6}
11q13.2	B3GNT1	Protein	-5.85	4.88×10^{-9}	0.09	rs3903072	530	3.50×10^{-6}
11q13.2	RP11-867G23.10	transcript	4.71	2.49×10^{-6}	0.03	rs3903072	594	2.61×10^{-4}
12p13.33	RP11-218M22.1	lncRNA	5.02	5.27×10^{-7}	0.19	rs12422552	13,641	5.17×10^{-7}
14q24.1	GALNT16	Protein	-8.27	1.38×10^{-16}	0.04	rs999737	691	8.57×10^{-4}
14q24.1	PLEKHD1	Protein	7.50	6.55×10^{-14}	0.02	rs999737	917	0.12
15q24.2	MAN2C1 ^f	Protein	-5.32	1.02×10^{-7}	0.39	rs2290203	15,851	9.56×10^{-8}
15q24.2	CTD-2323K18.1 ^f	lncRNA	-4.65	3.27×10^{-6}	0.07	rs2290203	15,619	3.16×10^{-6}

^a Genes that were siRNA-silenced for functional assays are bolded; SNPs used to predict gene expression are listed in the Supplementary Table 13

^b Protein: protein coding genes; lncRNA: long non-coding RNAs; transcript: processed transcript

^c P value: derived from association analyses; associations with $p \leq 5.82 \times 10^{-6}$ considered statistically significant based on Bonferroni correction of 8,597 tests (0.05/8,597); R²: prediction performance (R²) derived using GTEx data.

^d Risk SNPs identified in previous GWAS or fine-mapping studies. The risk SNP closest to the gene is presented. A full list of all risk SNPs, and their distances to the genes are presented in the **Supplementary Table 4**

^e Use of COJO method³⁶

^f Predicted expression of *MAN2C1* and *CTD-2323K18.1* was correlated (spearman R=0.76)

Table 2. Twenty-three expression-trait associations for genes located at genomic loci within 500 kb of any previous GWAS-identified breast cancer risk variants but not yet implicated as target genes of risk variants[#]

Region	Gene ^a	Type ^b	Z score	P value ^c	R ^{2c}	Closest risk SNP ^d	Distance to the closest risk SNP (kb)	P value after adjusting for adjacent risk SNPs ^e
1p11.2	<i>RP11-439A17.7</i>	lncRNA	-5.34	9.07×10^{-8}	0.22	rs11249433	442	0.02
1q21.1	<i>NUDT17</i>	Protein	-6.27	3.58×10^{-10}	0.01	rs12405132	56	0.08
1q21.1	<i>ANKRD34A</i>	Protein	-5.05	4.42×10^{-7}	0.01	rs12405132	169	4.28×10^{-5}
2p23.1-2p23.2	<i>ALK</i>	Protein	4.67	3.06×10^{-6}	0.06	rs4577244	295	2.70×10^{-6}
3p21.31	<i>PRSS46</i>	Protein	-5.83	5.68×10^{-9}	0.13	rs6796502	89	0.002
3q12.2	<i>RP11-114I8.4</i>	lncRNA	-5.84	5.19×10^{-9}	0.02	rs9833888	356	0.09
5p12	<i>RP11-53O19.1</i>	lncRNA	10.38	2.94×10^{-25}	0.03	rs10941679	39	7.46×10^{-4}
5q33.3	<i>UBLCP1</i>	Protein	5.93	3.04×10^{-9}	0.07	rs1432679	446	0.37
5q33.3	<i>RP11-32D16.1</i>	lncRNA	-5.41	6.37×10^{-8}	0.09	rs1432679	283	1.32×10^{-4}
6p22.2	<i>BTN3A2</i>	Protein	4.61	3.97×10^{-6}	0.28	rs71557345	229	0.72
6q23.1	<i>RP11-73O6.3</i> [†]	lncRNA	-6.61	3.74×10^{-11}	0.11	rs6569648	105	0.41
11p15.5	<i>AP006621.6</i> ^g	lncRNA	5.61	2.01×10^{-8}	0.34	rs6597981	21	0.52
11p15.5	<i>RPLP2</i> ^g	Protein	4.64	3.46×10^{-6}	0.27	rs6597981	7	0.51
14q32.33	<i>CTD-3051D23.1</i>	lncRNA	-5.06	4.21×10^{-7}	0.05	rs10623258	97	7.05×10^{-7}
16q12.2	<i>RP11-467J12.4</i>	lncRNA	8.04	9.02×10^{-16}	0.23	rs3112612	434	0.79
16q12.2	<i>CTD-3032H12.1</i>	lncRNA	4.92	8.58×10^{-7}	0.03	rs28539243	290	0.006
17q21.31	<i>LRRC37A</i> ^g	Protein	-5.89	3.85×10^{-9}	0.43	rs2532263	118	0.79
17q21.31	<i>KANSL1-AS1</i> ^g	lncRNA	-5.58	2.44×10^{-8}	0.62	rs2532263	18	0.95
17q21.31	<i>CRHR1</i> ^g	Protein	-5.29	1.22×10^{-7}	0.22	rs2532263	339	0.99
17q21.31	<i>LINC00671</i>	lncRNA	-5.85	4.95×10^{-9}	0.07	rs72826962	190	0.26
17q21.31	<i>LRRC37A2</i>	Protein	-5.77	7.93×10^{-9}	0.46	rs2532263	336	0.93
19p13.11	<i>HAPLN4</i>	Protein	-7.13	9.88×10^{-13}	0.02	rs2965183	172	0.22
19q13.31	<i>RP11-15A1.7</i> ^h	lncRNA	5.45	5.06×10^{-8}	0.02	rs3760982	215	0.28

[#] not yet reported from eQTL and/or functional studies as target genes of GWAS-identified risk variants and not harbor GWAS or fine-mapping identified risk variants

^a Genes that were siRNA-silenced for functional assays are bolded; SNPs used to predict gene expression are listed in the Supplementary Table 13

^b Protein: protein coding genes; lncRNA: long non-coding RNAs

^c *P* value: nominal *P* value from association analysis; the threshold after Bonferroni correction of 8,597 tests ($0.05/8,597=5.82\times10^{-6}$) was used; *R*²: prediction performance (*R*²) derived using GTEx data

^d Risk SNPs identified in previous GWAS or fine-mapping studies. The risk SNP closest to the gene is presented. A full list of all risk SNPs, and their distances to the genes are presented in the **Supplementary Table 4**

^e Use of COJO method³⁶; all index SNPs in the corresponding region were adjusted in the conditional analyses

^f Predicted expression of *RP11-73O6.3* and *L3MBTL3* was correlated (spearman *R*=0.88)

^g Predicted expression of *AP006621.6* and *RPLP2* was correlated; predicted expression of *LRRC37A*, *KANSL1-AS1*, and *CRHR1* was correlated (spearman *R*>0.1)

^h Predicted expression of *RP11-15A1.7* and *ZNF404* was correlated (spearman *R*=0.64)

Table 3. Eleven expression-trait associations for genes previously reported as potential target genes of GWAS-identified breast cancer risk variants or genes harboring risk variants

Region	Gene ^a	Type ^b	Z score	P value ^c	R ^{2c}	Closest risk SNP ^d	Distance to the closest risk SNP (kb)	P value after adjusting for adjacent risk SNPs ^e	Association direction reported previously ^f	Reference
1p36.13	<i>KLHDC7A</i>	Protein	-5.67	1.40×10^{-8}	0.04	rs2992756	0.085	0.06	-	⁷
2q33.1	<i>ALS2CRI2</i>	Protein	6.70	2.11×10^{-11}	0.10	rs1830298	intron of the gene	0.17	NA	³¹
2q33.1	<i>CASP8</i>	Protein	-8.05	8.51×10^{-16}	0.22	rs3769821	intron of the gene	0.16	-	^{31,32}
5q14.1	<i>ATG10</i>	Protein	-6.65	2.85×10^{-11}	0.51	rs7707921	intron of the gene	0.21	NA	⁹
5q14.2	<i>ATP6AP1L</i>	Protein	-4.98	6.32×10^{-7}	0.63	rs7707921	37	0.98	NA	⁹
6q23.1	<i>L3MBTL3</i> ^g	Protein	-6.69	2.27×10^{-11}	0.10	rs6569648	208	0.44	NA	⁶
6q25.1	<i>RMND1</i>	Protein	4.76	1.95×10^{-6}	0.13	rs3757322	169	1.11×10^{-4}	mixed	¹⁷
11q13.1	<i>SNX32</i>	Protein	4.70	2.60×10^{-6}	0.19	rs3903072	18	0.17	NA	³³
15q26.1	<i>RCCD1</i>	Protein	-7.18	7.23×10^{-13}	0.13	rs2290203	6	1.66×10^{-4}	-	¹⁰
17q22	<i>STXBP4</i>	Protein	6.69	2.21×10^{-11}	0.03	rs6504950	intron of the gene	0.90	+ in GTEx	^{34,35}
19q13.31	<i>ZNF404</i> ^h	Protein	7.42	1.15×10^{-13}	0.15	rs3760982	90	0.005	NA	⁸

^a Genes that were siRNA silenced for functional assays are bolded; SNPs used to predict gene expression are listed in the Supplementary Table 13

^b Protein: protein coding genes; lncRNA: long non-coding RNAs; NA: not available

^c P value: nominal P value from association analysis; the threshold after Bonferroni correction of 8,597 tests ($0.05/8,597=5.82 \times 10^{-6}$) was used; R²: prediction performance (R²) derived using GTEx data .

^d Risk SNPs identified in previous GWAS or fine-mapping studies. The risk SNP closest to the gene is presented. A full list of all risk SNPs, and their distances to the genes are presented in the **Supplementary Table 4**

^e Use of COJO method³⁶; all index SNPs in the corresponding region were adjusted for the conditional analyses

^f -: inverse association; +: positive association; mixed: both inverse and positive associations reported; NA: not available

^g Predicted expression of *L3MBTL3* and *RP11-73O6.3* was correlated (spearman R=0.88)

^h Predicted expression of *ZNF404* and *RP11-15A1.7* was correlated (spearman R=0.64)

Table 4. Genes at GWAS-identified breast cancer risk loci ($\pm 500\text{kb}$ of the index SNPs) whose predicted expression levels were associated with breast cancer risk at p -values between 5.82×10^{-6} and 1.05×10^{-3} (FDR corrected p -value ≤ 0.05)

Region	Gene	Type ^a	Z score	P value ^b	R^{2b}	Closest risk SNP ^c	Distance to the closest risk SNP (kb)	P value after adjusting for adjacent risk SNPs ^d
1p34.1	<i>UQCRH</i>	Protein	-3.90	9.51×10^{-5}	0.12	rs1707302	168	0.06
1p22.3	<i>LMO4</i>	Protein	-3.76	1.73×10^{-4}	0.09	rs12118297	15	0.002
2p23.3	<i>DNAJC27-AS1</i>	lncRNA	3.84	1.24×10^{-4}	0.03	rs6725517	65	0.13
4p14	<i>KLHL5</i>	Protein	3.52	4.35×10^{-4}	0.13	rs6815814	230	0.03
5q11.2	<i>AC008391.1</i>	miRNA	-4.03	5.60×10^{-5}	0.13	rs16886113	242	0.76
6p22.1	<i>HCG14</i>	lncRNA	-3.47	5.19×10^{-4}	0.11	rs9257408	61	0.03
6p22.2	<i>TRNAI2</i>	miRNA	-3.71	2.09×10^{-4}	0.02	rs71557345	307	0.007
6q25.1	<i>MTHFD1L</i>	Protein	3.85	1.17×10^{-4}	0.10	rs3757318	491	2.36×10^{-4}
8q24.21	<i>PVT1</i>	transcript	3.85	1.20×10^{-4}	0.03	rs11780156	81	1.09×10^{-4}
9q33.3	<i>RP11-123K19.1</i>	lncRNA	-4.10	4.05×10^{-5}	0.05	rs10760444	20	1.26×10^{-4}
10q25.2	<i>RP11-57H14.3</i>	lncRNA	3.42	6.16×10^{-4}	0.08	rs7904519	108	0.002
10q26.13	<i>RP11-500G22.2</i>	lncRNA	4.48	7.54×10^{-6}	0.15	rs2981582	336	0.91
11p15.5	<i>PTDSS2</i>	Protein	-3.47	5.16×10^{-4}	0.04	rs6597981	312	0.02
11p15.5	<i>AP006621.5</i>	Protein	4.35	1.37×10^{-5}	0.51	rs6597981	19	0.01
11p15.5	<i>PIDD1</i>	Protein	4.24	2.28×10^{-5}	0.45	rs6597981	intron of the gene	0.12
11p15.5	<i>MRPL23-AS1</i>	lncRNA	-3.86	1.12×10^{-4}	0.10	rs3817198	95	0.06
11q13.1-11q13.2	<i>PACSI</i>	Protein	-3.59	3.36×10^{-4}	0.06	rs3903072	255	0.001
12p11.22	<i>RP11-860B13.1</i>	lncRNA	3.46	5.42×10^{-4}	0.17	rs10771399	221	0.86
13q22.1	<i>KLF5</i>	Protein	-4.08	4.44×10^{-5}	0.22	rs6562760	306	NA
14q24.1	<i>CTD-2566J3.1</i>	lncRNA	-3.84	1.22×10^{-4}	0.04	rs2588809	64	0.55
14q32.33	<i>C14orf79</i>	Protein	4.37	1.22×10^{-5}	0.11	rs10623258	240	0.91
15q26.1	<i>FES</i>	Protein	4.37	1.26×10^{-5}	0.21	rs2290203	73	3.04×10^{-6}
16q12.2	<i>BBS2</i>	Protein	3.97	7.23×10^{-5}	0.26	rs2432539	80	0.36
16q12.2	<i>CRNDE</i>	lncRNA	3.28	1.05×10^{-3}	0.02	rs28539243	271	0.69
16q24.2	<i>RP11-482M8.1</i>	lncRNA	3.32	9.16×10^{-4}	0.02	rs4496150	441	0.19

17q11.2	<i>GOSR1</i>	Protein	3.79	1.51×10^{-4}	0.10	rs146699004	376	0.04
17q21.2	<i>ATP6V0A1</i>	Protein	3.61	3.02×10^{-4}	0.03	rs72826962	162	0.01
17q21.2	<i>RP11-400F19.8</i>	transcript	-3.96	7.65×10^{-5}	0.01	rs72826962	122	6.62×10^{-4}
17q21.31	<i>RP11-105N13.4</i>	transcript	-4.51	6.46×10^{-6}	0.02	rs2532263	359	NA
17q25.3	<i>CBX8</i>	Protein	4.38	1.16×10^{-5}	0.05	rs745570	6	0.99
19p13.11	<i>CTD-2538G9.5</i>	lncRNA	3.56	3.76×10^{-4}	0.01	rs8170	432	4.38×10^{-4}
19p13.11	<i>HOMER3</i>	Protein	-3.87	1.08×10^{-4}	0.10	rs4808801	469	0.18
20q11.22	<i>CTD-3216D2.5</i>	lncRNA	4.03	5.60×10^{-5}	0.16	rs2284378	281	9.24×10^{-4}
22q13.1	<i>TRIOBP</i>	Protein	3.34	8.34×10^{-4}	0.07	rs738321	396	0.003
22q13.1	<i>RP5-1039K5.13</i>	lncRNA	3.73	1.93×10^{-4}	0.01	rs738321	99	0.053
22q13.1	<i>CBY1</i>	Protein	3.91	9.34×10^{-5}	0.05	chr22:39359355	289	0.06
22q13.1	<i>APOBEC3A</i>	Protein	-4.11	3.98×10^{-5}	0.07	chr22:39359355	0.2	0.02
22q13.2	<i>RPI-85F18.6</i>	lncRNA	3.52	4.28×10^{-4}	0.12	rs73161324	460	0.72

^a Protein: protein coding genes; lncRNA: long non-coding RNAs; transcript: processed transcript

^b *P* value: nominal *P* value from association analysis; *R*²: prediction performance derived using GTEx data.

^c Risk SNPs identified in previous GWAS or fine-mapping studies. The risk SNP closest to the gene is presented. A full list of all risk SNPs, and their distances to the genes are presented in the **Supplementary Table 4**

^d Use of COJO method³⁶; all index SNPs in the corresponding region were adjusted for the conditional analyses

Supplementary Material

Identification of novel susceptibility loci and genes for breast cancer risk: A transcriptome-wide association study of 229,000 women of European descent

Lang Wu^{1,160}, Wei Shi^{2,160}, Jirong Long¹, Xingyi Guo¹, Kyriaki Michailidou^{3,4}, Jonathan Beesley², Manjeet K. Bolla³, Xiao-Ou Shu¹, Yingchang Lu¹, Qiuyin Cai¹, Fares Al-Ejeh², Esdy Rozali², Qin Wang³, Joe Dennis³, Bingshan Li¹⁵¹, Chenjie Zeng¹, Helian Feng^{5,6}, Alexander Gusev^{153, 154, 155}, Richard T. Barfield⁵, Irene L. Andrulis^{7,8}, Hoda Anton-Culver⁹, Volker Arndt¹⁰, Kristan J. Aronson¹¹, Paul L. Auer^{12,13}, Myrto Barrdahl¹⁴, Caroline Baynes¹⁵, Matthias W. Beckmann¹⁶, Javier Benitez^{17,18}, Marina Bermisheva^{19,20}, Carl Blomqvist^{21,159}, Natalia V. Bogdanova^{20,22,23}, Stig E. Bojesen²⁴⁻²⁶, Hiltrud Brauch²⁷⁻²⁹, Hermann Brenner^{10,29,30}, Louise Brinton³¹, Per Broberg³², Sara Y. Brucker³³, Barbara Burwinkel^{34,35}, Trinidad Caldés³⁶, Federico Canzian³⁷, Brian D. Carter³⁸, J. Esteban Castelao³⁹, Jenny Chang-Claude^{14,40}, Xiaoqing Chen², Ting-Yuan David Cheng⁴¹, Hans Christiansen²², Christine L. Clarke⁴², NBCS Collaborators^{43-120,44-124,45}, Margriet Collée⁴⁶, Sten Cornelissen⁴⁷, Fergus J. Couch⁴⁸, David Cox^{49,50}, Angela Cox⁵¹, Simon S. Cross⁵², Julie M. Cunningham⁴⁸, Kamila Czene⁵³, Mary B. Daly⁵⁴, Peter Devilee^{55,56}, Kimberly F. Doherty⁵⁷, Thilo Dörk²⁰, Isabel dos-Santos-Silva⁵⁸, Martine Dumont⁵⁹, Miriam Dwek⁶⁰, Diana M. Eccles⁶¹, Ursula Eilber¹⁴, A. Heather Eliassen^{6,62}, Christoph Engel⁶³, Mikael Eriksson⁵³, Laura Fachal¹⁵, Peter A. Fasching^{16,64}, Jonine Figueroa^{31,65}, Dieter Flesch-Janys^{66,67}, Olivia Fletcher⁶⁸, Henrik Flyger⁶⁹, Lin Fritschi⁷⁰, Marike Gabrielson⁵³, Manuela Gago-Dominguez^{71,72}, Susan M. Gapstur³⁸, Montserrat García-Closas³¹, Mia M. Gaudet³⁸, Maya Ghoussaini¹⁵, Graham G. Giles^{73,74}, Mark S. Goldberg^{75,76}, David E. Goldgar⁷⁷, Anna González-Neira¹⁷, Pascal Guénel⁷⁸, Eric Hahnen⁷⁹⁻⁸¹, Christopher A. Haiman⁸², Niclas Håkansson⁸³, Per Hall⁵³, Emily Hallberg⁸⁴, Ute Hamann⁸⁵, Patricia Harrington¹⁵, Alexander Hein¹⁶, Belynda Hicks⁸⁶, Peter Hillemanns²⁰, Antoinette Hollestelle⁸⁷, Robert N. Hoover³¹, John L. Hopper⁷⁴, Guanmengqian Huang⁸⁵, Keith Humphreys⁵³, David J. Hunter^{6,158}, Anna Jakubowska⁸⁸, Wolfgang Janni⁸⁹, Esther M. John⁹⁰⁻⁹², Nichola Johnson⁶⁸, Kristine Jones⁸⁶, Michael E. Jones⁹³, Audrey Jung¹⁴, Rudolf Kaaks¹⁴, Michael J. Kerin⁹⁴, Elza Khusnutdinova^{19,95}, Veli-Matti Kosma⁹⁶⁻⁹⁸, Vessela N. Kristensen⁹⁹⁻¹⁰¹, Diether Lambrechts^{102,103}, Loic Le Marchand¹⁰⁴, Jingmei Li¹⁵⁷, Sara Lindström^{5,105}, Jolanta Lissowska¹⁰⁶, Wing-Yee Lo^{27,28}, Sibylle Loibl¹⁰⁷, Jan Lubinski⁸⁸, Craig Luccarini¹⁵, Michael P. Lux¹⁶, Robert J. MacInnis^{73,74}, Tom Maishman^{61,108}, Ivana Maleva Kostovska^{20,109}, Arto Mannermaa⁹⁶⁻⁹⁸, JoAnn E. Manson^{6,110}, Sara Margolin¹¹¹, Dimitrios Mavroudis¹¹², Hanne Meijers-Heijboer¹⁵², Alfons Meindl¹¹³, Usha Menon¹¹⁴, Jeffery Meyer⁴⁸, Anna Marie Mulligan^{115,116}, Susan L. Neuhausen¹¹⁷, Heli Nevanlinna¹¹⁸, Patrick Neven¹¹⁹, Sune F. Nielsen^{24,25}, Børge G. Nordestgaard²⁴⁻²⁶, Olufunmilayo I. Olopade¹²⁰, Janet E. Olson⁸⁴, Håkan Olsson³², Paolo Peterlongo¹²¹, Julian Peto⁵⁸, Dijana Plaseska-Karanfilska¹⁰⁹, Ross Prentice¹², Nadege Presneau⁶⁰, Katri Pylkäs^{122,123}, Brigitte Rack⁸⁹, Paolo Radice¹²⁵, Nazneen Rahman¹²⁶, Gad Rennert¹²⁷, Hedy S. Rennert¹²⁷, Valerie Rhenius¹⁵, Atocha Romero^{36,128}, Jane Romm⁵⁷, Anja Rudolph¹⁴, Emmanouil Saloustros¹²⁹, Dale P. Sandler¹³⁰, Elinor J. Sawyer¹³¹, Marjanka K. Schmidt^{47,132}, Rita K. Schmutzler⁷⁹⁻⁸¹, Andreas Schneeweiss^{34,133}, Rodney J. Scott^{134,135}, Christopher Scott⁸⁴, Sheila Seal¹²⁶, Mitul Shah¹⁵, Martha J. Shrubsole¹, Ann Smeets¹¹⁹, Melissa C. Southey¹³⁶, John J. Spinelli^{137,138}, Jennifer Stone^{139,140}, Harald

Surowy^{34,35}, Anthony J. Swerdlow^{93,141}, Rulla M. Tamimi^{5,6,62}, William Tapper⁶¹, Jack A. Taylor^{130,142}, Mary Beth Terry¹⁴³, Daniel C. Tessier¹⁴⁴, Abigail Thomas⁸⁴, Kathrin Thöne⁶⁷, Rob A.E.M. Tollenaar¹⁴⁵, Diana Torres^{85,146}, Thérèse Truong⁷⁸, Michael Untch¹⁴⁷, Celine Vachon⁸⁴, David Van Den Berg⁸², Daniel Vincent¹⁴⁴, Quinten Waisfisz¹⁵², Clarice R. Weinberg¹⁴⁸, Camilla Wendt¹¹¹, Alice S. Whittmore^{91,92}, Hans Wildiers¹¹⁹, Walter C. Willett^{6,62,156}, Robert Winqvist^{122,123}, Alicja Wolk⁸³, Lucy Xia⁸², Xiaohong R. Yang³¹, Argyrios Ziogas⁹, Elad Ziv¹⁴⁹, kConFab/AOCS Investigators¹⁵⁰, Alison M. Dunning¹⁵, Paul D.P. Pharoah^{3,15}, Jacques Simard⁵⁹, Roger L. Milne^{73,74}, Stacey L. Edwards², Peter Kraft^{5,6}, Douglas F. Easton^{3,15}, Georgia Chenevix-Trench^{2*}, Wei Zheng^{1*}

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Supplementary Table 1. Internal performance of gene expression prediction models built using GTEx data

Prediction performance (R^2)	All	Protein	lncRNAs	miRNAs	Others*
Number of genes	15,148	10,483	4,277	68	320
0.01 [#]	12,824	8,874	3,628	57	265
0.04	7,655	5,244	2,200	38	173
0.09	3,818	2,601	1,106	19	92
0.16	1,573	1,035	479	8	51

Protein: protein coding genes; lncRNAs: long non-coding RNAs; miRNAs: microRNAs

* Including processed transcripts, immunoglobulin genes, and T cell receptor genes

[#] The R^2 of 0.01 is the internal prediction performance threshold according to which the prediction models were retained for external evaluation in the TCGA data

Supplementary Table 2. Genes with predicted expression levels associated with breast cancer risk at $p < 1.05 \times 10^{-3}$ (the significance level with false discovery rate correction)

Gene	Type*	Z score	P value	R ²	No. of predicting variants used	No. of predicting variants in model	Proportion of predicting variants used (%)
<i>RP11-53019.1</i>	lncRNA	10.38	2.94×10^{-25}	0.03	8	15	53
<i>LRRC3B</i>	Protein	-9.57	1.11×10^{-21}	0.17	46	46	100
<i>GALNT16</i>	Protein	-8.27	1.38×10^{-16}	0.04	53	53	100
<i>CASP8</i>	Protein	-8.05	8.51×10^{-16}	0.22	15	15	100
<i>RP11-467J12.4</i>	lncRNA	8.04	9.02×10^{-16}	0.23	142	142	100
<i>PLEKHD1</i>	Protein	7.50	6.55×10^{-14}	0.02	6	6	100
<i>ZNF404</i>	Protein	7.42	1.15×10^{-13}	0.15	32	32	100
<i>RCCD1</i>	Protein	-7.18	7.23×10^{-13}	0.13	22	22	100
<i>HAPLN4</i>	Protein	-7.13	9.88×10^{-13}	0.02	53	53	100
<i>ALS2CR12</i>	Protein	6.70	2.11×10^{-11}	0.10	4	4	100
<i>STXBP4</i>	Protein	6.69	2.21×10^{-11}	0.03	58	60	97
<i>L3MBTL3</i>	Protein	-6.69	2.27×10^{-11}	0.10	5	5	100
<i>ATG10</i>	Protein	-6.65	2.85×10^{-11}	0.51	57	61	93
<i>RP11-7306.3</i>	lncRNA	-6.61	3.74×10^{-11}	0.11	26	26	100
<i>NUDT17</i>	Protein	-6.27	3.58×10^{-10}	0.01	7	7	100
<i>UBLCP1</i>	Protein	5.93	3.04×10^{-9}	0.07	2	3	67
<i>LRRC37A</i>	Protein	-5.89	3.85×10^{-9}	0.43	31	32	97
<i>B3GNT1</i>	Protein	-5.85	4.88×10^{-9}	0.09	26	27	96
<i>LINC00671</i>	lncRNA	-5.85	4.95×10^{-9}	0.07	1	1	100
<i>RP11-114I8.4</i>	lncRNA	-5.84	5.19×10^{-9}	0.02	14	14	100
<i>PRSS46</i>	Protein	-5.83	5.68×10^{-9}	0.13	45	46	98
<i>LRRC37A2</i>	Protein	-5.77	7.93×10^{-9}	0.46	120	121	99
<i>KLHDC7A</i>	Protein	-5.67	1.40×10^{-8}	0.04	15	15	100
<i>AP006621.6</i>	lncRNA	5.61	2.01×10^{-8}	0.34	41	41	100
<i>KANSL1-AS1</i>	lncRNA	-5.58	2.44×10^{-8}	0.62	70	72	97

<i>RP11-15A1.7</i>	lncRNA	5.45	5.06×10^{-8}	0.02	2	2	100
<i>RP11-32D16.1</i>	lncRNA	-5.41	6.37×10^{-8}	0.09	44	46	96
<i>RP11-439A17.7</i>	lncRNA	-5.34	9.07×10^{-8}	0.22	93	94	99
<i>MAN2C1</i>	Protein	-5.32	1.02×10^{-7}	0.39	27	27	100
<i>CRHR1</i>	Protein	-5.29	1.22×10^{-7}	0.22	31	31	100
<i>RIC8A</i>	Protein	-5.27	1.40×10^{-7}	0.15	15	15	100
<i>ZSWIM5</i>	Protein	5.26	1.43×10^{-7}	0.17	67	67	100
<i>KLHDC10</i>	Protein	5.21	1.92×10^{-7}	0.14	52	53	98
<i>CTD-3051D23.1</i>	lncRNA	-5.06	4.21×10^{-7}	0.05	25	26	96
<i>ANKRD34A</i>	Protein	-5.05	4.42×10^{-7}	0.01	1	1	100
<i>MIR31HG</i>	lncRNA	-5.02	5.22×10^{-7}	0.12	1	1	100
<i>RP11-218M22.1</i>	lncRNA	5.02	5.27×10^{-7}	0.19	47	48	98
<i>ATP6AP1L</i>	Protein	-4.98	6.32×10^{-7}	0.63	64	67	96
<i>CTD-3032H12.1</i>	lncRNA	4.92	8.58×10^{-7}	0.03	23	23	100
<i>UBD</i>	Protein	-4.87	1.10×10^{-6}	0.13	31	31	100
<i>RMND1</i>	Protein	4.76	1.95×10^{-6}	0.13	91	91	100
<i>RP11-867G23.10</i>	transcript	4.71	2.49×10^{-6}	0.03	5	5	100
<i>SNX32</i>	Protein	4.70	2.60×10^{-6}	0.19	17	17	100
<i>ALK</i>	Protein	4.67	3.06×10^{-6}	0.06	47	48	98
<i>CTD-2323K18.1</i>	transcript	-4.65	3.27×10^{-6}	0.07	23	23	100
<i>RPLP2</i>	Protein	4.64	3.46×10^{-6}	0.27	45	45	100
<i>SPATA18</i>	Protein	-4.62	3.86×10^{-6}	0.11	43	43	100
<i>BTN3A2</i>	Protein	4.61	3.97×10^{-6}	0.28	66	66	100
<i>RP11-105N13.4</i>	transcript	-4.51	6.46×10^{-6}	0.02	15	16	94
<i>SLC39A9</i>	Protein	-4.48	7.32×10^{-6}	0.03	24	24	100
<i>RP11-500G22.2</i>	lncRNA	4.48	7.54×10^{-6}	0.15	8	8	100
<i>FAT4</i>	Protein	4.45	8.44×10^{-6}	0.06	42	54	78
<i>CRIP2</i>	Protein	4.44	9.14×10^{-6}	0.03	12	12	100
<i>RP11-432I5.1</i>	lncRNA	4.40	1.06×10^{-5}	0.03	11	14	79
<i>CBX8</i>	Protein	4.38	1.16×10^{-5}	0.05	12	12	100
<i>C14orf79</i>	Protein	4.37	1.22×10^{-5}	0.11	5	5	100
<i>RHOD</i>	Protein	4.37	1.23×10^{-5}	0.03	24	24	100

<i>FES</i>	Protein	4.37	1.26×10^{-5}	0.21	23	23	100
<i>AP006621.5</i>	Protein	4.35	1.37×10^{-5}	0.51	46	46	100
<i>NUP107</i>	Protein	4.30	1.69×10^{-5}	0.14	4	4	100
<i>GSTM4</i>	Protein	-4.29	1.78×10^{-5}	0.06	9	9	100
<i>YBEY</i>	Protein	4.26	2.01×10^{-5}	0.40	27	27	100
<i>PIDD1</i>	Protein	4.24	2.28×10^{-5}	0.45	61	61	100
<i>RP11-126L15.4</i>	lncRNA	-4.19	2.74×10^{-5}	0.05	59	59	100
<i>AC010136.2</i>	lncRNA	-4.14	3.52×10^{-5}	0.21	1	1	100
<i>APOBEC3A</i>	Protein	-4.11	3.98×10^{-5}	0.07	33	33	100
<i>RP11-123K19.1</i>	lncRNA	-4.10	4.05×10^{-5}	0.05	21	21	100
<i>GABPB1-AS1</i>	transcript	4.10	4.21×10^{-5}	0.45	28	28	100
<i>CTD-3110H11.1</i>	lncRNA	4.09	4.31×10^{-5}	0.53	25	26	96
<i>EDEM2</i>	Protein	4.09	4.39×10^{-5}	0.03	58	59	98
<i>KLF5</i>	Protein	-4.08	4.44×10^{-5}	0.22	30	30	100
<i>HSF2</i>	Protein	-4.05	5.02×10^{-5}	0.04	45	45	100
<i>SMN2</i>	Protein	-4.04	5.44×10^{-5}	0.19	33	34	97
<i>XXbac-BPG170G13.32</i>	lncRNA	4.03	5.50×10^{-5}	0.14	50	56	89
<i>AC008391.1</i>	miRNA	-4.03	5.60×10^{-5}	0.13	7	7	100
<i>CTD-3216D2.5</i>	lncRNA	4.03	5.60×10^{-5}	0.16	57	57	100
<i>CPNE1</i>	Protein	-4.02	5.80×10^{-5}	0.33	36	36	100
<i>GSTM3</i>	Protein	-3.98	6.95×10^{-5}	0.18	23	23	100
<i>BBS2</i>	Protein	3.97	7.23×10^{-5}	0.26	20	20	100
<i>RP11-400F19.8</i>	transcript	-3.96	7.65×10^{-5}	0.01	22	26	85
<i>PILRA</i>	Protein	3.94	8.16×10^{-5}	0.54	25	25	100
<i>STAG3L5P-PVRIG2P-PILRB</i>	transcript	3.91	9.27×10^{-5}	0.32	42	43	98
<i>CBY1</i>	Protein	3.91	9.34×10^{-5}	0.05	19	21	90
<i>UQCRH</i>	Protein	-3.90	9.51×10^{-5}	0.12	35	35	100
<i>ALS2CL</i>	Protein	-3.90	9.69×10^{-5}	0.23	1	3	33
<i>ATF4</i>	Protein	-3.90	9.74×10^{-5}	0.11	95	97	98
<i>CCBL2</i>	Protein	3.90	9.78×10^{-5}	0.01	13	17	76
<i>HOMER3</i>	Protein	-3.87	1.08×10^{-4}	0.10	16	16	100

<i>CMTR2</i>	Protein	-3.86	1.11×10^{-4}	0.01	22	38	58
<i>MRPL23-AS1</i>	lncRNA	-3.86	1.12×10^{-4}	0.10	13	14	93
<i>ARHGEF19</i>	Protein	-3.86	1.15×10^{-4}	0.13	95	96	99
<i>NNT-AS1</i>	lncRNA	3.86	1.15×10^{-4}	0.06	40	40	100
<i>MTHFD1L</i>	Protein	3.85	1.17×10^{-4}	0.10	24	24	100
<i>PVT1</i>	transcript	3.85	1.20×10^{-4}	0.03	14	17	82
<i>CTD-2566J3.1</i>	lncRNA	-3.84	1.22×10^{-4}	0.04	16	16	100
<i>PDLIM4</i>	Protein	-3.84	1.22×10^{-4}	0.08	42	43	98
<i>MYRF</i>	Protein	3.84	1.24×10^{-4}	0.01	10	10	100
<i>DNAJC27-AS1</i>	lncRNA	3.84	1.24×10^{-4}	0.03	22	22	100
<i>ATP5I</i>	Protein	-3.82	1.34×10^{-4}	0.02	9	9	100
<i>GOSR1</i>	Protein	3.79	1.51×10^{-4}	0.10	13	13	100
<i>RP11-335O13.7</i>	lncRNA	-3.77	1.63×10^{-4}	0.08	34	34	100
<i>RP11-550I24.2</i>	transcript	-3.76	1.67×10^{-4}	0.05	61	61	100
<i>LMO4</i>	Protein	-3.76	1.73×10^{-4}	0.09	1	1	100
<i>RP5-1039K5.13</i>	lncRNA	3.73	1.93×10^{-4}	0.01	37	38	97
<i>TRNAI2</i>	miRNA	-3.71	2.09×10^{-4}	0.02	12	12	100
<i>RP4-625H18.2</i>	lncRNA	-3.70	2.12×10^{-4}	0.02	5	5	100
<i>ZNF334</i>	Protein	-3.69	2.22×10^{-4}	0.12	55	55	100
<i>PILRB</i>	Protein	3.68	2.29×10^{-4}	0.30	70	71	99
<i>METTL10</i>	Protein	-3.68	2.35×10^{-4}	0.17	25	25	100
<i>SH3TC2</i>	Protein	3.67	2.42×10^{-4}	0.09	42	43	98
<i>CTD-2026K11.3</i>	lncRNA	3.67	2.46×10^{-4}	0.01	20	20	100
<i>CTD-2026K11.2</i>	lncRNA	3.66	2.52×10^{-4}	0.12	109	130	84
<i>TMC4</i>	Protein	3.66	2.54×10^{-4}	0.21	6	6	100
<i>RP5-1139B12.4</i>	lncRNA	-3.66	2.55×10^{-4}	0.17	47	47	100
<i>TBX5</i>	Protein	3.64	2.73×10^{-4}	0.11	85	85	100
<i>SNUPN</i>	Protein	-3.63	2.86×10^{-4}	0.03	4	4	100
<i>RP11-1055B8.4</i>	lncRNA	3.62	2.92×10^{-4}	0.20	5	5	100
<i>PSORS1C2</i>	Protein	3.62	2.96×10^{-4}	0.41	29	32	91
<i>IST1</i>	Protein	3.62	3.00×10^{-4}	0.01	18	18	100
<i>ATP6V0A1</i>	Protein	3.61	3.02×10^{-4}	0.03	98	99	99

<i>KLC1</i>	Protein	-3.61	3.08×10^{-4}	0.07	37	37	100
<i>GPR144</i>	Protein	3.59	3.31×10^{-4}	0.12	53	75	71
<i>PACS1</i>	Protein	-3.59	3.36×10^{-4}	0.06	49	49	100
<i>ECT2L</i>	Protein	3.58	3.47×10^{-4}	0.14	3	3	100
<i>CTD-2538G9.5</i>	lncRNA	3.56	3.76×10^{-4}	0.01	7	7	100
<i>AZGP1</i>	Protein	-3.55	3.79×10^{-4}	0.03	5	5	100
<i>OXLD1</i>	Protein	3.55	3.86×10^{-4}	0.15	31	31	100
<i>CPLX1</i>	Protein	-3.54	4.03×10^{-4}	0.05	17	17	100
<i>DGKQ</i>	Protein	3.54	4.06×10^{-4}	0.25	85	85	100
<i>RP11-757G1.6</i>	lncRNA	3.53	4.17×10^{-4}	0.19	33	33	100
<i>CTA-109P11.4</i>	lncRNA	-3.52	4.26×10^{-4}	0.10	10	10	100
<i>RP1-85F18.6</i>	lncRNA	3.52	4.28×10^{-4}	0.12	88	88	100
<i>TBX5-AS1</i>	lncRNA	3.52	4.31×10^{-4}	0.09	55	61	90
<i>KLHL5</i>	Protein	3.52	4.35×10^{-4}	0.13	106	109	97
<i>MUTYH</i>	Protein	3.51	4.47×10^{-4}	0.04	12	12	100
<i>TRIM4</i>	Protein	-3.50	4.64×10^{-4}	0.43	72	74	97
<i>MIR1909</i>	miRNA	3.50	4.68×10^{-4}	0.04	33	34	97
<i>SLC22A5</i>	Protein	-3.50	4.72×10^{-4}	0.19	28	28	100
<i>CCDC18</i>	Protein	-3.48	5.08×10^{-4}	0.38	94	94	100
<i>PTDSS2</i>	Protein	-3.47	5.16×10^{-4}	0.04	31	31	100
<i>HCG14</i>	lncRNA	-3.47	5.19×10^{-4}	0.11	2	2	100
<i>SMIM8</i>	Protein	3.47	5.20×10^{-4}	0.06	20	20	100
<i>MAP3K14-AS1</i>	lncRNA	-3.46	5.31×10^{-4}	0.04	3	3	100
<i>FAM149B1</i>	Protein	-3.46	5.35×10^{-4}	0.03	12	12	100
<i>RP11-860B13.1</i>	lncRNA	3.46	5.42×10^{-4}	0.17	14	14	100
<i>PAIP1</i>	Protein	-3.45	5.67×10^{-4}	0.02	2	2	100
<i>GSTM5</i>	Protein	-3.44	5.92×10^{-4}	0.28	20	20	100
<i>RP11-57H14.3</i>	lncRNA	3.42	6.16×10^{-4}	0.08	2	2	100
<i>BRMS1</i>	Protein	-3.40	6.62×10^{-4}	0.05	7	7	100
<i>KDM6B</i>	Protein	-3.40	6.73×10^{-4}	0.07	36	52	69
<i>IGKV2D-24</i>	IG_gene	-3.40	6.74×10^{-4}	0.02	1	1	100
<i>RP11-174G6.5</i>	lncRNA	3.39	7.00×10^{-4}	0.05	26	27	96

<i>POLR2J</i>	Protein	-3.39	7.01×10^{-4}	0.28	86	86	100
<i>RP11-580I16.2</i>	lncRNA	3.38	7.17×10^{-4}	0.04	4	4	100
<i>RP13-20L14.1</i>	lncRNA	-3.37	7.52×10^{-4}	0.02	8	9	89
<i>RP11-553A10.1</i>	Protein	3.36	7.76×10^{-4}	0.03	31	33	94
<i>RP11-363E6.3</i>	lncRNA	-3.36	7.83×10^{-4}	0.05	37	37	100
<i>TSPAN5</i>	Protein	-3.35	8.11×10^{-4}	0.04	12	12	100
<i>PSORS1C1</i>	Protein	3.34	8.28×10^{-4}	0.35	17	20	85
<i>TRIOBP</i>	Protein	3.34	8.34×10^{-4}	0.07	22	23	96
<i>CLEC18A</i>	Protein	-3.34	8.37×10^{-4}	0.43	32	32	100
<i>DFNA5</i>	Protein	-3.33	8.55×10^{-4}	0.19	28	28	100
<i>TMEM136</i>	Protein	3.33	8.56×10^{-4}	0.07	68	78	87
<i>C9orf3</i>	Protein	3.33	8.64×10^{-4}	0.03	23	26	88
<i>GPR156</i>	Protein	3.33	8.67×10^{-4}	0.19	69	71	97
<i>IL10RB-AS1</i>	lncRNA	-3.33	8.68×10^{-4}	0.17	91	92	99
<i>BDH2</i>	Protein	-3.33	8.72×10^{-4}	0.23	41	41	100
<i>ZNF165</i>	Protein	3.33	8.76×10^{-4}	0.06	17	17	100
<i>LINC00092</i>	lncRNA	-3.32	9.03×10^{-4}	0.08	43	43	100
<i>RP11-482M8.1</i>	lncRNA	3.32	9.16×10^{-4}	0.02	37	37	100
<i>USP19</i>	Protein	-3.31	9.28×10^{-4}	0.02	5	6	83
<i>MMP24</i>	Protein	-3.31	9.40×10^{-4}	0.13	2	2	100
<i>CTD-2196P11.2</i>	lncRNA	3.29	1.01×10^{-3}	0.04	28	29	97
<i>NR1H3</i>	Protein	3.29	1.01×10^{-3}	0.17	52	53	98
<i>FLOT1</i>	Protein	-3.28	1.03×10^{-3}	0.10	60	63	95
<i>BAZ1B</i>	Protein	-3.28	1.04×10^{-3}	0.14	63	63	100
<i>AH11</i>	Protein	3.28	1.05×10^{-3}	0.23	13	14	93
<i>CRNDE</i>	lncRNA	3.28	1.05×10^{-3}	0.02	22	25	88
<i>AL450992.2</i>	lncRNA	-3.28	1.05×10^{-3}	0.03	6	6	100

* Protein: protein coding genes; lncRNA: long non-coding RNAs; miRNA: microRNA; transcript: processed transcript; IG_gene: immunoglobulin genes.

P value: nominal *p* value from association analysis; R^2 : prediction performance (R^2) derived using GTEx data.

Supplementary Table 3. Associations of predicted expression of identified genes with breast cancer risk in each of the three assessed datasets (OncoArray, iCOGS, and GWAS sets)

Gene name	OncoArray z-score	OncoArray <i>p</i> -value	iCOGS z-score	iCOGS <i>p</i> -value	GWAS z-score	GWAS <i>p</i> -value	Cochran's Q	I ²
Table 1								
<i>ZSWIM5</i>	2.98	0.003	4.32	1.57×10^{-5}	1.39	0.17	0.32	0
<i>LRRC3B</i>	-7.48	7.19×10^{-14}	-4.89	1.02×10^{-6}	-3.61	3.11×10^{-4}	2.09	0.04
<i>SPATA18</i>	-3.09	0.002	-2.59	0.01	-2.33	0.02	0.21	0
<i>UBD</i>	-1.55	0.12	-4.07	4.67×10^{-5}	-3.46	5.48×10^{-4}	1.54	0
<i>KLHDC10</i>	2.15	0.03	4.39	1.16×10^{-5}	2.87	0.004	0.92	0
<i>MIR31HG</i>	-4.35	1.35×10^{-5}	-2.90	0.004	-0.53	0.60	0.98	0
<i>RIC8A</i>	-3.28	0.001	-3.12	0.002	-2.71	0.007	0.17	0
<i>B3GNT1</i>	-2.70	0.007	-5.00	5.83×10^{-7}	-2.38	0.02	0.82	0
<i>RP11-867G23.10</i>	2.78	0.005	3.13	0.002	2.18	0.03	0.04	0
<i>RP11-218M22.1</i>	3.84	1.22×10^{-4}	3.33	8.82×10^{-4}	0.86	0.39	0.35	0
<i>GALNT16</i>	-4.45	8.74×10^{-6}	-6.17	6.82×10^{-10}	-3.67	2.40×10^{-4}	0.38	0
<i>PLEKHD1</i>	5.21	1.85×10^{-7}	3.96	7.43×10^{-5}	3.96	7.36×10^{-5}	0.90	0
<i>MAN2C1</i>	-4.08	4.47×10^{-5}	-3.49	4.88×10^{-4}	-0.86	0.39	0.43	0
<i>CTD-2323K18.1</i>	-3.69	2.23×10^{-4}	-2.62	0.009	-1.27	0.21	0.42	0
Table 2								
<i>RP11-439A17.7</i>	-4.35	1.37×10^{-5}	-3.39	6.90×10^{-4}	-0.32	0.75	0.88	0
<i>NUDT17</i>	-3.53	4.19×10^{-4}	-4.99	5.91×10^{-7}	-1.98	0.047	0.30	0
<i>ANKRD34A</i>	-4.27	1.97×10^{-5}	-2.54	0.01	-1.13	0.26	0.94	0
<i>ALK</i>	3.84	1.23×10^{-4}	3.23	0.001	-0.08	0.94	0.84	0
<i>PRSS46</i>	-4.33	1.51×10^{-5}	-3.51	4.41×10^{-4}	-1.78	0.08	0.31	0
<i>RP11-114I8.4</i>	-4.20	2.66×10^{-5}	-3.15	0.002	-2.74	0.006	0.48	0
<i>RP11-53O19.1</i>	8.29	1.17×10^{-16}	5.75	8.85×10^{-9}	3.23	0.001	2.16	0.07
<i>UBLCP1</i>	4.72	2.34×10^{-6}	3.12	0.002	1.98	0.047	0.80	0
<i>RP11-32D16.1</i>	-3.75	1.75×10^{-4}	-3.66	2.51×10^{-4}	-1.53	0.13	0.09	0
<i>BTN3A2</i>	3.16	0.002	2.74	0.006	2.06	0.04	0.12	0
<i>RP11-73O6.3</i>	-5.34	9.31×10^{-8}	-2.24	0.03	-4.32	1.53×10^{-5}	3.39	0.41

<i>AP006621.6</i>	3.58	3.40×10^{-4}	3.92	8.75×10^{-5}	1.98	0.048	0.01	0
<i>RPLP2</i>	3.43	5.93×10^{-4}	2.77	0.006	1.57	0.12	0.19	0
<i>CTD-3051D23.1</i>	-2.60	0.009	-3.36	7.85×10^{-4}	-3.34	8.30×10^{-4}	0.45	0
<i>RP11-467J12.4</i>	5.75	8.73×10^{-9}	5.41	6.28×10^{-8}	1.93	0.054	0.38	0
<i>CTD-3032H12.1</i>	2.93	0.003	2.95	0.003	2.60	0.009	0.15	0
<i>LRRC37A</i>	-4.13	3.56×10^{-5}	-3.08	0.002	-3.07	0.002	0.58	0
<i>KANSL1-AS1</i>	-3.83	1.28×10^{-4}	-3.17	0.002	-2.61	0.009	0.27	0
<i>CRHR1</i>	-3.58	3.39×10^{-4}	-2.81	0.005	-2.93	0.003	0.45	0
<i>LINC00671</i>	-4.40	1.11×10^{-5}	-4.15	3.32×10^{-5}	-0.25	0.80	0.82	0
<i>LRRC37A2</i>	-3.93	8.47×10^{-5}	-3.18	0.001	-2.96	0.003	0.39	0
<i>HAPLN4</i>	-5.49	4.01×10^{-8}	-5.10	3.46×10^{-7}	-0.31	0.75	1.28	0
<i>RP11-15A1.7</i>	3.65	2.59×10^{-4}	4.26	2.00×10^{-5}	0.78	0.44	0.38	0
Table 3								
<i>KLHDC7A</i>	-4.69	2.77×10^{-6}	-3.53	4.11×10^{-4}	-0.62	0.53	0.91	0
<i>ALS2CR12</i>	4.98	6.25×10^{-7}	2.80	0.005	4.24	2.21×10^{-5}	2.09	0.04
<i>CASP8</i>	-5.97	2.42×10^{-9}	-3.63	2.78×10^{-4}	-4.66	3.20×10^{-6}	2.30	0.13
<i>ATG10</i>	-3.00	0.003	-5.83	5.60×10^{-9}	-2.65	0.008	1.27	0
<i>ATP6AP1L</i>	-2.40	0.02	-4.24	2.20×10^{-5}	-1.87	0.06	0.51	0
<i>L3MBTL3</i>	-5.42	5.89×10^{-8}	-2.38	0.02	-4.13	3.65×10^{-5}	3.13	0.36
<i>RMND1</i>	3.14	0.002	2.76	0.006	2.41	0.02	0.18	0
<i>SNX32</i>	2.41	0.02	3.80	1.45×10^{-4}	1.78	0.08	0.27	0
<i>RCCD1</i>	-5.58	2.36×10^{-8}	-4.08	4.56×10^{-5}	-2.21	0.03	0.81	0
<i>STXBP4</i>	4.77	1.85×10^{-6}	4.01	6.05×10^{-5}	2.46	0.01	0.28	0
<i>ZNF404</i>	4.76	1.96×10^{-6}	5.28	1.28×10^{-7}	2.28	0.02	0.06	0
Table 4								
<i>UQCRH</i>	-3.13	0.002	-2.14	0.03	-1.19	0.23	0.32	0
<i>LMO4</i>	-2.42	0.02	-2.53	0.01	-1.42	0.16	0.002	0
<i>DNAJC27-AS1</i>	3.41	6.47×10^{-4}	1.37	0.17	1.77	0.08	1.12	0
<i>KLHL5</i>	2.34	0.02	1.96	0.05	1.59	0.11	0.09	0
<i>AC008391.1</i>	-2.84	0.004	-3.00	0.003	-0.36	0.72	0.27	0
<i>HCG14</i>	-2.65	0.008	-2.54	0.01	0.02	0.99	0.37	0
<i>TRNAI2</i>	-2.26	0.02	-2.46	0.01	-1.67	0.09	0.02	0

<i>MTHFD1L</i>	2.26	0.02	2.81	0.005	1.58	0.11	0.02	0
<i>PVT1</i>	2.12	0.03	2.73	0.006	1.82	0.07	0.06	0
<i>RP11-123K19.1</i>	-3.80	1.42×10^{-4}	-1.49	0.14	-1.44	0.15	1.37	0
<i>RP11-57H14.3</i>	3.54	3.98×10^{-4}	1.50	0.13	0.09	0.93	1.31	0
<i>RP11-500G22.2</i>	3.09	0.002	3.15	0.002	1.01	0.31	0.10	0
<i>PTDSS2</i>	-1.69	0.09	-2.98	0.003	-1.33	0.18	0.25	0
<i>AP006621.5</i>	2.80	0.005	3.13	0.002	1.26	0.21	0.03	0
<i>PIDD1</i>	1.61	0.11	3.70	2.16×10^{-4}	2.42	0.02	0.82	0
<i>MRPL23-AS1</i>	-2.29	0.02	-2.04	0.04	-2.89	0.004	0.48	0
<i>PACSI</i>	-1.40	0.16	-3.53	4.19×10^{-4}	-1.09	0.27	0.81	0
<i>RP11-860B13.1</i>	2.86	0.004	2.15	0.03	0.66	0.51	0.26	0
<i>KLF5</i>	-2.16	0.03	-2.38	0.017	-3.16	0.002	0.54	0
<i>CTD-2566J3.1</i>	-2.53	0.01	-2.65	0.008	-1.19	0.24	0.02	0
<i>C14orf79</i>	3.60	3.17×10^{-4}	1.89	0.06	2.03	0.04	0.86	0
<i>FES</i>	3.48	4.95×10^{-4}	1.82	0.07	2.34	0.02	0.90	0
<i>BBS2</i>	2.65	0.008	3.08	0.002	0.53	0.59	0.21	0
<i>CRNDE</i>	2.82	0.005	0.50	0.61	2.76	0.006	1.90	0
<i>RP11-482M8.1</i>	2.54	0.01	1.82	0.07	1.37	0.17	0.18	0
<i>GOSR1</i>	2.87	0.004	1.61	0.11	2.22	0.03	0.62	0
<i>ATP6V0A1</i>	2.23	0.03	2.74	0.006	0.94	0.34	0.06	0
<i>RP11-400F19.8</i>	-4.18	2.91×10^{-5}	0.36	0.72	-3.47	5.28×10^{-4}	5.84	0.66
<i>RP11-105N13.4</i>	-2.92	0.004	-2.64	0.008	-2.32	0.02	0.15	0
<i>CBX8</i>	1.82	0.07	3.61	3.04×10^{-4}	2.44	0.01	0.60	0
<i>CTD-2538G9.5</i>	1.61	0.11	3.17	0.002	1.48	0.14	0.39	0
<i>HOMER3</i>	-1.67	0.09	-2.92	0.004	-2.45	0.01	0.41	0
<i>CTD-3216D2.5</i>	1.40	0.16	3.10	0.002	3.18	0.001	0.98	0
<i>TRIOBP</i>	3.77	1.63×10^{-4}	0.55	0.58	0.92	0.36	2.46	0.19
<i>RP5-1039K5.13</i>	2.43	0.02	1.68	0.09	2.67	0.008	0.56	0
<i>CBY1</i>	2.13	0.03	2.60	0.009	2.29	0.02	0.14	0
<i>APOBEC3A</i>	-3.44	5.87×10^{-4}	-1.37	0.17	-2.60	0.009	1.40	0
<i>RP1-85F18.6</i>	1.68	0.09	2.94	0.003	1.48	0.14	0.24	0

Supplementary Table 4. Full list of all index SNPs within the same genomic loci/region of the identified associated genes in Tables 1-4 and their distances with the associated genes

Gene	Index SNP(s) [#]	Distance to the index SNP (kb)
Table 1		
<i>ZSWIM5</i>	rs1707302	829
<i>LRRC3B</i>	rs12493607	3931
	rs653465	591
	rs4973768	705
<i>SPATA18</i>	rs6815814	14,101
<i>UBD</i>	rs9257408	597
<i>KLHDC10</i>	rs4593472	892
<i>MIR31HG</i>	rs1011970	502
<i>RIC8A</i>	rs6597981	588
	rs3817198	1694
<i>B3GNT1</i>	rs3903072	530
<i>RP11-867G23.10</i>	rs3903072	594
<i>RP11-218M22.1</i>	rs12422552	13,641
<i>GALNT16</i>	rs999737	691
<i>PLEKHD1</i>	rs999737	917
<i>MAN2C1</i>	rs2290203	15,851
<i>CTD-2323K18.1</i>	rs2290203	15,619
Table 2		
<i>RP11-439A17.7</i>	rs11249433	442
<i>NUDT17</i>	rs12405132	56
<i>ANKRD34A</i>	rs12405132	169
<i>ALK</i>	rs4577244	295
<i>PRSS46</i>	rs6796502	89
<i>RP11-114I8.4</i>	rs9833888	356
<i>RP11-53O19.1</i>	rs10941679	39
	rs4415084	82

<i>UBLCP1</i>	rs1432679	446
<i>RP11-32D16.1</i>	rs1432679	283
<i>BTN3A2</i>	rs71557345	229
<i>RP11-73O6.3</i>	rs6569648	105
<i>AP006621.6</i>	rs6597981	21
	rs909116	1160
	rs3817198	1127
<i>RPLP2</i>	rs6597981	7
	rs909116	1129
	rs3817198	1096
<i>CTD-3051D23.1</i>	rs10623258	97
<i>RP11-467J12.4</i>	rs12922061	434-1595
	rs17817449	
	rs11075995	
	rs3112612	
	rs3803662	
	rs28539243	
<i>CTD-3032H12.1</i>	rs12922061	290-2385
	rs17817449	
	rs11075995	
	rs3112612	
	rs3803662	
	rs28539243	
<i>LINC00671</i>	rs72826962	190
<i>LRRC37A</i>	rs2532263	118
<i>KANSL1-AS1</i>	rs2532263	18
<i>CRHR1</i>	rs2532263	339
<i>LRRC37A2</i>	rs2532263	336
<i>HAPLN4</i>	rs8170	1977
	rs2363956	1972
	rs4808801	795
	rs2965183	172
<i>RP11-15A1.7</i>	rs3760982	215

Table 3		
<i>KLHDC7A</i>	rs2992756	0.085
<i>ALS2CR12</i>	rs3769821	30
	rs13393577	11075
	rs1830298	inside the gene
<i>CASP8</i>	rs3769821	inside the gene
	rs13393577	11144
<i>ATG10</i>	rs7707921	inside the gene
<i>ATP6AP1L</i>	rs7707921	37
<i>L3MBTL3</i>	rs6569648	208
<i>RMND1</i>	rs9383951	169-2117
	rs9485372	
	rs3757322	
	rs9397437	
	rs851984	
	rs9918437	
	rs2747652	
<i>SNX32</i>	rs3903072	18
	rs75915166	3755
	rs78540526	3707
<i>RCCD1</i>	rs2290203	6
<i>STXBP4</i>	rs6504950	inside the gene
	rs2787486	inside the gene
<i>ZNF404</i>	rs3760982	90
Table 4		
<i>UQCRH</i>	rs1707302	168
<i>LMO4</i>	rs17426269	342
	rs12118297	15
<i>DNAJC27-AS1</i>	rs6725517	65
	rs200648189	455
<i>KLHL5</i>	rs6815814	230
<i>AC008391.1</i>	rs16886113	242
	rs16886181	276

	rs16886397	381
	rs2229882	416
	rs7726354	504
	rs62355902	301
<i>HCG14</i>	rs9257408	61
<i>TRNAI2</i>	rs71557345	307
	rs3757318	491
	rs2046210	525
<i>MTHFD1L</i>	rs9383938	564
	rs11780156	81
	rs13281615	451
<i>PVT1</i>	rs1562430	419
<i>RP11-123K19.1</i>	rs10760444	20
<i>RP11-57H14.3</i>	rs7904519	108
	rs2981582	336
	rs11199914	594
	rs35054928	347
<i>RP11-500G22.2</i>	rs45631563	339
<i>PTDSS2</i>	rs6597981	312
<i>AP006621.5</i>	rs6597981	19
<i>PIDD1</i>	rs6597981	inside the gene
<i>MRPL23-AS1</i>	rs3817198	95
<i>PACSI</i>	rs3903072	255
	rs10771399	221
<i>RP11-860B13.1</i>	rs7297051	241
<i>KLF5</i>	rs6562760	306
	rs2588809	64
<i>CTD-2566J3.1</i>	rs999737	438
<i>C14orf79</i>	rs10623258	240
<i>FES</i>	rs2290203	73
<i>BBS2</i>	rs2432539	80
<i>CRNDE</i>	rs28539243	271
<i>RP11-482M8.1</i>	rs4496150	441

<i>GOSR1</i>	rs146699004	376
<i>ATP6V0A1</i>	rs72826962	162
<i>RP11-400F19.8</i>	rs72826962	122
<i>RP11-105N13.4</i>	rs2532263	359
<i>CBX8</i>	rs745570	6
<i>CTD-2538G9.5</i>	rs8170	432
	rs2363956	437
	rs67397200	444
<i>HOMER3</i>	rs4808801	469
	rs2965183	494
<i>CTD-3216D2.5</i>	rs2284378	281
<i>TRIOBP</i>	rs738321	396
<i>RP5-1039K5.13</i>	rs738321	99
<i>CBY1</i>	rs738321	484
	chr22:39359355	289
<i>APOBEC3A</i>	rs738321	780
	chr22:39359355	0.2
<i>RP1-85F18.6</i>	rs73161324	460
	rs6001930	689

[#] Index SNPs identified in previous GWAS or fine-mapping studies

Supplementary Table 5. In-depth individual level association analyses of predicted expression of 41 identified genes with breast cancer risk in iCOGS and OncoArray datasets identified similar results to those obtained using summary statistics

Gene name	iCOGS dataset individual level analysis (n=84,740)		iCOGS dataset summary statistics analysis (n=89,677)		OncoArray dataset individual level analysis (n=112,133)		OncoArray dataset summary statistics analysis (n=106,776)	
	z-score ^a	p-value ^a	z-score ^a	p-value ^a	z-score ^b	p-value ^b	z-score ^b	p-value ^b
Table 1								
<i>ZSWIM5</i>	3.86	1.12×10^{-4}	4.32	1.57×10^{-5}	3.50	4.73×10^{-4}	2.98	0.003
<i>LRRC3B</i>	-4.76	1.95×10^{-6}	-4.89	1.02×10^{-6}	-7.44	1.04×10^{-13}	-7.48	7.19×10^{-14}
<i>SPATA18</i>	-2.02	0.04	-2.59	0.01	-2.89	3.90×10^{-3}	-3.09	0.002
<i>KLHDC10</i>	3.53	4.12×10^{-4}	4.39	1.16×10^{-5}	2.39	0.02	2.15	0.03
<i>MIR31HG</i>	-2.87	4.07×10^{-3}	-2.90	0.004	-4.99	6.11×10^{-7}	-4.35	1.35×10^{-5}
<i>RIC8A</i>	-3.11	1.86×10^{-3}	-3.12	0.002	-4.15	3.26×10^{-5}	-3.28	0.001
<i>B3GNT1</i>	-3.68	2.35×10^{-4}	-5.00	5.83×10^{-7}	-3.18	1.49×10^{-3}	-2.70	0.007
<i>RP11-218M22.1</i>	2.82	4.82×10^{-3}	3.33	8.82×10^{-4}	3.58	3.47×10^{-4}	3.84	1.22×10^{-4}
<i>GALNT16</i>	-5.07	3.93×10^{-7}	-6.17	6.82×10^{-10}	-4.70	2.62×10^{-6}	-4.45	8.74×10^{-6}
<i>PLEKHD1</i>	2.92	3.50×10^{-3}	3.96	7.43×10^{-5}	5.73	1.01×10^{-8}	5.21	1.85×10^{-7}
<i>MAN2C1</i>	-3.24	1.19×10^{-3}	-3.49	4.88×10^{-4}	-3.69	2.24×10^{-4}	-4.08	4.47×10^{-5}
<i>CTD-2323K18.1</i>	-2.91	3.56×10^{-3}	-2.62	0.009	-3.63	2.88×10^{-4}	-3.69	2.23×10^{-4}
Table 2								
<i>RP11-439A17.7</i>	-3.37	7.61×10^{-4}	-3.39	6.90×10^{-4}	-3.51	4.50×10^{-4}	-4.35	1.37×10^{-5}
<i>ALK</i>	3.27	1.06×10^{-3}	3.23	0.001	4.51	6.62×10^{-6}	3.84	1.23×10^{-4}
<i>PRSS46</i>	-3.22	1.26×10^{-3}	-3.51	4.41×10^{-4}	-5.00	5.80×10^{-7}	-4.33	1.51×10^{-5}
<i>RP11-114I8.4</i>	-3.22	1.28×10^{-3}	-3.15	0.002	-3.77	1.65×10^{-4}	-4.20	2.66×10^{-5}
<i>UBLCP1</i>	2.17	0.03	3.12	0.002	5.10	3.44×10^{-7}	4.72	2.34×10^{-6}
<i>RP11-32D16.1</i>	-2.68	7.31×10^{-3}	-3.66	2.51×10^{-4}	-4.63	3.63×10^{-6}	-3.75	1.75×10^{-4}
<i>BTN3A2</i>	1.51	0.13	2.74	0.006	3.65	2.65×10^{-4}	3.16	0.002
<i>RP11-73O6.3</i>	-1.62	0.11	-2.24	0.03	-5.72	1.08×10^{-8}	-5.34	9.31×10^{-8}
<i>AP006621.6</i>	4.29	1.82×10^{-5}	3.92	8.75×10^{-5}	3.45	5.58×10^{-4}	3.58	3.40×10^{-4}

<i>RPLP2</i>	2.93	3.44×10^{-3}	2.77	0.006	3.39	6.92×10^{-4}	3.43	5.93×10^{-4}
<i>CTD-3051D23.1</i>	-2.83	4.62×10^{-3}	-3.36	7.85×10^{-4}	-2.64	8.39×10^{-3}	-2.60	0.009
<i>RP11-467J12.4</i>	4.78	1.71×10^{-6}	5.41	6.28×10^{-8}	5.63	1.83×10^{-8}	5.75	8.73×10^{-9}
<i>CTD-3032H12.1</i>	3.79	1.50×10^{-4}	2.95	0.003	3.33	8.60×10^{-4}	2.93	0.003
<i>LRRC37A</i>	-3.07	2.11×10^{-3}	-3.08	0.002	-3.75	1.77×10^{-4}	-4.13	3.56×10^{-5}
<i>KANSL1-AS1</i>	-3.12	1.83×10^{-3}	-3.17	0.002	-3.53	4.10×10^{-4}	-3.83	1.28×10^{-4}
<i>CRHR1</i>	-2.67	7.59×10^{-3}	-2.81	0.005	-3.35	7.94×10^{-4}	-3.58	3.39×10^{-4}
<i>HAPLN4</i>	-4.73	2.26×10^{-6}	-5.10	3.46×10^{-7}	-5.87	4.44×10^{-9}	-5.49	4.01×10^{-8}
<i>RP11-15A1.7</i>	3.57	3.54×10^{-4}	4.26	2.00×10^{-5}	4.71	2.45×10^{-6}	3.65	2.59×10^{-4}
Table 3								
<i>KLHDC7A</i>	-2.87	4.06×10^{-3}	-3.53	4.11×10^{-4}	-4.51	6.54×10^{-6}	-4.69	2.77×10^{-6}
<i>ALS2CRI2</i>	2.47	0.01	2.80	0.005	5.09	3.53×10^{-7}	4.98	6.25×10^{-7}
<i>CASP8</i>	-3.72	2.03×10^{-4}	-3.63	2.78×10^{-4}	-5.85	4.98×10^{-9}	-5.97	2.42×10^{-9}
<i>ATG10</i>	-4.55	5.28×10^{-6}	-5.83	5.60×10^{-9}	-4.04	5.44×10^{-5}	-3.00	0.003
<i>ATP6AP1L</i>	-3.33	8.80×10^{-4}	-4.24	2.20×10^{-5}	-3.72	2.02×10^{-4}	-2.40	0.02
<i>L3MBTL3</i>	-1.77	0.08	-2.38	0.02	-5.77	8.06×10^{-9}	-5.42	5.89×10^{-8}
<i>RMND1</i>	2.44	0.01	2.76	0.006	3.64	2.68×10^{-4}	3.14	0.002
<i>SNX32</i>	3.56	3.70×10^{-4}	3.80	1.45×10^{-4}	2.99	2.76×10^{-3}	2.41	0.02
<i>RCCD1</i>	-3.49	4.92×10^{-4}	-4.08	4.56×10^{-5}	-5.76	8.26×10^{-9}	-5.58	2.36×10^{-8}
<i>STXBP4</i>	3.53	4.22×10^{-4}	4.01	6.05×10^{-5}	5.26	1.42×10^{-7}	4.77	1.85×10^{-6}
<i>ZNF404</i>	4.76	1.91×10^{-6}	5.28	1.28×10^{-7}	5.97	2.44×10^{-9}	4.76	1.96×10^{-6}

^a adjusted for study, the first eight principal components, and a principal component derived specifically for the study LMBC (set to zero for all other studies).

^b adjusted for country and the first ten principal components.

Supplementary Table 6. INQUISIT scores of the identified genes showing a significant association with breast cancer risk in the TWAS ($p \leq 5.82 \times 10^{-6}$)

Gene	Distal	Promoter	GTEEx eQTL
From Table 1			
<i>ZSWIM5</i>	none	none	
<i>LRRC3B</i>	none	none	
<i>SPATA18</i>	none	none	
<i>UBD</i>	0.05	none	
<i>KLHDC10</i>	none	none	
<i>MIR31HG</i>	none	none	
<i>RIC8A</i>	none	none	
<i>B3GNT1</i>	none	none	
<i>RP11-867G23.10</i>	none	none	
<i>RP11-218M22.1</i>	none	none	
<i>GALNT16</i>	none	none	
<i>PLEKHD1</i>	none	none	
<i>MAN2C1</i>	none	none	
<i>CTD-2323K18.1</i>	none	none	
From Table 2			
<i>RP11-439A17.7</i>	none	none	yes
<i>NUDT17</i>	3	none	
<i>ANKRD34A</i>	1	none	
<i>ALK</i>	none	none	
<i>PRSS46</i>	none	none	
<i>RP11-114I8.4</i>	none	none	
<i>RP11-53O19.1</i>	none	none	
<i>UBLCP1</i>	none	none	
<i>RP11-32D16.1</i>	none	none	
<i>BTN3A2</i>	none	none	yes
<i>RP11-73O6.3</i>	none	none	
<i>AP006621.6</i>	none	none	yes

<i>RPLP2</i>	1	none	
<i>CTD-3051D23.1</i>	none	none	
<i>RP11-467J12.4</i>	none	none	
<i>CTD-3032H12.1</i>	none	none	
<i>LINC00671</i>	none	none	
<i>LRRC37A2</i>	1	none	
<i>LRRC37A</i>	1	none	
<i>KANSL1-AS1</i>	3	none	
<i>CRHR1</i>	1	none	
<i>HAPLN4</i>	1	none	
<i>RP11-15A1.7</i>	None	none	
From Table 3			
<i>KLHDC7A</i>	none	3	
<i>ALS2CR12</i>	1	none	
<i>CASP8</i>	3	none	
<i>ATG10</i>	3	4	
<i>ATP6AP1L</i>	0.1	none	
<i>L3MBTL3</i>	2	2	
<i>RMND1</i>	4	none	
<i>SNX32</i>	2	none	
<i>RCCD1</i>	5	none	
<i>STXBP4</i>	1	none	
<i>ZNF404</i>	2	none	

Supplementary Table 7. Canonical pathways, diseases and bio functions, and networks associated with identified breast cancer associated genes, and highly co-expressed protein-coding genes of the identified novel susceptibility long non-coding RNAs

Gene(s)	Top canonical pathways	Related diseases and disorders	Molecular and Cellular Functions	Top networks	List of highly co-expressed genes for each long non-coding RNA
Protein-coding genes with Bonferroni corrected significant associations	Granzyme B Signaling ($p=0.024$); Inflammasome pathway ($p=0.030$); Tumoricidal Function of Hepatic Natural Killer Cells ($p=0.036$); Cytotoxic T Lymphocyte-mediated Apoptosis of Target Cells ($p=0.046$)	Cancer; Developmental Disorder; Hematological Disease; Hereditary Disorder; Immunological Disease	Cell Death and Survival; Cell-To-Cell Signaling and Interaction; Cellular Compromise; Cell Cycle; Cellular Morphology	Cell Death and Survival, Cellular Compromise, Nervous System Development and Function; Cancer, Dermatological Diseases and Conditions, Organismal Injury and Abnormalities; Cardiovascular System Development and Function, Cell Cycle, Cellular Development; Cellular Assembly and Organization, DNA Replication, Recombination, and Repair, Cell Cycle; Developmental Disorder, Hereditary Disorder, Ophthalmic Disease	NA
<i>MIR31HG</i>	BER pathway ($p=7.56 \times 10^{-3}$); Dermatan Sulfate Biosynthesis (Late Stages) ($p=0.026$); Chondroitin Sulfate Biosynthesis (Late Stages) ($p=0.028$); Ephrin A Signaling ($p=0.030$); Heparan Sulfate Biosynthesis (Late Stages)	Cardiovascular Disease; Connective Tissue Disorders; Dermatological Diseases and Conditions; Developmental Disorder;	Cell-To-Cell Signaling and Interaction; Cellular Assembly and Organization; Cellular Movement; Gene Expression; Molecular	Cancer, Organismal Injury and Abnormalities, Reproductive System Disease; Cell Cycle, Connective Tissue Disorders, Dermatological Diseases and Conditions; Cardiovascular Disease, Cellular Development, Organismal Injury and Abnormalities; Cancer,	<i>STMN4, ROCK1, APOL2, PRSS35, RPP38, RPUSD3, HS3ST6, LRR1, DIRC1, KLHL38, POLE, TREX2, CACNA1H, AC078883.4, RP5-826L7.1, MYLKP1, TSSK1A, MTHFD1P1, RP11-527F13.1, RP11-32B5.1, PRKCQ-AS1, RP11-834C11.3, CTD-2127H9.1, RP11-454K7.1, CTD-2561B21.10</i>

	($p=0.030$)	Hereditary Disorder	Transport	Gastrointestinal Disease, Organismal Injury and Abnormalities; Hereditary Disorder, Neurological Disease, Organismal Injury and Abnormalities	
<i>RP11-218M22.1</i>	Netrin Signaling ($p=0.024$); ATM Signaling ($p=0.037$); Role of BRCA1 in DNA Damage Response ($p=0.048$)	Cancer; Dermatological Diseases and Conditions; Developmental Disorder; Hereditary Disorder; Neurological Disease	Cell Cycle; DNA Replication, Recombination, and Repair; Cell Death and Survival; Cell Morphology; Cellular Assembly and Organization	Cell Cycle, Cellular Development, Cellular Growth and Proliferation; Cancer, Cell Death and Survival, Organismal Injury and Abnormalities; Developmental Disorder, Hereditary Disorder, Organismal Injury and Abnormalities	<i>FFAR2,PKIB,TP53BP2,LSM14B,NSA2,SYAP1,ZNF738,MAGEF1,FOX12,DCC,NCR1,XRCC2,BLM,RP11-94I2.1,LINC00160,AC092664.1,RP11-83M16.2,CTD-2325P2.4,CTD-3099C6.7</i>
<i>CTD-2323K18.1</i>	D-glucuronate Degradation I ($p=3.31 \times 10^{-3}$); Methylglyoxal Degradation III ($p=0.012$); Mevalonate Pathway I ($p=0.013$); Superpathway of Geranylgeranyldiphosphate Biosynthesis I (via Mevalonate) ($p=0.018$); Tryptophan Degradation X (Mammalian, via Tryptamine) ($p=0.020$);	Cancer; Cardiovascular Disease; Dermatological Diseases and Conditions; Endocrine System Disorders; Hereditary Disorder	DNA Replication, Recombination, and Repair; Post-Translational Modification; Carbohydrate Metabolism; Cell Morphology; Cellular Assembly and Organization	Cellular Assembly and Organization, Hereditary Disorder, Organismal Injury and Abnormalities; Cellular Development, Cellular Growth and Proliferation, Cell Death and Survival; Cell Morphology, Cellular Function and Maintenance, Hematological System Development and Function; Cell Cycle, Cell Morphology, Organ Morphology	<i>VCAN,UBE2T,SUV39H1,MVK,AKR1A1,ZC3H13,MCM8,CASD1,CBLN2,DTL,DGKQ,RPL7A,CCDC74B,CTRC,RHEBL1,SNUPN,PKIA,KIF24,BMP2,MUC19,LINC00612,RP11-157J24.1,LINC00035,RP11-460N20.4,FTH1P1,HNRNPA1P27,FAM203A,RP5-903G2.2,RP11-532F12.5,RP11-340F14.5,RP11-120M18.2,RP11-168F9.2</i>
<i>RP11-439A17.7</i>	Tetrahydrobiopterin Biosynthesis I ($p=2.21 \times 10^{-3}$); Tetrahydrobiopterin	Developmental Disorder; Hereditary Disorder;	Cell Signaling; DNA Replication, Recombination, and Repair;	Cell Morphology, Gastrointestinal Disease, Organismal Injury and Abnormalities; Cellular	<i>TBC1D23,SPR,GNA13,TRMT5,BAIAP2L2,GATA5,GUCY2D,NIPA2,CEP170,ADAMTS16,GTF2H2C,CEND1,IFITM1,SRRM2-AS1,</i>

	Biosynthesis II ($p=2.21 \times 10^{-3}$); Relaxin Signaling ($p=4.13 \times 10^{-3}$); Synaptic Long Term Depression ($p=4.38 \times 10^{-3}$); Endothelin-1 Signaling ($p=6.60 \times 10^{-3}$)	Metabolic Disease; Neurological Disease; Ophthalmic Disease	Nucleic Acid Metabolism; Small Molecule Biochemistry; Cell Morphology	Development, Reproductive System Development and Function, Cell Cycle; Organ Morphology, Reproductive System Development and Function, Connective Tissue Disorders	<i>AC091167.3, RP11-30P6.1, RP5-956O18.3, RP11-137H2.4, COL6A4P1, RP5-836N10.1, VN1R20P, RP11-381E24.1, TET2-AS1, RP11-361I14.2, RP11-732A19.9, CTD-2329K10.1, LA16c-385E7.1, RN7SL15P, SH3GL1P2, MIR3942, RP11-820I16.1, HMGB2P1, RP11-479O17.10</i>
<i>RP11-114I8.4</i>	ErbB2-ErbB3 Signaling ($p=0.043$); ErbB4 Signaling ($p=0.045$)	Dermatological Diseases and Conditions; Developmental Disorder; Hereditary Disorder; Metabolic Disease; Organismal Injury and Abnormalities	Cellular Function and Maintenance; Molecular Transport; Cell Morphology; Gene Expression; Protein Trafficking	Cell Morphology, Cellular Compromise, Cellular Function and Maintenance; Lipid Metabolism, Small Molecule Biochemistry, Dermatological Diseases and Conditions; Hereditary Disorder, Nephrosis, Ophthalmic Disease; Molecular Transport, Cellular Assembly and Organization, Cell Morphology; Cellular Assembly and Organization, Cellular Function and Maintenance, Cell Signaling	<i>ARHGAP31, PEX3, DPP8, CPNE3, KDELR3, A4GNT, RIBC2, NCBP1, SLC25A26, ANKAR, CERS3, CCDC28B, PRORS1P, NRG4, FAM26F, ZNF789, NFKBIL1, Y_RNA, RP1-13D10.2, LINC00205, AC002117.1, RP13-216E22.4, MED4-AS1, ZRANB2-AS1, AD000090.2, RP11-206L10.9, RP11-353N4.2, RP11-499P20.2, TMEM161B-AS1, AC008592.4, RP11-15A1.2, CTD-2639E6.9</i>
<i>RP11-53O19.1</i>	Inosine-5'-phosphate Biosynthesis II ($p=5.44 \times 10^{-3}$); Retinoate Biosynthesis II ($p=7.25 \times 10^{-3}$); Purine Nucleotides De Novo Biosynthesis II ($p=0.020$); Cleavage and	Dermatological Diseases and Conditions; Developmental Disorder; Hereditary Disorder; Neurological	Cell Cycle; Cell Morphology; Cellular Assembly and Organization; Cellular Function and Maintenance; Nucleic Acid	Cell Cycle, Cell-To-Cell Signaling and Interaction, Cellular Growth and Proliferation; Cell Death and Survival, Neurological Disease, Organismal Injury and Abnormalities; Cancer, Cell Death and Survival, Cell-	<i>MCM10, RRP15, EPN2, MTMR3, CPSF6, RBBP5, FBXO30, PAICS, CAPN11, RNF144B, HAUS2, KIF21A, FAM222A, CTDSPL, HNRNPU, TOMM70A, RIBC1, RRP1B, RBP7, FUBP1, S100Z, C17orf66, NUDT4, DSG4, MED16, OR10A6, GCNT4, TMEM139, ZNF320, C11orf72, CXorf38, ZNF566, ZNF197, TNFRSF18, MAGI2,</i>

	Polyadenylation of Pre-mRNA ($p=0.022$); Epithelial Adherens Junction Signaling ($p=0.028$)	Disease; Ophthalmic Disease	Metabolism	To-Cell Signaling and Interaction; Cardiovascular Disease, Cell Death and Survival, Cell Morphology; Embryonic Development, Organismal Development, Tissue Morphology	VWC2, GLRA4, AC104472.1, UBE2Q2P2, AC073621.2, AC073850.6, FGF12-AS2, AC073342.12, RP11-557H15.4, RP11-640M9.1, RN7SL331P, AP000322.53, RP11-51J9.4, RP11-1055B8.4, RP11-135L13.4, RP11-64C12.8
RP11-32D16.1	AMPK Signaling ($p=1.96 \times 10^{-4}$); Tyrosine Degradation I ($p=2.20 \times 10^{-4}$); Phenylalanine Degradation IV (Mammalian, via Side Chain) ($p=1.95 \times 10^{-3}$); LPS/IL-1 Mediated Inhibition of RXR Function ($p=3.11 \times 10^{-3}$); Valine Degradation I ($p=3.23 \times 10^{-3}$)	Metabolic Disease; Endocrine System Disorders; Gastrointestinal Disease; Hepatic System Disease; Organismal Injury and Abnormalities	Lipid Metabolism; Small Molecule Biochemistry; Energy Production; Molecular Transport; Carbohydrate Metabolism	Lipid Metabolism, Molecular Transport, Small Molecule Biochemistry; Cell Signaling, Nucleic Acid Metabolism, Small Molecule Biochemistry; Cell Cycle, Gene Expression, Organ Morphology; Carbohydrate Metabolism, Molecular Transport, Small Molecule Biochemistry; Skeletal and Muscular Disorders, Cell Morphology, Organ Development	TNMD, CX3CL1, DAPK2, HEBP2, TRAF3IP2, GYG2, LIPE, SEPHS1, FMO2, APMAP, SLC6A2, FAH, ETFB, SH3D19, CPT1A, TMED5, ITGB1BP1, KCNIP2, CAT, WIPF3, ACVR1C, PCK1, ARHGEF6, SLC7A10, INPP5K, KL, TCN1, ADAM19, SDS, CD36, PPP1R1A, TTLL4, SLC19A3, RTN1, ETFA, ADAMTS18, SELENBP1, CDKN2B, TM7SF2, FLI1, PDE3B, PLOD2, RWDD2B, NAA11, RPUSD3, C2CD2, HPD, PFKFB1, FBXO27, CCDC51, CTSB, SUN1, NIPSNAP3B, AQP7, ZNF219, GPT2, PLIN1, ACAA2, GPD1, PLIN4, ANGPTL4, GLYCTK, GSG1L, MRAP, FABP4, PFKFB3, MUC7, AQPEP, DCTN2, FOXG1, CIDEA, PLA2G16, BOK, RPLP2, PNPLA2, COX14, ADIPOQ, ABAT, TUSC5, NAT8L, CIDEC, DHRS4L2, MAOA, MAN2A2, VKORC1L1, KCNKG, NUDT16, GJC2, LINC00222, PCDHA7, AC022007.5, MIR135A1, ZNF259P1, KCNIP2-AS1, AC022596.6, ADIPOQ-AS1, RP11-445L13__B.3, RP1-28O10.1, AC008738.1, VN1R108P, RP11-573D15.1, RP5-1172A22.1,

					<i>RP1-293L6.1, LINC00263, TRHDE-AS1, AC159540.2, VWFP1, GLYCTK-AS1, CEBPA, RP11-768F21.1, CTD-2589H19.4, RP13-884E18.2, RP11-1101K5.1, PAICSP4, AP006621.8, RP11-317P15.5, RP11-663N22.1</i>
<i>RP11-7306.3</i>	Pentose Phosphate Pathway (Oxidative Branch) ($p=5.88 \times 10^{-3}$); Selenocysteine Biosynthesis II (Archaea and Eukaryotes) ($p=8.81 \times 10^{-3}$); GDP-mannose Biosynthesis ($p=8.81 \times 10^{-3}$); p53 Signaling ($p=9.13 \times 10^{-3}$); Tryptophan Degradation to 2-amino-3-carboxymuconate Semialdehyde ($p=0.010$)	Cardiovascular Disease; Connective Tissue Disorders; Developmental Disorder; Hematological Disease; Hereditary Disorder	Cell Death and Survival; Carbohydrate Metabolism; Cell Cycle; Cell Morphology; Cell-To-Cell Signaling and Interaction	Cellular Development, Cellular Growth and Proliferation, Reproductive System Development and Function; Cell-mediated Immune Response, Cellular Development, Cellular Function and Maintenance	<i>HIVEP2, LAMA3, SEPHS1, ARHGAP28, DHX32, GGA1, EIF3D, PSMD7, GPI, URGCP, XPNPEP1, PPP1R9B, SMURF2, DDX25, NR5A1, TP53BP2, PIK3R1, SYB U, CDYL, NCAM2, DHRS4, G6PD, ZER1, SHANK1, HAAO, SH3TC2, PACS1, L3MBTL3, LINC00162, RP11-536C5.7, RP11-213G2.2, AC002401.1, MTX1P1, RP11-247I13.11, LINC00461, RP11-281P23.2, RP11-10A14.4, RP11-578O24.2, FTL1P14, AC005702.1, RP11-138E2.1, RP11-216P16.2</i>
<i>AP006621.6</i>	Primary Immunodeficiency Signaling ($p=1.40 \times 10^{-3}$); Acetate Conversion to Acetyl-CoA ($p=5.04 \times 10^{-3}$); T Cell Receptor Signaling ($p=6.49 \times 10^{-3}$); G12/13 Signaling ($p=9.50 \times 10^{-3}$); Tec Kinase Signaling ($p=0.016$)	Cancer; Cardiovascular Disease; Connective Tissue Disorders; Dermatological Diseases and Conditions; Developmental Disorder	Cellular Function and Maintenance; Cell Death and Survival; Cell Morphology; Cell-To-Cell Signaling and Interaction; Cellular Development	Humoral Immune Response, Protein Synthesis, Hematological System Development and Function; Cellular Compromise, Cell Cycle, Cellular Assembly and Organization; Cell Cycle, Hereditary Disorder, Neurological Disease; Embryonic Development, Organismal Development, Tissue Development	<i>BTK, HERPUD1, PRDM1, KCNAB2, ZFAND6, DERL3, SRRD, ACSS3, LAX1, ZBP1, RGS13, TEC, DUOXA2, RPL11, UGCG, LPCAT1, GF11B, RNF187, SHCBP1, AP006621.1, PIDD, NUGGC, IGHA1, FIS1, IGLC6, RPL32P1, RP11-162O12.2, LINC00568, GS1-124K5.11, LINC00582, AC007285.7, AC005162.5, RP11-181G12.4, LINC01010, RPS20P21, H2AFJ, RP11-510M2.2, RP11-1084E5.1, AP006621.5, AC091171.1, RP11-554D14.4, RP11-421I10.1,</i>

					<i>RP11-61A14.3,RP11-325K4.3,RP11-174G6.5,RP11-849F2.8,RP4-713A8.1,STAG3L5P-PVRIG2P-PILRB</i>
<i>CTD-3051D23.1</i>	Granulocyte Adhesion and Diapedesis ($p=0.039$); Agranulocyte Adhesion and Diapedesis ($p=0.043$); IL-22 Signaling ($p=0.045$); Role of JAK family kinases in IL-6-type Cytokine Signaling ($p=0.046$); B Cell Development ($p=0.050$)	Cancer; Cardiovascular Disease; Developmental Disorder; Endocrine System Disorders; Hematological Disease	Cellular Development; Cell Morphology; Cellular Growth and Proliferation; Lipid Metabolism; Molecular Transport	Cell Cycle, Cell Death and Survival, Cellular Compromise; Cellular Development, Cellular Growth and Proliferation, Hematological System Development and Function; Cellular Assembly and Organization, Cellular Function and Maintenance, Tissue Morphology; Connective Tissue Disorders, Organismal Injury and Abnormalities, Reproductive System Development and Function; Infectious Diseases, Cancer, Organismal Injury and Abnormalities	<i>RC3H2,TDRD3,GPATCH2L,IL5RA,EZR,TFIP11,MIB1,CD79A,EPB41L5,SLC1A4,STAT5A,CASZ1,VSTM2L,SPIRE1,KLF4,NAA30,TBX19,RNF145,AZGP1,SCIMP,PPP1R32,VANGL2,LEO1,LMAN2,SERPINA6,NAT1,NDUFA11,SOX11,TSSK6,PCED1B,CRELD2,LCN12,CCDC157,NKAPL,TMEM240,IGLV2-23,IGLV3-19,IGLC3,CCL27,HLA-FAS1,RP11-216N14.5,AC068587.2,RP11-557J10.4,CTD-2240H23.2,KB-1460A1.5,LA16c-431H6.6,RP11-283I3.6,RP11-597M12.1,RP11-694I15.7,RP4-734G22.3</i>
<i>RP11-467J12.4</i>	Glycerol-3-phosphate Shuttle ($p=2.32 \times 10^{-3}$); Glycerol Degradation I ($p=5.78 \times 10^{-3}$)	Cancer; Organismal Injury and Abnormalities; Reproductive System Disease; Cardiovascular Disease; Developmental Disorder	Cell-To-Cell Signaling and Interaction; Cellular Assembly and Organization; Cellular Growth and Proliferation; Cell Morphology; Cellular Development	Cardiovascular System Development and Function, Cellular Development, Cellular Function and Maintenance; Cell Morphology, Connective Tissue Development and Function, Tissue Morphology	<i>PREX2,CD82,MARCH2,INTS10,TPD52L1,MORN1,HSDL2,CHST8,FGD3,IL17B,BTBD3,PRSS23,ANKFN1,ZKSCAN2,SUSD3,ALDH16A1,HDAC11,GPD1,EMR1,CTDNEP1,SRGAP3,SFTA2,AC074391.1,AL021068.1,RP11-553A21.3,RP4-669P10.16,LL0XNC01-237H1.3,DPY19L2P4,AC007750.5,KRT8P36,RP11-368I23.2,RP11-16P20.3,FMRI-AS1</i>

<i>CTD-3032H12.1</i>	ERK/MAPK Signaling ($p=2.00 \times 10^{-4}$); FLT3 Signaling in Hematopoietic Progenitor Cells ($p=1.14 \times 10^{-3}$); Acute Myeloid Leukemia Signaling ($p=1.50 \times 10^{-3}$); - Adrenergic Signaling ($p=1.92 \times 10^{-3}$); Corticotropin Releasing Hormone Signaling ($p=3.69 \times 10^{-3}$)	Inflammatory Response; Cancer; Organismal Injury and Abnormalities; Auditory Disease; Cardiovascular Disease	Cell-To-Cell Signaling and Interaction; Cell Death and Survival; Cell Cycle; Cell Morphology; Cellular Function and Maintenance	Embryonic Development, Organ Development, Organismal Development; Cell Morphology, Cell Death and Survival, Cellular Development; Lipid Metabolism, Molecular Transport, Small Molecule Biochemistry; Post-Translational Modification, Cell Morphology, Cellular Function and Maintenance; Dermatological Diseases and Conditions, Organismal Injury and Abnormalities, Hair and Skin Development and Function	<i>MBTD1,AGA,CHI3L2,FNDC3B,GRHL2,CRLS1,ESR1,DHRS7,HTATSF1,DNAJB6,GATA3,ATE1,STC2,MOGS,PDCL3,SLC1A4,NR5A2,IPO13,STAT5A,TRPV5,EPS15L1,YWHAH,RAF1,HRASLS2,GLCE,C1RL,GRTP1,IGLON5,UCK2,TNFRSF21,RNF44,RASSF3,C17orf103,SH3RF2,PLXDC1,ADCY9,C1orf50,MOCS2,FBP1,TAF1D,DEGS2,PLA2G4F,C3orf33,IRX5,IRX3,SETD3,C1orf64,PIP5K1C,EIF4EBP1,HUS1B,ZKSCAN7,CALM1,CAPN8,SPINK9,SNORA40,LINC00277,RP11-223A3.1,ATP8A2P2,AK3P3,RP11-5P18.5,LINC00941,KIFC1,RP11-69L16.4,RP1-93H18.1,RP11-64D22.2,RGAG1,CTB-33O18.1,RP11-676M6.1,POLG2,AC040173.1,RP11-114F10.2,LINC00567,AC009133.20,RP11-106E15.1,CTD-2206N4.4,CTD-3214H19.6,CTD-3099C6.11,CTD-2376I4.2,RP11-383I23.2</i>
<i>KANSL1-AS1</i>	Endoplasmic Reticulum Stress Pathway ($p=0.024$); Tumoricidal Function of Hepatic Natural Killer Cells ($p=0.027$); Cytotoxic T Lymphocyte-mediated Apoptosis of Target Cells ($p=0.035$); TWEAK Signaling ($p=0.038$)	Developmental Disorder; Hereditary Disorder; Neurological Disease; Organismal Injury and Abnormalities; Psychological Disorders	Cell Morphology; Cellular Function and Maintenance; Lipid Metabolism; Molecular Transport; Small Molecule Biochemistry	Gene Expression, Cell Cycle, Lipid Metabolism; Cell Cycle, Reproductive System Development and Function, Embryonic Development; Developmental Disorder, Hereditary Disorder, Ophthalmic Disease; Cell Cycle, Endocrine System Development and Function, Lipid Metabolism	<i>BTN3A1,MRPL43,USP14,KANSL1,EMC6,NLGN4X,TMEM219,CDC42SE2,ZNF230,NEU3,CASP7,TRAPPC2L,ACYP2,LRRC37A,CECR6,FAM227A,NUDT17,EPHB4,CRHR1-IT1,ARHGAP19,LRRC37A4P,MICD,RP4-782L23.1,FAM215B,AC007386.4,LRRC37A2,DECR2,CCDC153,BANF1P1,AC004449.6,CTD-2555A7.3,RP11-259G18.2,RP11-259G18.3,RP11-1055B8.8,DND1P1,</i>

					<i>RP11-798G7.8,RP11-622C24.1</i>
<i>LINC00671</i>	Dolichyl-diphosphooligosaccharide Biosynthesis ($p=0.016$); Hereditary Breast Cancer Signaling ($p=0.017$); Antiproliferative Role of TOB in T Cell Signaling ($p=0.037$); Inhibition of Angiogenesis by TSP1 ($p=0.046$)	Inflammatory Response; Cancer; Cardiovascular Disease; Developmental Disorder; Gastrointestinal Disease	DNA Replication, Recombination, and Repair; Cell-To-Cell Signaling and Interaction; Cellular Function and Maintenance; Cell Cycle; Cellular Development	Cell Death and Survival, Cancer, Organismal Injury and Abnormalities; Cell Morphology, Developmental Disorder, Digestive System Development and Function; Cellular Movement, Nervous System Development and Function, Embryonic Development; Cell Morphology, Cellular Function and Maintenance, Cellular Movement; Organ Morphology, Organismal Development, Organismal Injury and Abnormalities	<i>RHBDF1,NCAPD2,UBE2D1,PALB2,PHF14,EHD1,AMOTL2,CHST10,TXLNG2P,KDELC1,TUFT1,TRIM50,CABYR,N6AMT1,SIGLEC11,C2orf44,TGFBR2,MAP9,ARSK,WEE1,TEF,DPAGT1,C11orf68,HIC1,LINC00670,CEP97,TNFAIP2,KTN1-AS1,ZNF567,Y_RNA,LINC00449,LINC00963,SYNJ2BP,RPL39P5,LRRC37A11P,U3,SMIM13,RP11-61N20.3,RP11-222A11.1,RN7SL165P,RN7SL244P,RP11-463J10.3,RP11-407G23.4,AOC4P,RP11-2117.4,LINC00565,RP11-703I16.1,MIR24-2,CLEC4GP1</i>
<i>RP11-15A1.7</i>	Induction of Apoptosis by HIV1 ($p=1.09 \times 10^{-4}$); Docosaheanoic Acid (DHA) Signaling ($p=1.72 \times 10^{-3}$); Molecular Mechanisms of Cancer ($p=2.33 \times 10^{-3}$); CD27 Signaling in Lymphocytes ($p=2.93 \times 10^{-3}$); Small Cell Lung Cancer Signaling ($p=5.60 \times 10^{-3}$)	Infectious Diseases; Cancer; Cardiovascular Disease; Dermatological Diseases and Conditions; Developmental Disorder	Cellular Compromise; Cellular Assembly and Organization; Cell Morphology; Cell Death and Survival; Cell-To-Cell Signaling and Interaction	Cell Morphology, Cellular Assembly and Organization, Behavior; Cell Morphology, Cellular Function and Maintenance, Cellular Compromise; Cell Signaling, Nucleic Acid Metabolism, Molecular Transport; Cell Death and Survival, Cellular Development, Cellular Growth and Proliferation	<i>SLC25A13,BID,WAPAL,PBX4,RASD1,KLF9,ADCY7,ZNF211,TMEM115,CDKN2D,TULP4,WTAP,ZNF7,ANXA8L1,PAN3,ZNF761,UVSSA,INTS1,FAM102A,BCL2L1,C19orf18,PER1,CDC42EP4,HKR1,GPR1,ANKRD19P,ZNF34,DNM3,ZFP2,ZNF155,FAM71F2,AKR1C7P,RAB1C,RP11-538P18.2,ZBED5-AS1,RP11-301H24.3,RP11-420A23.1,RP11-521B24.5,CHMP4BP1,NT5CP1,RP11-109N23.6,SMIM6,HCCAT3,RP11-727F15.13,RP11-130L8.2,RP11-274B21.9,RP5-1024N4.4,RP11-434H6.7</i>

NA: not available

Supplementary Table 8. Cell line and media information.

Cell Line	Media constituents
MCF10A	DMEM/F12 + 5% Horse Serum + 20ng/mL EGF + 0.5µg/mL Hydrocortisone + 100ng/mL Cholera Toxin + 10 µg/mL Insulin from bovine pancreas + 1% Penicillin-Streptomycin
Bre80-Tert	DMEM/F12 + 5% Horse Serum + 20ng/mL EGF + 0.5µg/mL Hydrocortisone + 100ng/mL Cholera Toxin + 10 µg/mL Insulin from bovine pancreas + 1% Penicillin-Streptomycin
184A1	MEGM + BPE 52ug/mL + HC 500ng/mL + EGF 10ng/ml + I 5ug/ml + transferrin 5ug/mL + cholera toxin 1ng/mL
ZR751	RPMI-1640 + 10% Fetal Bovine Serum + 1% Penicillin-Streptomycin + 10 µg/mL Insulin from bovine pancreas
MCF7	RPMI-1640 + 10% Fetal Bovine Serum + 1% Penicillin-Streptomycin
KPL1	DMEM + 10% Fetal Bovine Serum + 1% Penicillin-Streptomycin
T47D	RPMI-1640 + 10% Fetal Bovine Serum + 1% Penicillin-Streptomycin
SKBR3	DMEM + 10% Fetal Bovine Serum + 1% Penicillin-Streptomycin
BT474	RPMI-1640 + 10% Fetal Bovine Serum + 1% Penicillin-Streptomycin
MDA-MB-453	DMEM/F12 + 20% Fetal Bovine Serum + 1% Penicillin-Streptomycin + 10 µg/mL Insulin from bovine pancreas
MDA-MB-231	DMEM + 10% Fetal Bovine Serum + 1% Penicillin-Streptomycin
MDA-MB-436	DMEM + 10% Fetal Bovine Serum + 1% Penicillin-Streptomycin

BT549	RPMI-1640 + 10% Fetal Bovine Serum + 1% Penicillin-Streptomycin
MDA-MB-157	DMEM + 10% Fetal Bovine Serum + 1% Penicillin-Streptomycin
HCC1937	RPMI-1640 + 10% Fetal Bovine Serum + 1% Penicillin-Streptomycin
HS578T	DMEM + 10% Fetal Bovine Serum + 1% Penicillin-Streptomycin
SUM159PT	RPMI-1640 + 10% Fetal Bovine Serum + 1% Penicillin-Streptomycin
MDA-MB-468	DMEM + 10% Fetal Bovine Serum + 1% Penicillin-Streptomycin

Supplementary Table 9. siRNA sequences.

Name	Sense sequence (5'-3')	Antisense sequence (5'-3')
RMND1-1	CCACGGAUAUGUUGAAGUATT	UACUUCAACAUAUCCGUGGGA
RMND1-2	CAAACCAAAUCUGUUGGGUUCUAAA	UUUAGAACCCAACAGAUUUGGUUUG
KLHDC10-1	CAACCUAUAUGUGUUUGGAGGUUAU	AUAACCUCCAAACACAUAUAGGUUG
KLHDC10-2	GAGAUAUCUGGAAGUUGAAUCUGCA	UGCAGAUUCAACUCCAGAUUAUCUC
ZSWIM5-1	GGGAAAGUGAAAGACUACUCUUUAA	UAAAAGAGUAGUCUUUCACUUUCCC
ZSWIM5-2	CCUCAUUGGCCAUGAGCCAUCUUA	UUAAGAUGGCUCAUGGCCAAUGAGG
UBLCP1-1	GCACCUAAAUCGUGAUAAATT	UUUAUCACGAUUUAGGUGCGC
UBLCP1-2	CAGGAGUAUUCAGUGACCACACUUU	AAAGUGUGGUCACUGAAUACUCCUG
PLEKHD1-1	UCAAGAGAGCUIUUCUGCUUUACUA	UAGUAAAGCAGAAAGCUCUCUUUGA
PLEKHD1-2	AAGAUGCCUUAAGGGUGUAGAACA	UGUUCUACACCCUUAAGGCAUCUUG
ALS2CR12-1	AACUCCACAGGGAGUCCAAGCUAA	UUAGCUUGGAACUCCCUGUGGAGUU
ALS2CR12-2	CAGCAAGGCAAGAAGAGACUAAUAA	UUAUUAGUCUCUUCUUGCCUUGCUG
STXBP4-1	(CCUGGAGGAGACUGUUAUA)dTdT	(UAUAACAGUCUCCUCCAGG)dAdA
STXBP4-2	(GGACCUCAAGCCUCAACAU)dTdT	(AUGUUGAGGCUUGAGGUCC)dAdT
ZNF404-1	UGCGUACCAUCAGGAGACAUGGAAA	UUUCCAUGUCUCCUGAUGGUACGCA
ZNF404-2	GGGAAACGUUUAGAUUAUAUCGACA	UGUCGAUAUAUUCUAAACGUUUC
PIDD-1	GACUGUCCUGACCUCAGAtt	UCUGAGGUCAGGAACAGUCtg
PIDD-2	AGGGCAGAAUCUGCUUUGUCUUCUA	UAGAAGACAAAGCAGAUUCUGCCCU
NRBF2-1	UGUGAAAUGCGCUGCGUAUUU	AUACGCAGCGCAUUUCACAUU
NRBF2-2	CCGGAGGAGGAAGUGGUGAGGUUGU	ACAACCUCACCACUCCUCCUCCGG
NRBF2-3	AGGAAGUGGUGAGGUUGUUGCUCU	AGGAGCAACAACCUCACCACUUCU
ABHD8-1	GAGCAAUCUUAAGCGCUAUGCCAA	UUGGCAUAGCGCUUGAAGAUUGCUC
ABHD8-2	CAUCCUACGGUGUCUCUUUCUGCA	UGCAGAAAGAGACACCGUAGGAAUG
RP11-218M22-R1-1	UGAGCGCAGGAACCAUGGUCUUCU	AUGAAGACCAUGGUUCCUGCGCUA
RP11-218M22-R1-2	CGCAGGAACCAUGGUCUUCUUGCU	AGCAAUGAAGACCAUGGUUCCUGCG
RP11-218M22-R2-1	CCAGUGGGUUUGGAUUAUAAUCCUGA	UCAGGAUUAUAUCCAAACCCACUGG
RP11-218M22-R2-2	CAGACUGCGAGACAAUCUCUCUUA	UAAAGAGAGAUUGUCUCGCAGUCUG
AP006621.6-1	GGGUACCUUACACUGGGCGUCAGAA	UUCUGACGCCAGGUGAAGGUACCC

AP006621.6-2	UCACCUGGGCGUCAGAAGCACUUGA	UCAAGUGCUUCUGACGCCCAGGUGA
RP11-467J12.4-1	CACCAUAUCAUGGUUCCCACUAGCA	UGCUAGUGGGAACCAUGAU AUGGUG
RP11-467J12.4-2	UAUGAGAGUUCCAGUUGCUCCACAA	UUGUGGAGCAACUGGAACUCUCAUA
RP11-15A1.7-1	CACCCUCCUCAUACUUCCGUAGUUU	AAACUACGGAAGUAUGAGGAGGGUG
RP11-15A1.7-2	GGAAUCCACCUAAGUGUCUAUCAAU	AUUGAUAGACACUUAGGUGGAU UCC
CTD-3032H12.1-1	CAAGCUCCCGAGGCGAUCUGCUGUU	AACAGCAGAU CGCCUCGGGAGCUUG
CTD-3032H12.1-2	AGGCCCAAGUCGCAGUUCUCGUGAA	UUCACGAGAACUGCGACUUGGGCCU
B2M-1	CCAGCGUACUCCAAAGAUUTT	AAUCUUUGGAGUACGCUGGTT
B2M-2	GGTTTACTCACGTCATCCATT	TGGATGACGTGAGTAAACCTT
ARHGDIA-1	CCCGUCUAACCAUGAUGCCUUAACA	UGUUAAGGCAUCAUGGUUAGACGGG
ARHGDIA-2	CCUUAACAUGUGGAGUGUACCGUGG	CCACGGUACACUCCACAUGUUAAGG
ZAP70-1	UAACCUCCUCAUAGCUGACAUUGAA	UUCA AUGUCAGCUAUGAGGAGGUUA
ZAP70-2	CCGAAUGCAUCAACUCCGCAAGUU	AACUUGCGGAAGUUGAUGCAUUCGG

Supplementary Table 10. Literature reported link between genes identified in our study that have not been reported from eQTL and/or following functional studies as target genes of risk variants (Tables 1-2) and breast cancer

Gene	Reported link with breast cancer	Study type	Consistency with the direction of effect identified in our study	PMID of literature
Table 1				
<i>ZSWIM5</i>	NA	NA	NA	NA
<i>LRRC3B</i>	inhibits bupivacaine-induced breast cancer cell invasion	<i>In vitro</i>	consistent	29085514
	reduced expression in breast cancer tissues compared with breast fibroma tissues; low gene expression associated with higher tissue grade	Human tissues		24839112
	methyated and/or deleted in ~32% breast carcinoma samples	Human tissues		22321817
<i>SPATA18</i>	downregulated ~ 5-folds in human ductal breast carcinomas compared with normal breast samples	Human tissues	consistent	21300779; 16473279
<i>UBD</i>	inhibits growth in MCF-7 breast carcinoma cells	<i>In vitro</i>	consistent	12170760
	increased expression in breast cancer tissues compared with surrounding tissues; expression correlated with triple-negative breast cancer (TNBC)	Human tissues	inconsistent	26185453
<i>KLHDC10</i>	NA	NA	NA	NA
<i>MIR31HG</i>	down-regulated in TNBC cell lines of basal subtype; heavily methylated in the TNBC cell lines	<i>In vitro</i>	consistent	22289355
	increased expression in breast cancer tissues compared with normal control; expression associated with advanced pathologic stage and tumor size; knockdown decreases breast cancer cell proliferation, induces apoptosis, inhibits migration/invasion and impedes tumorigenesis	<i>In vitro, in vivo, and human tissues</i>	inconsistent	24631686
<i>RIC8A</i>	undergoes a classical double-hit genetic inactivation in a breast	<i>In vitro and in</i>	consistent	19432969

	cancer cell line; loss of expression in a subgroup of aggressive <i>TP53</i> mutant breast cancers	<i>vivo</i>		
<i>B3GNT1</i>	NA	NA	NA	NA
<i>RP11-867G23.10</i>	NA	NA	NA	NA
<i>RP11-218M22.1</i>	NA	NA	NA	NA
<i>GALNT16</i>	NA	NA	NA	NA
<i>PLEKHD1</i>	NA	NA	NA	NA
<i>MAN2C1</i>	NA	NA	NA	NA
<i>CTD-2323K18.1</i>	NA	NA	NA	NA
Table 2				
<i>RP11-439A17.7</i>	NA	NA	NA	NA
<i>NUDT17</i>	NA	NA	NA	NA
<i>ANKRD34A</i>	NA	NA	NA	NA
<i>ALK</i>	overexpressed in 36% of breast cancer patients; gene amplification present in 13.3 % of cases; overexpression associated with aggressive behavior	Human tissues	consistent	26384210
	amplified in a large proportion of Inflammatory Breast Cancers (IBC), a highly aggressive subtype of breast cancer	<i>In vitro</i> and human tissues		22215853
	copy number gain observed in 47.2% of IBC patients; copy number gain associated with poorer recurrence free survival	Human tissues		25803816
<i>PRSS46</i>	NA	NA	NA	NA
<i>RP11-114I8.4</i>	NA	NA	NA	NA
<i>RP11-53O19.1</i>	NA	NA	NA	NA
<i>UBLCP1</i>	NA	NA	NA	NA
<i>RP11-32D16.1</i>	NA	NA	NA	NA
<i>BTN3A2</i>	higher expression associated with improved distant metastasis-free survival in HR-/HER2+ breast cancer	Human tissues	NA	28409241
<i>RP11-73O6.3</i>	NA	NA	NA	NA

<i>AP006621.6</i>	NA	NA	NA	NA
<i>RPLP2</i>	differentially expressed for breast cancer apoptosis (both up- and down-regulation)	<i>In vitro</i>	NA	22133146
<i>CTD-3051D23.1</i>	NA	NA	NA	NA
<i>RP11-467J12.4</i>	NA	NA	NA	NA
<i>CTD-3032H12.1</i>	NA	NA	NA	NA
<i>LINC00671</i>	NA	NA	NA	NA
<i>LRRC37A2</i>	NA	NA	NA	NA
<i>LRRC37A</i>	NA	NA	NA	NA
<i>KANSL1-AS1</i>	NA	NA	NA	NA
<i>CRHR1</i>	encodes a receptor of corticotropin-releasing hormone (CRH), which suppresses TGFβ1-induced Epithelial-Mesenchymal Transition in breast cancer cells	<i>In vitro</i>	consistent	24412750 26138318
<i>HAPLN4</i>	NA	NA	NA	NA
<i>RP11-15A1.7</i>	NA	NA	NA	NA

NA: not available

Supplementary Table 11. Performance of prediction models and association results for breast cancer target genes reported previously in GWAS-identified loci

Chromosome regions	Target genes	Reference	Evidence from original paper for supporting this gene as the target gene	Performance of expression prediction model (R^2) in GTEx/TCGA	Association of predicted expression with breast cancer risk
1p33	<i>NSUN4</i>	¹	eQTL analyses in GTEx, TCGA (tumor tissue) and METABRIC (tumor adjacent normal tissue), prediction by ChIA-PET in MCF7 cells	0.01/0.006	$p=1.95 \times 10^{-4}$ (z: negative)
1p36.22	<i>PEX14</i>	²	eQTL analyses in TCGA (tumor and adjacent normal tissue)	0.02/0	$p=0.002$ (z: positive)
2p23.2	<i>TRMT61B</i>	³	eQTL analyses in TCGA (tumor tissue) and Norwegian normal breast cohort (normal tissue)	0.23/0.33	$p=0.30$
2q33	<i>PPIL3</i> , <i>CASP8</i>	^{3,4}	eQTL analyses in TCGA (tumor tissue); eQTL analyses in TCGA (tumor adjacent normal tissue) and Westra <i>et al.</i> (peripheral blood samples)	0.44/0.59, 0.22/0.30	$p=0.02$ (z: positive), $p=8.51 \times 10^{-16}$ (z: negative)
2q35	<i>IGFBP5</i>	⁵	eQTL analyses in the Norwegian Breast Cancer Study and METABRIC (tumor adjacent normal tissue) (marginal significant associations with levels of one of the tested probes, but not any others)	0.04/0.004	NA
4q24	<i>TET2</i>	⁶	eQTL analyses in TCGA (tumor tissue) and METABRIC (tumor adjacent normal tissue)	0.007/0.02	$p=0.08$
5p12	<i>FGF10</i> , <i>MRPS30</i>	⁷	eQTL analyses in GTEx (normal tissue) and Norwegian Breast Cancer Study (tumor and tumor adjacent normal tissue); eQTL analyses in GTEx (normal tissue), and Norwegian Breast Cancer Study and TCGA (both tumor and tumor adjacent normal tissue)	0.02/0, 0.006/0.16	$p=0.26$, $p=1.43 \times 10^{-25}$ (z: positive)
5p15.33	<i>TERT</i>	⁸	luciferase reporter assays	NA	NA
5q11.2	<i>MAP3K1</i>	⁹⁻¹¹	Chromosome Conformation Capture and luciferase reporter assays etc, while, no	0.06/0	$p=0.32$

			detectable differences in expression were found across genotypes of the index SNPs		
5q14	<i>ATP6AP1L</i>	¹²	eQTL analyses in TCGA (tumor tissue)	0.63/0.32	$p=6.32 \times 10^{-7}$ (z: negative)
6p24.3	<i>GCNT2</i>	¹³	eQTL analyses in TCGA (tumor tissue)	NA	NA
6q25	<i>ESR1</i> , <i>RMND1</i> , <i>CCDC170</i> , <i>AKAP12</i>	^{14,15}	eQTL analyses in TCGA (tumor tissue) and METABRIC (tumor and tumor adjacent normal tissue); eQTL analyses in TCGA (tumor tissue); eQTL analyses in TCGA (tumor tissue) and GTEx (normal tissue); eQTL analyses in TCGA (tumor tissue)	NA, 0.13/0.02, 0.02/NA, NA	NA, $p=1.95 \times 10^{-6}$ (z: positive), $p=0.002$ (z: negative), NA
7q35	<i>OR2A7</i>	¹⁰	eQTL analyses in TCGA (tumor tissue)	0.23/0.12	$p=0.34$
8q24	<i>POU5F1B</i> , <i>PVT1</i>	¹⁶	eQTL analyses in TCGA (tumor tissue)	NA, 0.03/0.01	NA, $p=1.12 \times 10^{-4}$ (z: positive)
9q31.2	<i>KLF4</i>	^{11,17}	eQTL analyses in TCGA (tumor tissue)	0.02/0	$p=0.007$ (z: positive)
10q21.2	<i>NRBF2</i>	¹⁸	eQTL analyses in Normal breast I (normal tissue) and Breast carcinomas I (tumor tissue)	NA	NA
10q26.13	<i>FGFR2</i>	¹⁹	prediction by ChIA-PET in MCF7 cells, while no association in eQTL analyses in METABRIC (tumor tissue)	0.13/0.02	$p=0.73$
11p15.5	<i>TH</i>	¹⁰	eQTL analyses in TCGA (tumor tissue)	NA	NA
11q13.1	<i>AP5B1</i>	¹⁰	eQTL analyses in TCGA (tumor tissue)	NA	NA
11q13.3	<i>CCND1</i>	²⁰	eQTL analyses in the Helsinki Breast Cancer Study (tumor tissue) suggests borderline association for one SNP rs554219 in a recessive model; while there was no linear trend, and no signal detected in analyses of 40 normal breast tissue samples or TCGA tumor samples	NA	NA
15q26.1	<i>RCCD1</i>	²¹	eQTL analyses in TCGA (tumor and adjacent normal tissue)	0.13/0.07	$p=3.33 \times 10^{-13}$ (z: negative)
16q12.1	<i>TOX3</i>	^{10,11}	eQTL analyses in TCGA (tumor tissue)	$0.02/4.27 \times 10^{-5}$	$p=0.09$
16q13	<i>AMFR</i>	¹	eQTL analyses in METABRIC (tumor adjacent normal tissue); prediction by ChIA-PET in MCF7	NA	NA

			cells		
16q23.2	<i>DYNLRB2</i>	¹⁰	eQTL analyses in TCGA (tumor tissue)	NA	NA
17q22	<i>STXBP4</i>	²²	Index SNP associated with differential transcript expression in TCGA (tumor tissue)	0.03/0.01	$p=2.21 \times 10^{-11}$ (z: positive)
19p13	<i>LRRC25</i> , <i>ABHD8</i>	^{10,23,24}	eQTL analyses in TCGA (tumor tissue); eQTL analyses in normal breast tissue	5.36×10^{-6} /0, NA	$p=0.65$, NA
19q13.31	<i>ZNF404</i> , <i>ZNF155</i>	^{2,10}	eQTL analyses in TCGA (tumor tissue); eQTL analyses in TCGA (tumor tissue)	0.15/0.21, 0.13/0.19	$p=1.15 \times 10^{-13}$ (z: positive), $p=0.03$ (z: positive)
21q22.12	<i>KCNE1</i> , <i>RUNX1</i> , <i>RCAN1</i>	²⁵	eQTL analyses in TCGA (tumor tissue); eQTL analyses in METABRIC (tumor tissue); eQTL analyses in METABRIC (tumor tissue)	0.08/0.06, 0.04/0, NA	$p=0.65$, $p=0.76$, NA

NA, not applicable

Supplementary Table 12. Primer sequences.

Name	Sequence 5' -> 3'
GUSB Fwd	GAAAATATGTGGTTGGAGAGCTCATT
GUSB Rev	CGAGTGAAGATCCCCTTTTTA
PUM1 Fwd	AATGCAGGCGCGAGAAAT
PUM1 Rev	TTGTGCAGCTGAGGAACTAATGA
RPLP0 Fwd	CCATTGAAATCCTGAGTGATGTG
RPLP0 Rev	CTTCGCTGGCTCCCCTTT
ZSWIM5_H_FWD1	AAGACGGTGGCGGAAAAGTG
ZSWIM5_H_REV1	GAAGGACCAGTAGACGATGCG
ZSWIM5_H_FWD2	AGTCGGCTTTCATCTGAGTGG
ZSWIM5_H_REV2	AGGAAGACGCAATTTGACTTGG
ZSWIM5_H_FWD3	CTATCTCCGAAACCCTTTTCCAG
ZSWIM5_H_REV3	TGTGGTGTGCCGTGATTAAATA
KLHDC10_H_FWD1	CTCAACCGCTTCGTGCAAC
KLHDC10_H_REV1	CCTAACTGGGTCCCATCGTATTT
KLHDC10_H_FWD2	TACGATGGGACCCAGTTAGGA
KLHDC10_H_REV2	TGTGGCCTCTCAAAAACCTGT
KLHDC10_H_FWD3	GCACGAAGTGGACATCGTTG
KLHDC10_H_REV3	CCTCCCGATTTCATCATAATCTGG
UBLCP1_H_FWD1	GTGGACAGGAGTATTCAGTGACC
UBLCP1_H_REV1	CAAGTAACTTTTGGCGTTCTGG
UBLCP1_H_FWD2	CTCGCAGAGTGAAAGAGTACAAA
UBLCP1_H_REV2	GCACAAGACCTGTGGTCAAATA
PLEKHD1_H_FWD1	TCCCGGCGGTTTTTCATCATC
PLEKHD1_H_REV1	CCACTGGGTCTGCTCAAACCT
PLEKHD1_H_FWD2	GGAAGAGACCGAAGAAGCTCTGC
PLEKHD1_H_REV2	TGCAAGGACTCCGTGAGGT
ALS2CR12_H_FWD1	ACTTGGGACCACGGAAGCTA
ALS2CR12_H_REV1	GGAGCTGGTACAAGAGGAGTTA
ALS2CR12_H_FWD2	ATGCACAAGCCCTTATCCTAGA
ALS2CR12_H_REV2	AGAGGCCAATCTCCAGAACA

RMND1_H_FWD1	CAGTGCCGAAGAATCGGTCAT
RMND1_H_REV1	CGAGCAGCATTTAATGGAGACA
RMND1_H_FWD2	GCACACCTTCCAACCATGAAA
RMND1_H_REV2	TGGATGCTTTTAGTGGTCTCTTC
RMND1_H_FWD3	GAGACCACTAAAAGCATCCAGG
RMND1_H_REV3	GCAGTGCATTAGGTCCTCGT
STXBP4_H_FWD1	CCTTGGCCTGAAGGTACTAGG
STXBP4_H_REV1	AGCAGATTCTAACCTCAACTTGG
STXBP4_H_FWD2	GAATCTGCTTGGGAGATAGCATT
STXBP4_H_REV2	TGAGGCTTGAGGTCCATATTCT
STXBP4_H_FWD3	ATCCCTCTGTTCGCTTTAAGGC
STXBP4_H_REV3	TCAGGGCTTGGTGTTGTTCC
ZNF404_H_FWD1	AAGTAAATGCGTACCATCAGGAG
ZNF404_H_REV1	TCCCACTTTAGGTCTCTGTTGT
ZNF404_H_FWD2	GGCCTTTGTTCGCAGCTATCT
ZNF404_H_REV2	AGGCTTGAGCCCTTACCAAAA
ZNF404_H_FWD3	GGCCTTTTGTAGAGGCTCTCA
ZNF404_H_REV3	AAGGTCTCCAACACGACTGAA
PIDD1_H_FWD1	TCAGAGGATTCGGACGCAG
PIDD1_H_REV1	GTGAGTGCTCAGACGCAAGAA
PIDD1_H_FWD2	GAGCCTCGTCGAGTCTCCAT
PIDD1_H_REV2	GGCCCAGTACAACAGGTGC
PIDD1_H_FWD3	CTCACCCACCTGTACGCAC
PIDD1_H_REV3	CAGAGCGATGAGGTTACAC
NRBF2_H_FWD1	CAGACGAGCAGACCGTTTATT
NRBF2_H_REV1	TGCTGGGCTTTCAATCTTTCTT
ABHD8_H_FWD1	GGGGTGACCGACGGTATCT
ABHD8_H_REV1	GGCTTGACCTCTACAAAGGTG
ABHD8_H_FWD2	TCGAGCCGACCTCCTACAC
ABHD8_H_REV2	TTTGCAGCTAGTGATGCGCTT
ABHD8_H_FWD3	CTGAGGACATGCGAGCAATCT
ABHD8_H_REV3	GAAAGAGACACCGTAGGAATGG

RP11-218M22.1_H_R1_FWD1	CGGGAAAAGATGGAGTGAAGGT
RP11-218M22.1_H_R1_REV1	GGCACTTCCGCTAATGCTG
RP11-218M22.1_H_R1_FWD2	TGAGCCGGGAAAAGATGGAGT
RP11-218M22.1_H_R1_REV2	GCACTTCCGCTAATGCTGAGG
RP11-218M22.1_H_R2_FWD1	CACTGAGAGAAGCAGGAGAATGT
RP11-218M22.1_H_R2_REV1	AAGAGAGATTGTCTCGCAGTC
RP11-218M22.1_H_R2_FWD2	ACTGAGAGAAGCAGGAGAATGT
RP11-218M22.1_H_R2_REV2	AAAGAGAGATTGTCTCGCAGTC
AP006621.6_H_FWD1	TCCTGAGGGCCGACTCTAC
AP006621.6_H_REV1	CGTCTTAGCGGCTGTCACTT
AP006621.6_H_FWD2	ACTGAGAGAAGCAGGAGAATGTT
AP006621.6_H_REV2	CACTAAAGAGAGATTGTCTCGCA
RP11-467J12.4_H_FWD1	GGGGTGGTGGGTGTCACTAA
RP11-467J12.4_H_REV1	ATTCACCTTCACCAGGGCAC
RP11-467J12.4_H_FWD2	TCACTAAAAGGAACCAGCCCC
RP11-467J12.4_H_REV2	CTCTGACTGATTACCTTCACCA
RP11-15A1.7_H_FWD1	CAGAGTGTGTCTGGACTCCG
RP11-15A1.7_H_REV1	CCAGGCGCTCAGAGATATGG
RP11-15A1.7_H_FWD2	GCGACTCAGAGTGTGTCTGG
RP11-15A1.7_H_REV2	ATGGAATACGTTCCCGGTGG
CTD-3032H12.1_H_FWD1	CCTACACGAGGCCAGAGATCC
CTD-3032H12.1_H_REV1	CCTAACAGCAGATCGCCTCG
CTD-3032H12.1_H_FWD2	GCCCGTGGCCTACACGAG
CTD-3032H12.1_H_REV2	CGGGTCTTCCTTTGTGTCCAG
B2M-FWD-1	GAGGCTATCCAGCGTACTCCA
B2M-REV-1	CGGCAGGCATACTCATCTTTT
B2M-FWD-2	CTCACGTCATCCAGCAGAGA
B2M-REV-2	CGGCAGGCATACTCATCTTT
B2M-FWD-3	AGGCTATCCAGCGTACTCCA
B2M-REV-3	CGGCAGGCATACTCATCTTT
ARHGDIA-FWD-1	GGATGAGCACTCGGTCAACTA
ARHGDIA-REV-1	GGCCTCCTTGTACTTTCGCAG

ARHGDIA-FWD-2	GAGCCTGCGAAAGTACAAGG
ARHGDIA-REV-2	TCCTTCAGCACAAACGACTG
ARHGDIA-FWD-2	TGCCTCTGCCTTTTCTGTCT
ARHGDIA-REV-3	GCACTTGGTCCCTTGTTTGT
ZAP70-FWD-1	CGAGCGTGTATGAGAGCCC
ZAP70-REV-1	ATGAGGAGGTTATCGCGCTTC
ZAP70-FWD-2	ACGCCAAGATCAGCGACTTT
ZAP70-REV-2	GGGTGCGTACCACTTGAGC
ZAP70-FWD-3	CTGGAGCTATGGGGTCACCA
ZAP70-REV-3	CAGGCTGTAGTAACAGGCTCG

Supplementary Table 13. Predicting variants in gene expression prediction models for the identified associated genes after Bonferroni correction

<i>RP11-439A17.7</i>	rs17023394, rs12037207, rs838518, rs17023457, rs838522, rs699774, rs838532, rs838530, rs838528, rs3820032, rs3753264, rs3753263, rs3753262, rs2185556, rs10754396, rs12405488, rs1417610, rs1417609, rs12024495, rs2050892, rs10923824, rs4659178, rs10802098, rs10923836, rs2275609, rs7547046, rs3949342, rs947130, rs4659182, rs7553527, rs6692504, rs4659200, rs346670, rs2024838, rs10754414, rs12046880, rs10802122, rs4659221, rs10494228, rs347910, rs838990, rs404937, rs380155, rs598100, rs12025390, rs4659226, rs663807, rs616111, rs539304, rs539426, rs17258425, rs4391705, rs532208, rs17186233, rs753424, rs10923902, rs947269, rs3009197, rs3009196, rs2994815, rs2994816, rs2994817, rs3009182, rs3009184, rs3009186, rs2487573, rs835578, rs4659245, rs4659246, rs4659247, rs2843021, rs12075536, rs947273, rs699780, rs835574, rs835573, rs2793830, rs6688004, rs4659248, rs2453056, rs5025718, rs2493420, rs1493696, rs2493411, rs2453044, rs10903159, rs4844381, rs12145080, rs11249348, rs6600671, rs2319969, rs2319971, rs11249431, rs2222371
<i>ZSWIM5</i>	rs6690437, rs12754891, rs12091565, rs11210998, rs2120823, rs17386059, rs12732315, rs16832024, rs12744658, rs12749754, rs1889759, rs6688710, rs7525308, rs6429550, rs7517439, rs7517639, rs6692487, rs11579580, rs12733586, rs7528461, rs12139143, rs6703452, rs7531019, rs11211053, rs11577974, rs12735637, rs12125367, rs12755554, rs11211059, rs7519454, rs263997, rs263992, rs263991, rs263989, rs183809, rs11556200, rs264025, rs264022, rs12749130, rs12738542, rs7553658, rs6692713, rs4399199, rs12126314, rs12126318, rs2202152, rs11579411, rs10789463, rs6429566, rs12743512, rs937291, rs10789465, rs2275276, rs11580609, rs7903, rs11211129, rs644915, rs512026, rs518365, rs12141928, rs6696085, rs12406217, rs12141269, rs17102087, rs12137934, rs12146051, rs12142240
<i>KLHDC7A</i>	rs4920399, rs11203247, rs17435018, rs7517220, rs6665151, rs11261017, rs11261020, rs4920322, rs4920323, rs11261021, rs2992745, rs3000058, rs2816030, rs2230705, rs6683394
<i>ALK</i>	rs4665406, rs7576048, rs13029274, rs12995493, rs10190267, rs2940806, rs12052472, rs1992810, rs2276551, rs2276549, rs4666201, rs2293564, rs12997783, rs4666202, rs7561975, rs6731724, rs12993746, rs12997218, rs11897665, rs7564775, rs7562088, rs6753532, rs4414641, rs4665485, rs11127243, rs13010777, rs12476676, rs12465499, rs4233750, rs12478888, rs12478928, rs6547981, rs7603844, rs7560160, rs4502372, rs5018731, rs11690664, rs7576793, rs12714298, rs829602, rs2253121, rs1474194, rs17324662, rs7594598, rs7597567, rs13388219, rs13385578, rs1862960
<i>CASP8</i>	rs6728002, rs3754935, rs7603014, rs6735656, rs6754084, rs1861270, rs2293554, rs10931936, rs1035142, rs700635, rs6743068, rs13016963, rs9288316, rs7560328, rs2597900
<i>ALS2CR12</i>	rs1035142, rs13016963, rs9288316, rs7560328
<i>PRSS46</i>	rs11716028, rs6808473, rs7632165, rs11717357, rs9843503, rs4413345, rs12495098, rs10510751, rs3918357, rs743661,

	rs916092, rs1520484, rs9819159, rs9836993, rs9880885, rs9829227, rs9810013, rs3796367, rs3796369, rs3796370, rs9834713, rs9835025, rs11915788, rs9820361, rs9820372, rs9820785, rs9820845, rs9820861, rs9821418, rs9841203, rs9841229, rs9864097, rs9868357, rs7632176, rs7623501, rs7626129, rs7428736, rs7428787, rs7639979, rs12489663, rs12496832, rs6793235, rs6787229, rs6784957, rs376737976, rs1014229, rs11714840
<i>LRRC3B</i>	rs2052760, rs11711082, rs11719046, rs11715434, rs10510576, rs11719770, rs11719901, rs17018100, rs994169, rs973603, rs11712421, rs17018155, rs1158545, rs1158544, rs12633309, rs17018167, rs2036430, rs2036428, rs9820211, rs1602349, rs4435583, rs1907178, rs6808839, rs1488240, rs9845198, rs6794554, rs6551142, rs7646852, rs1488215, rs17018761, rs1386884, rs11717214, rs1915915, rs9841537, rs12495557, rs1522140, rs7618127, rs13080907, rs13098500, rs6765909, rs4586769, rs10510594, rs1603051, rs7611368, rs11707188, rs3892373
<i>NUDT17</i>	rs9286836, rs11587364, rs12402787, rs12732381, rs2040085, rs2040086, rs34695381
<i>ANKRD34A</i>	rs704985
<i>RP11-114I8.4</i>	rs12639465, rs6441308, rs1287283, rs7616988, rs9873709, rs10511177, rs9875640, rs6807176, rs6799379, rs6806178, rs9815439, rs1021341, rs9814359, rs6790535
<i>SPATA18</i>	rs7440594, rs10012938, rs10434448, rs17577020, rs4864836, rs6856794, rs11724730, rs225160, rs225163, rs13989, rs225165, rs225170, rs419792, rs17612170, rs11947242, rs4470701, rs7665551, rs730284, rs4865271, rs12510605, rs9683559, rs7692441, rs7690931, rs1501614, rs1841263, rs6858566, rs10517269, rs11939448, rs11938159, rs6838718, rs7693568, rs6835977, rs17644026, rs11133238, rs4864440, rs11133239, rs11929934, rs13434989, rs10012324, rs17082294, rs10517278, rs4864717, rs17646076
<i>RP11-53O19.1</i>	rs80316101, rs150134525, rs7720551, rs76768074, rs148946381, rs181072007, rs186001811, rs111765202, rs35601455, rs112494990, rs144785376, rs62366821, rs112679498, rs13179565, rs201180654, rs191324191
<i>ATG10</i>	rs4703825, rs12187089, rs11738172, rs432872, rs457049, rs456778, rs463247, rs457700, rs386424, rs462122, rs11740142, rs1384256, rs11741569, rs1485587, rs11740648, rs11741303, rs2860007, rs11748868, rs12515069, rs1428939, rs178957, rs2407064, rs10068160, rs3857369, rs10061458, rs17245188, rs9293290, rs4703537, rs891159, rs4703870, rs10066167, rs10065463, rs1543911, rs2059891, rs6895884, rs6888977, rs6884232, rs1019806, rs4703879, rs2407153, rs11747683, rs749402, rs749401, rs862240, rs146991557, rs226204, rs11743578, rs2407156, rs17247678, rs6861268, rs58757861, rs715888, rs1827391, rs168534, rs355285, rs4703914, rs6885480, rs10473857, rs4605761, rs1505073, rs40214, rs256795
<i>ATP6AP1L</i>	rs6881927, rs12517153, rs13174473, rs10042996, rs12188888, rs2972230, rs10514220, rs1561150, rs442417, rs461802, rs4703852, rs2406905, rs6892261, rs16899359, rs7727483, rs178957, rs3857369, rs10061458, rs9293290, rs4703537, rs10066167, rs10065463, rs3738, rs226202, rs226199, rs6880209, rs178931, rs226196, rs862240, rs146991557, rs862239, rs3756683, rs3734115, rs224844, rs224843, rs6872917, rs12187334, rs4703894, rs905221, rs2385882, rs12153244, rs10062095, rs2015904, rs2015911, rs4354012, rs4282294, rs16899813, rs924615, rs10514242, rs10041952, rs16899864, rs9293313, rs17205439, rs10462383, rs6863549, rs13165887, rs13166060, rs10474061,

	rs10474062, rs10051128, rs12658884, rs7713051, rs1363948, rs9283781, rs11744735, rs433820, rs37558, rs4703936
<i>UBLCP1</i>	rs31199, rs10054046, rs10070382
<i>RP11-32D16.1</i>	rs11466807, rs11744671, rs11466784, rs3097837, rs254664, rs1896606, rs13155377, rs13159354, rs7709209, rs2984629, rs17242576, rs11741271, rs2988321, rs1952657, rs12187534, rs10045014, rs12188478, rs10515764, rs17055744, rs2419654, rs1317414, rs864821, rs824869, rs824871, rs699083, rs10040448, rs13171583, rs11750665, rs11747709, rs11741746, rs13175305, rs11743605, rs7715522, rs11748882, rs31193, rs13155183, rs1345721, rs7736575, rs2009612, rs1469070, rs7730360, rs13172711, rs4130208, rs2287763, rs6881547, rs12187372
<i>BTN3A2</i>	rs9467504, rs13203202, rs2328879, rs6456693, rs10946795, rs6924948, rs9467701, rs6923139, rs9467704, rs6903015, rs9467707, rs6939978, rs9366653, rs9379851, rs9348709, rs9393705, rs9393706, rs9393707, rs9393708, rs9358932, rs9358934, rs9357006, rs9379855, rs9348712, rs9379856, rs9379857, rs9379858, rs9393710, rs9379859, rs9358935, rs12173854, rs9379864, rs12176317, rs12174602, rs12174631, rs13216828, rs9393713, rs9393714, rs9358937, rs2073529, rs2073531, rs9348716, rs9366655, rs1977, rs1978, rs1979, rs3799380, rs4518487, rs6456728, rs9348721, rs9467772, rs3799383, rs6920256, rs16901784, rs2494691, rs2451746, rs9366673, rs9393777, rs17687319, rs17687372, rs17687396, rs17739727, rs12213055, rs9468076, rs2076305, rs2064219
<i>L3MBTL3</i>	rs9321204, rs9388762, rs7754426, rs6569648, rs7740107
<i>RP11-73O6.3</i>	rs9388721, rs9372945, rs17755387, rs9402157, rs4499953, rs17811901, rs12191170, rs12202100, rs6569644, rs9388762, rs7754426, rs7740107, rs4364506, rs9492440, rs10499172, rs7759381, rs9492441, rs7769599, rs12190724, rs7764762, rs4548027, rs9492443, rs12198331, rs9492445, rs9492446, rs9492447
<i>RMND1</i>	rs11759741, rs11751703, rs17800315, rs17800327, rs11759502, rs11757075, rs2223451, rs7451945, rs742315, rs9371462, rs7769835, rs6557545, rs9384561, rs9371463, rs11752947, rs9383832, rs11155740, rs7742124, rs4870532, rs17080057, rs17080062, rs17080069, rs17080087, rs17080089, rs17080091, rs17080093, rs17080102, rs9372087, rs10872665, rs1494309, rs782665, rs803401, rs2295083, rs9397029, rs742829, rs6902496, rs12203650, rs7349940, rs12205664, rs1575219, rs2180927, rs9478167, rs730489, rs4869987, rs4869988, rs6923696, rs17362091, rs4869991, rs6939639, rs2179544, rs12210237, rs3800270, rs6904364, rs6557140, rs9322321, rs9397050, rs7752091, rs3736175, rs6557141, rs9383571, rs954238, rs2982558, rs2982556, rs1124674, rs9479117, rs1709180, rs3020404, rs2982683, rs9340978, rs3798577, rs9383963, rs7450824, rs2813549, rs2813550, rs2763033, rs2635469, rs1324452, rs2813504, rs12174080, rs2788381, rs1890100, rs3816850, rs4472361, rs7776230, rs725235, rs4870095, rs6913579, rs4870101, rs4870102, rs214955, rs214992
<i>KLHDC10</i>	rs4728160, rs721691, rs2896415, rs4731568, rs7800983, rs11764547, rs10246160, rs10225672, rs2727455, rs10480805, rs1046691, rs10246707, rs17162050, rs10249037, rs17162123, rs9656386, rs17162136, rs10273782, rs6467267, rs17162295, rs10500121, rs10253233, rs6962745, rs7803795, rs7801603, rs7805980, rs10279425, rs12531444, rs12534580, rs901799, rs1488009, rs1574704, rs6943386, rs7793239, rs2129902, rs6467309, rs10257888, rs10263075, rs1035596, rs10281580, rs10272075, rs10272206, rs10248834, rs10248294, rs10279517, rs6969737, rs6945822,

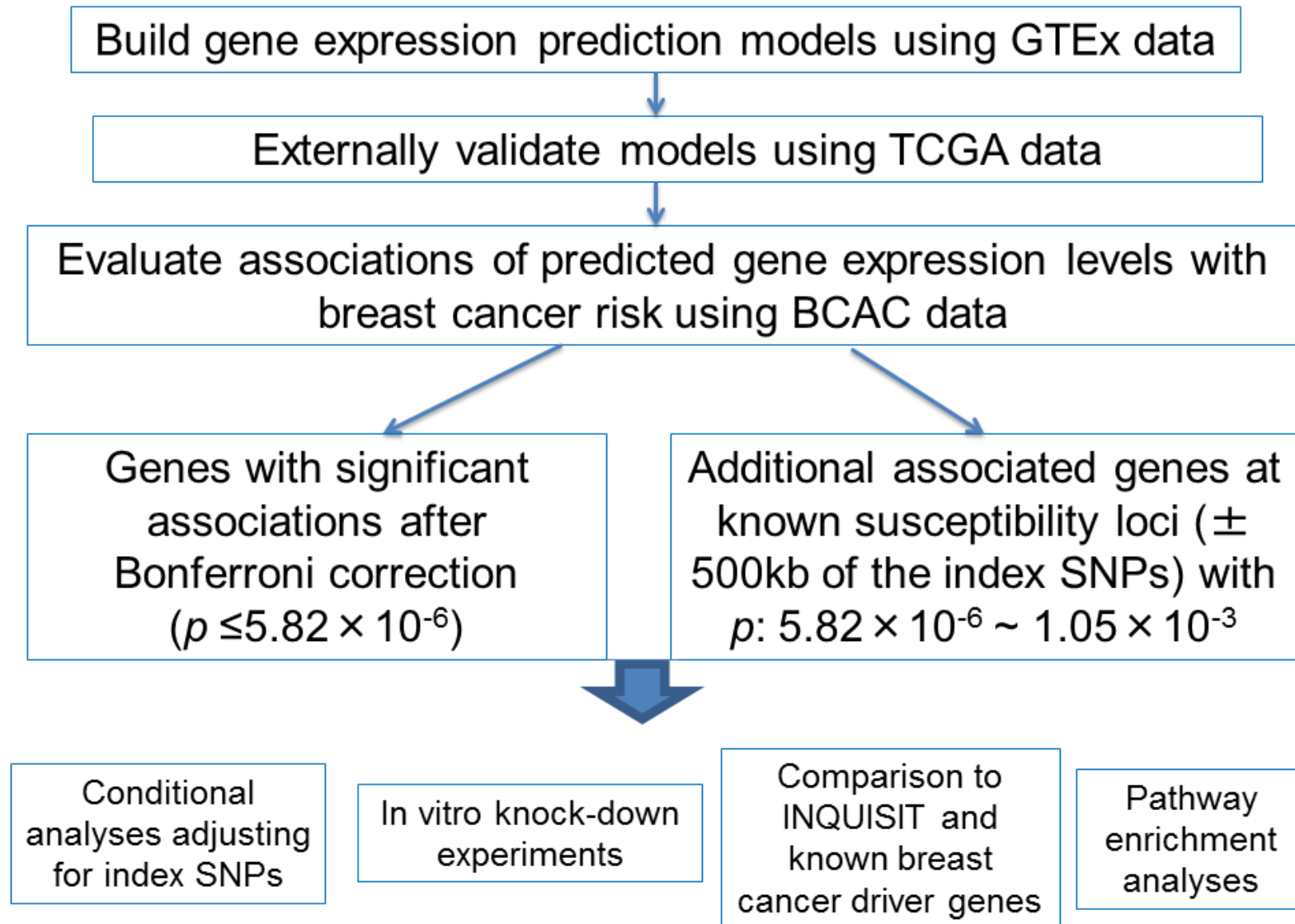
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<i>RPLP2</i>	rs11245936, rs7103978, rs28514396, rs2943510, rs11029039, rs6578471, rs4752763, rs11604009, rs10400297, rs10838484, rs1038727, rs7110331, rs575488, rs2292958, rs2292963, rs1317356, rs11041082, rs7127542, rs6598055, rs7925234, rs11246052, rs9737419, rs10902120, rs11602841, rs7947900, rs7395116, rs11246068, rs4131942, rs7117996, rs10794314, rs10794315, rs11246108, rs11246130, rs11246131, rs10902165, rs12806187, rs4963166, rs7635, rs10902208, rs7952095, rs12277141, rs10902221, rs11246314, rs11246316, rs28633403
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<i>RP11-867G23.10</i>	rs78407319, rs190536043, rs118019315, rs2270448, rs118151305
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<i>GALNT16</i>	rs17105278, rs7150454, rs12889206, rs8003738, rs4902567, rs1810623, rs6573834, rs1570106, rs17105586, rs916962, rs2247048, rs2525521, rs1476586, rs1859302, rs2525523, rs2525524, rs2525525, rs2525526, rs2525527, rs2842331, rs7153476, rs8007194, rs2257111, rs2257116, rs2257127, rs4899246, rs10137893, rs4902611, rs181464, rs17835996, rs1275195, rs1950712, rs7143336, rs1890941, rs2185492, rs7155178, rs7151003, rs8008770, rs12889279, rs1958184, rs10873226, rs12884186, rs11158830, rs1009256, rs2182972, rs9323533, rs41350744, rs1469253, rs7141710, rs4902805, rs12590132, rs1015585, rs4902806
<i>PLEKHD1</i>	rs9323513, rs11158749, rs10134446, rs2189517, rs7140266, rs2525530

<i>CTD-3051D23.1</i>	rs8095, rs1053419, rs11850704, rs4906423, rs4555088, rs3809461, rs3809457, rs7144812, rs4454893, rs7142224, rs10136937, rs4340263, rs10135130, rs3809454, rs2841214, rs2582559, rs4983589, rs3825761, rs7151594, rs880616, rs10139596, rs1882848, rs10140111, rs11625865, rs4983590, rs11160839
<i>MAN2C1</i>	rs2075589, rs4886649, rs1809714, rs1984586, rs1984587, rs7164976, rs3866545, rs7183520, rs7166852, rs7164429, rs8028182, rs12708519, rs8029112, rs28610581, rs8030802, rs6495182, rs8023268, rs8023815, rs11636031, rs11636199, rs12708520, rs7163907, rs28693593, rs4886716, rs4075522, rs7184046, rs13380103
<i>CTD-2323K18.1</i>	rs17336243, rs2304900, rs12908919, rs1822324, rs12911696, rs12899456, rs12909554, rs8038911, rs12905302, rs1809714, rs1984586, rs1984587, rs7164976, rs3866545, rs7183520, rs7166852, rs7164429, rs9673084, rs5745935, rs8027749, rs11635996, rs3210683, rs1128585
<i>RCCD1</i>	rs11855570, rs8030486, rs2227935, rs7167216, rs734252, rs2677744, rs1266489, rs1266483, rs4773, rs1550636, rs2290202, rs2301825, rs3826033, rs4392040, rs7402585, rs12915069, rs9744944, rs8028382, rs4583214, rs4244910, rs4932591, rs4306482
<i>RP11-467J12.4</i>	rs17257857, rs12597737, rs7200881, rs17201162, rs12598784, rs1344490, rs3910446, rs9889099, rs1362380, rs9936470, rs16950876, rs8048212, rs17268400, rs3095536, rs3095537, rs1076081, rs1861315, rs16951015, rs1074734, rs16951035, rs1345312, rs16951056, rs12597728, rs4477699, rs4480800, rs1362558, rs11075488, rs9921890, rs1861527, rs3095599, rs3095600, rs194392, rs194394, rs12930211, rs7191789, rs1362553, rs8048309, rs1362554, rs1345389, rs2075236, rs3095660, rs1420546, rs3095661, rs40841, rs1362560, rs3095616, rs1420548, rs8051542, rs4784220, rs12922061, rs11647542, rs11866049, rs16951465, rs11867085, rs3104823, rs3112587, rs16951525, rs12925035, rs4784253, rs12919531, rs4238756, rs4783785, rs1420257, rs6499105, rs7205069, rs17298178, rs17370363, rs11639509, rs7500472, rs12919591, rs12922267, rs2387879, rs551415417, rs9925367, rs9936502, rs10153135, rs16951919, rs4783804, rs9925003, rs7198530, rs12933494, rs12919486, rs12930884, rs8058720, rs3760010, rs4456500, rs8051064, rs8047647, rs1833205, rs1833207, rs12051480, rs8045574, rs2388117, rs9924562, rs2160290, rs3743772, rs7186754, rs3095633, rs17802269, rs16952362, rs1477199, rs11861870, rs16952517, rs6499642, rs6499643, rs16945088, rs11075994, rs7195539, rs6499653, rs12149433, rs9926180, rs2111113, rs7500562, rs13335343, rs1362570, rs16952634, rs9937234, rs10852525, rs12921721, rs1344503, rs12232391, rs8053966, rs17821714, rs11864972, rs7185301, rs7185479, rs17224310, rs17224394, rs11643535, rs7194907, rs8053888, rs7185783, rs16952730, rs9933107, rs8056502, rs8061928, rs16952756, rs13335453, rs9922370, rs1971037, rs1125337, rs1125338, rs11076015
<i>CTD-3032H12.1</i>	rs6499657, rs17823199, rs17196003, rs748815, rs13333140, rs16953503, rs12926529, rs9926409, rs1420289, rs2160294, rs933517, rs1420290, rs1420292, rs8059628, rs1186818, rs7205346, rs16953806, rs12325292, rs7198507, rs4435250, rs4783866, rs2589010, rs8053467
<i>LINC00671</i>	rs72826975
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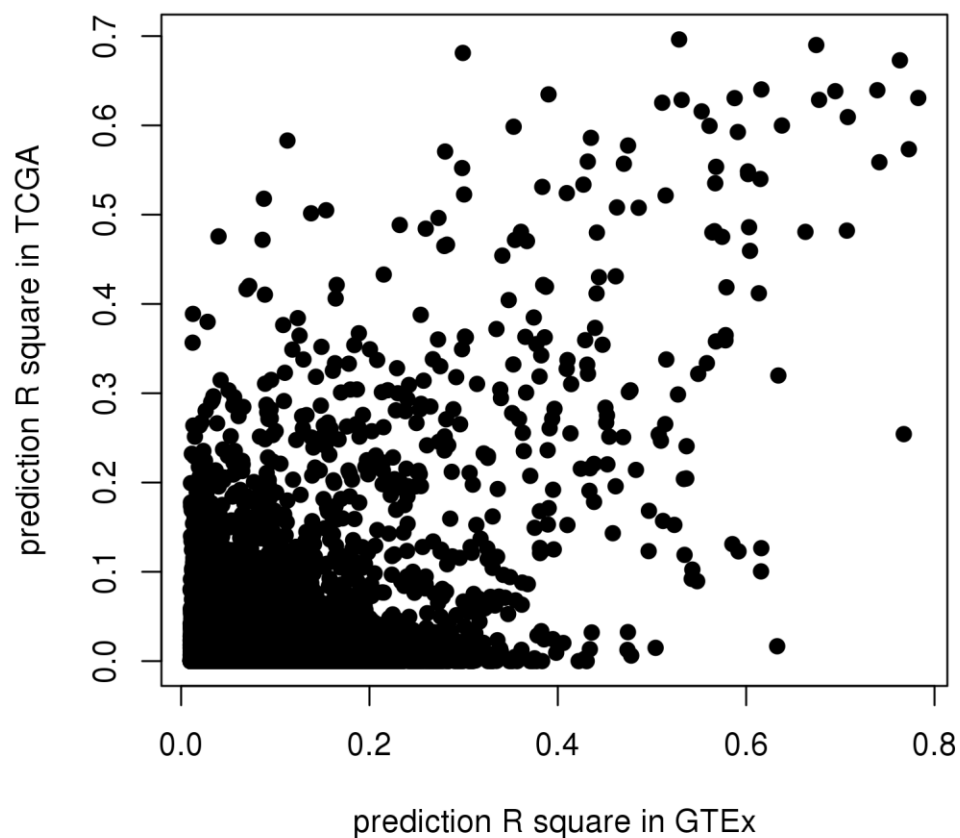
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<i>KANSLI-ASI</i>	rs1133458, rs7213493, rs9911183, rs11079502, rs2072090, rs16939943, rs7222751, rs7214661, rs7222377, rs1059504, rs1230106, rs439558, rs17687796, rs1358071, rs1989480, rs8082105, rs12953076, rs12938031, rs4076452, rs9892359, rs171441, rs242939, rs171443, rs17690703, rs753235, rs2435205, rs3785883, rs2471738, rs2532345, rs2696531, rs148126555, rs538628, rs169201, rs199439, rs199457, rs199456, rs199451, rs199448, rs199445, rs199443, rs199535, rs199534, rs9896243, rs199533, rs35732828, rs2074404, rs199498, rs199497, rs3851781, rs7214920, rs7213046, rs1004485, rs3851788, rs2317996, rs2317995, rs3851797, rs2316757, rs3851800, rs6504640, rs8078196, rs8074704, rs9905068, rs7210126, rs2316759, rs7221913, rs11653838, rs7216950, rs4968304, rs4074249, rs11079750, rs12950699, rs11871364
<i>CRHRI</i>	rs12942842, rs17544947, rs6503404, rs10853009, rs1071682, rs1000354, rs12947718, rs12942666, rs7222389, rs34018943, rs11012, rs9730, rs17631303, rs2077606, rs3946526, rs17631676, rs9890016, rs2435200, rs4630591, rs183211, rs199436, rs199438, rs142167, rs7224296, rs199453, rs199452, rs199449, rs199444, rs199442, rs199524, rs199520
<i>STXBP4</i>	rs2164160, rs1434739, rs974588, rs11653274, rs11079129, rs17709877, rs17675596, rs7210359, rs12952253, rs7209926, rs7208123, rs9916547, rs8065361, rs9891704, rs9900816, rs9907961, rs17745183, rs2787497, rs2787481, rs244317, rs7218719, rs7223718, rs7213282, rs3931318, rs11079178, rs7218226, rs8069305, rs8073227, rs4794602, rs17211444, rs11653426, rs906580, rs12940838, rs8066578, rs11656691, rs12938239, rs12936661, rs8076964, rs7208778, rs13380851, rs11656314, rs11656373, rs6504993, rs6504994, rs12951953, rs6504995, rs11658412, rs10515108, rs10515109, rs7218936, rs376416681, rs7223017, rs7225097, rs8068391, rs7220718, rs2055661, rs8069548, rs16956323, rs9903179, rs9902928, rs9903177
<i>HAPLN4</i>	rs12976333, rs12985909, rs1363119, rs10415765, rs10423237, rs10415319, rs10401774, rs8102478, rs3746183, rs3746181, rs1363120, rs888663, rs888669, rs35742476, rs10420384, rs2303692, rs7246788, rs2891676, rs4808803, rs11672385, rs1971093, rs8111582, rs8111397, rs10426768, rs4808807, rs2013069, rs11670392, rs7249760, rs10409346, rs10409408, rs4808135, rs1469412, rs6512269, rs4580317, rs4808812, rs8106068, rs4808814, rs10422974, rs10415320, rs7249453, rs34992005, rs17214879, rs247765, rs247781, rs248969, rs248950, rs7247625, rs7248612, rs2116889, rs12980778, rs2080261, rs7250793, rs258583
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Supplementary Figure 1. Study design flow chart

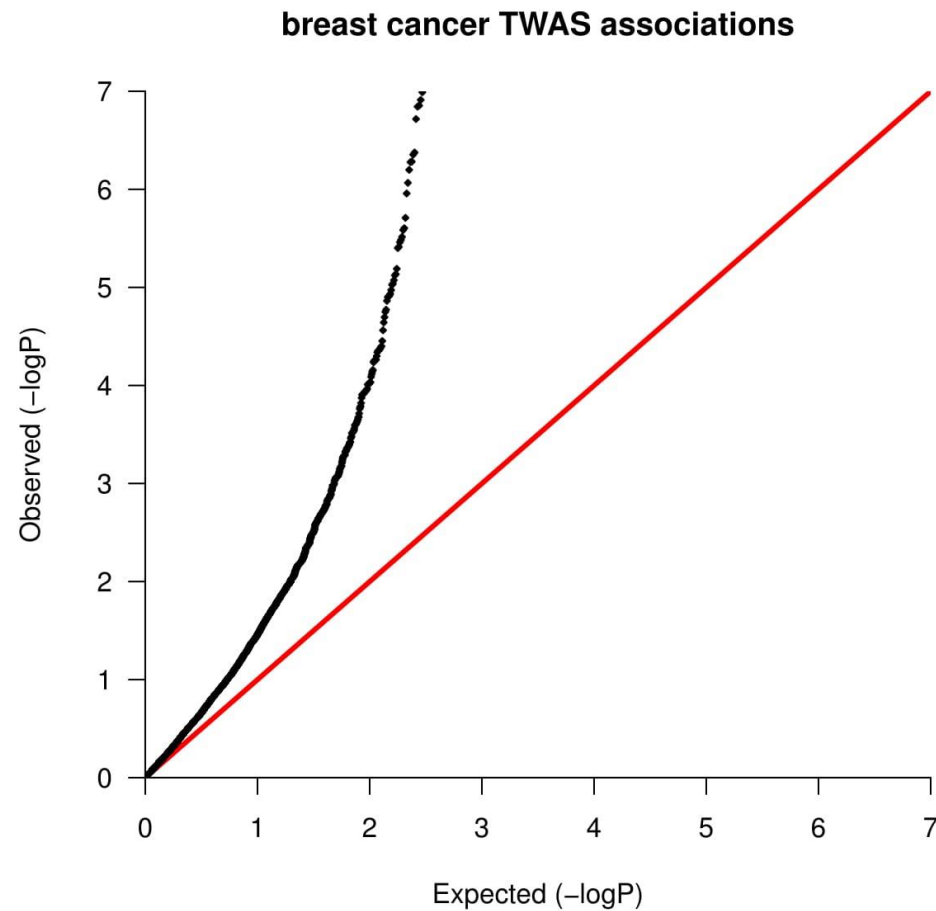


Supplementary Figure 2. Performance of expression prediction models in GTEx and TCGA datasets for genes with at least 10% correlation in GTEx data. The x axis represents the prediction performance (R^2) in GTEx dataset. The y axis represents the prediction performance in TCGA dataset. Each dot represents the expression prediction model for one gene. There is a trend that genes with a high internal prediction performance in GTEx data also have a high external prediction performance in TCGA data (correlation coefficient: 0.55).

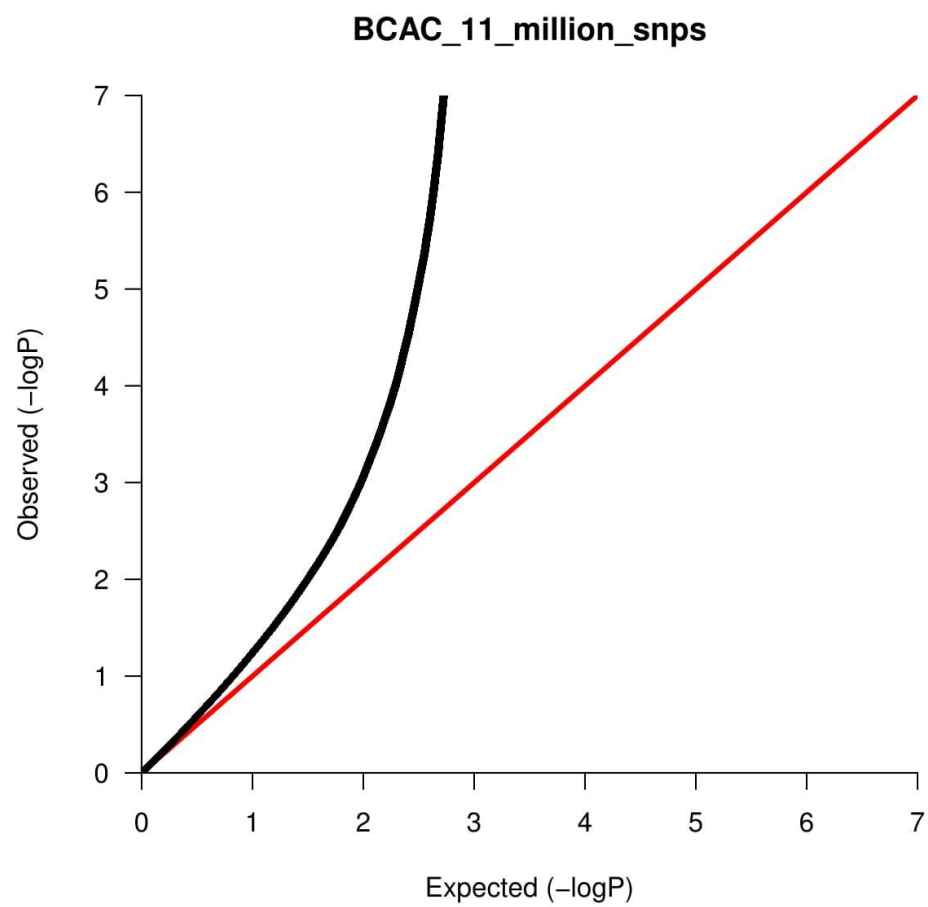


Supplementary Figure 3. Quantile-quantile plots. (A) Quantile-quantile plot of P values in $-\log$ scale of associations between genetically predicted expression levels of 8,597 genes and breast cancer risk; (B) Quantile-quantile plot of P values in $-\log$ scale of associations between the over 250,000 SNPs predicting expression levels of the 8,597 genes and breast cancer risk in BCAC; (C) Quantile-quantile plot of P values in $-\log$ scale of associations between all 11.8 million SNPs and breast cancer risk in BCAC

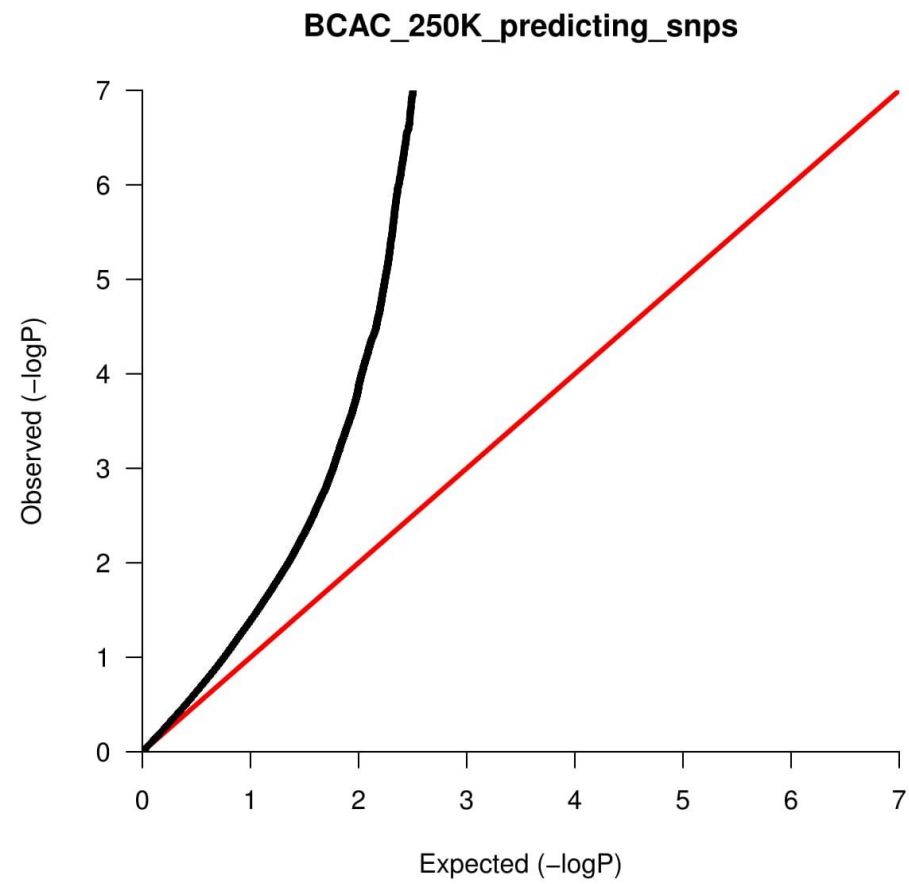
(A)



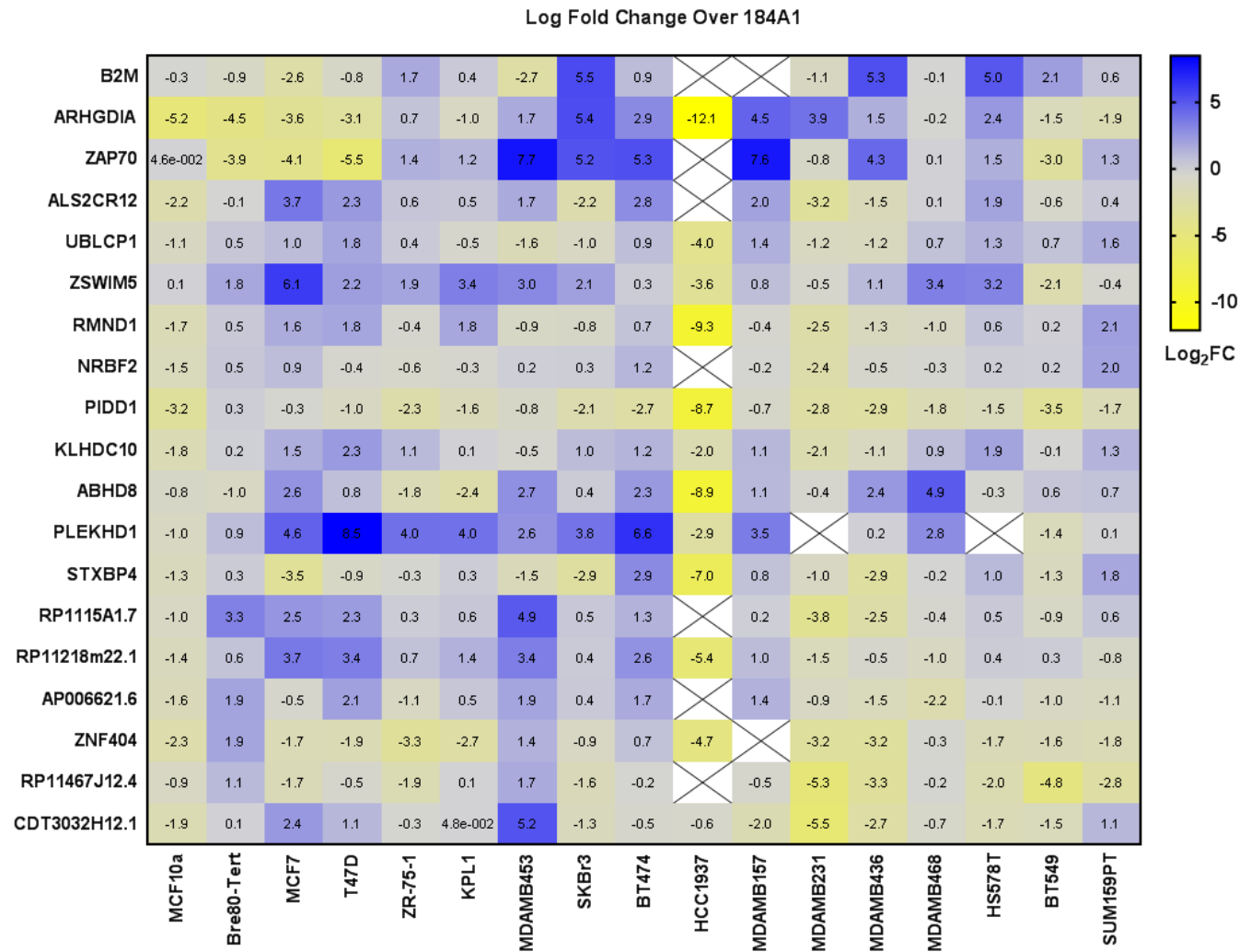
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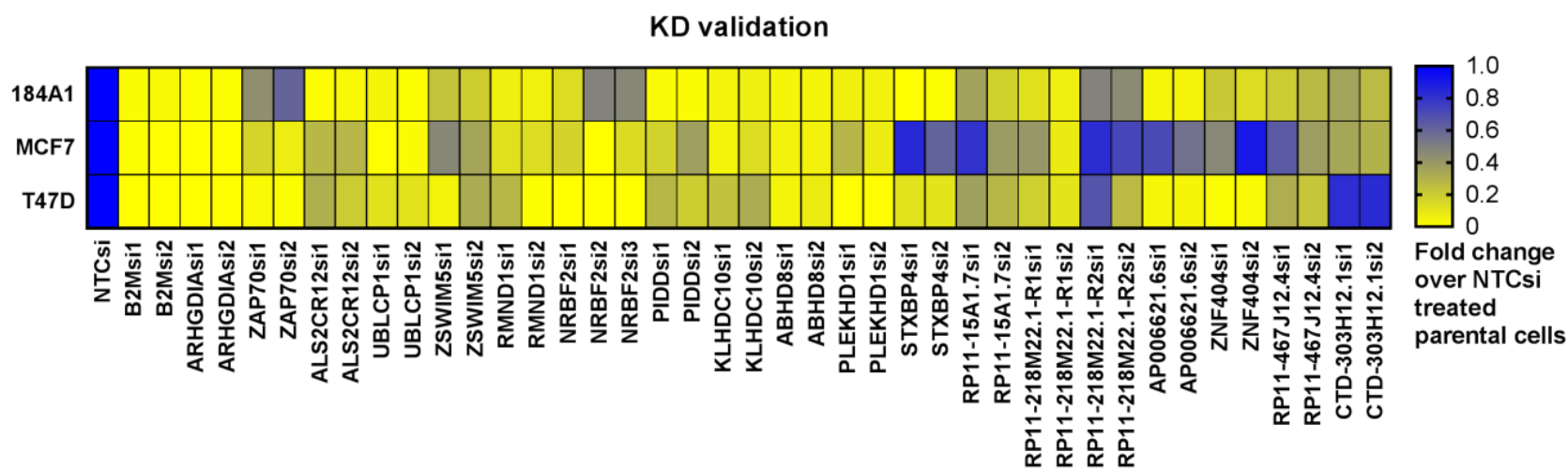
(C)



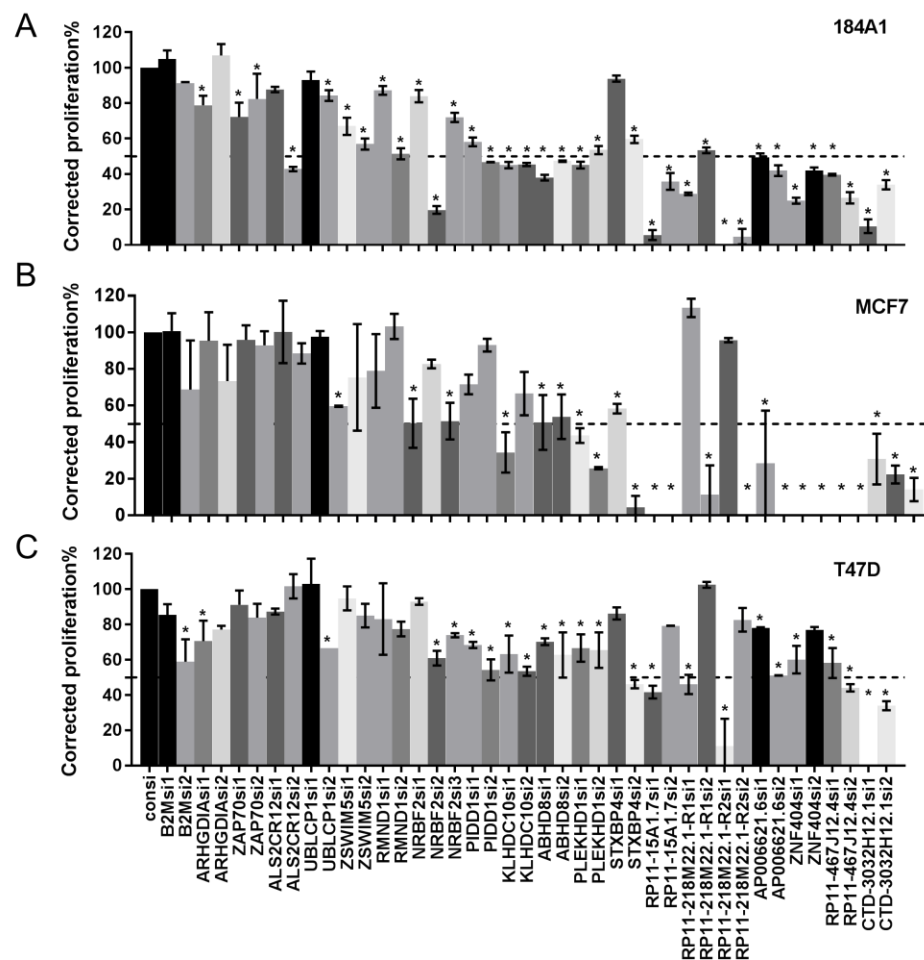
Supplementary Figure 4. Heat map of log fold change (FC) of selected genes normalized to expression levels in 184A1 breast cells. Two or three primer sets were designed for each gene (y-axis) and mRNA levels quantified by qPCR in indicated cells lines (x-axis), including 184A1. The FC of genes normalized to that in 184A1 = mRNA level in indicated cells / mRNA level in 184A1. The \log_2 FC over 184A1 is depicted as a heat map. An X represents “not detectable” with all primer sets.



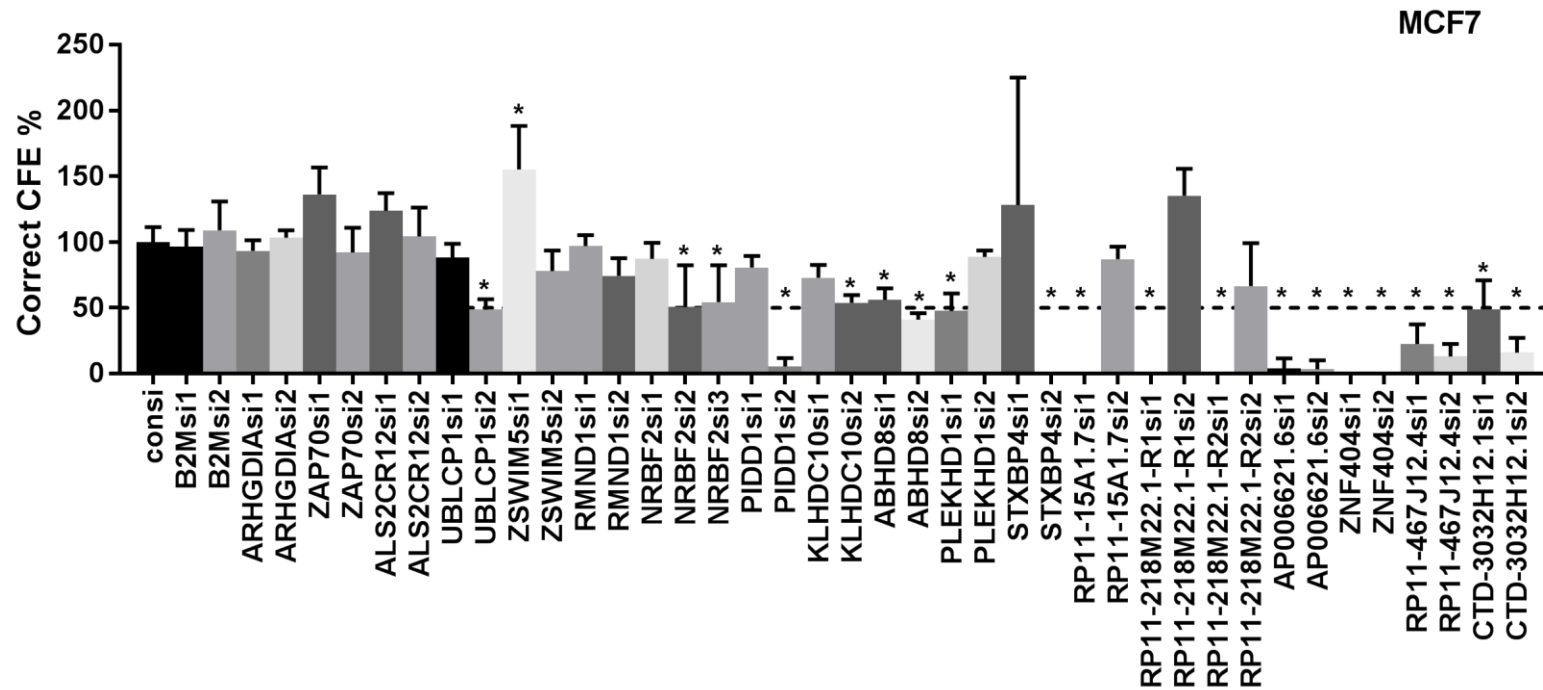
Supplementary Figure 5. Validation of knockdown. 184A1, MCF7 and T47D cells, transfected with the indicated siRNAs, were harvested after 36 hours for qPCR analysis to assess knockdown efficiency. The fold changes over NTCsi-transfected parental cells were plotted.



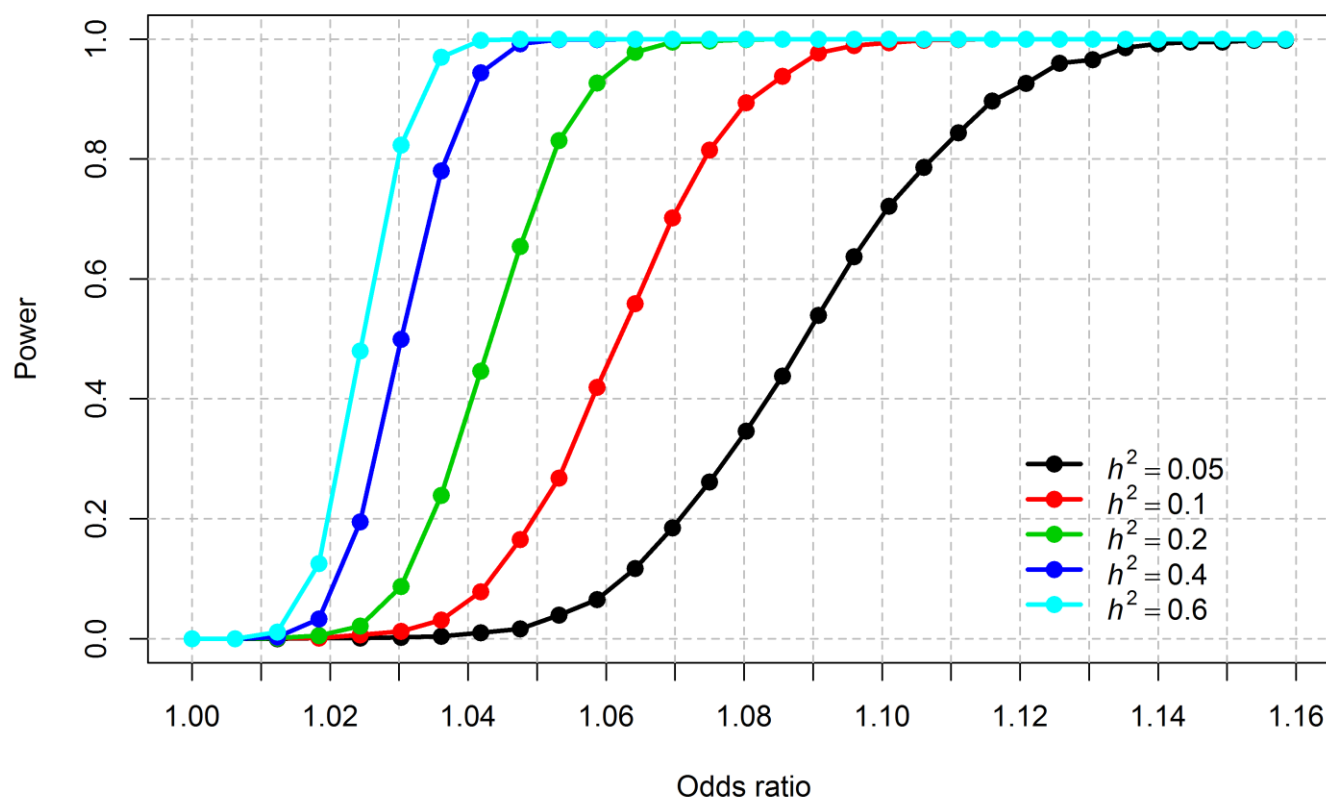
Supplementary Figure 6. Proliferation in breast cells using two independent siRNAs (related to Figure 2A). (A) 184A1, (B) MCF7 or (C) T47D cells were transfected with indicated siRNAs over seven days and phase-contrast images collected using an IncuCyte ZOOM. Each cell proliferation time-course was normalized to the baseline confluency and analyzed in GraphPad Prism. Corrected proliferation % = 100 +/- (relative proliferation in indicated siRNA - proliferation in control siRNA (consi))/knockdown efficiency. Error bars, SD ($N=2$). P -values were determined by one-way ANOVA followed by Dunnett's multiple comparisons test: * P -value < 0.05.



Supplementary Figure 7. Colony formation efficiency in MCF7 cells using two independent siRNAs (related to Fig 2B). MCF7 cells were transfected with indicated siRNAs, then reseeded after 16 hours for colony formation (CF) assays. At day 14, colonies were fixed with methanol, stained with crystal violet, scanned and batch analyzed by ImageJ. Corrected CF efficiency (CFE) % = 100 +/- (relative CFE in indicated siRNA - CFE in control siRNA (consi))/knockdown efficiency. Error bars, SD (*N*=2). *P*-values were determined by one-way ANOVA followed by Dunnett's multiple comparisons test: **P*-value < 0.05.



Supplementary Figure 8. Power calculation of the TWAS analysis. The simulation analysis is based on 122,977 cases and 105,974 controls. The gene expression was generated from the empirical distribution of predicted gene expression levels in the BCAC. Statistical power was calculated at $P < 5.82 \times 10^{-6}$ (the significance level used in main TWAS analyses) according to cis-heritability (h^2) which we aim to capture using gene expression prediction models (R^2). The figure shows results per one standard deviation increase (or decrease) in the gene expression based on 1000 replicates.



Supplementary Note

Acknowledgements for BCAC studies

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Consortia Membership

NBCS Collaborators

Prof. Em. Anne-Lise Børresen-Dale, PhD (Department of Cancer Genetics, Institute for Cancer Research, Oslo University Hospital Radiumhospitalet, Oslo, Norway), Dr. Kristine K. Sahlberg, PhD (Department of Research, Vestre Viken Hospital, Drammen, Norway and Department of Cancer Genetics, Institute for Cancer Research, Oslo University Hospital Radiumhospitalet, Oslo, Norway), Dr. Lars Ottestad, MD (Department of Cancer Genetics, Institute for Cancer Research, Oslo University Hospital Radiumhospitalet, Oslo, Norway), Prof. Em. Rolf Kåresen, MD (Institute of Clinical Medicine, Faculty of Medicine, University of Oslo, Oslo, Norway and Department of Breast- and Endocrine Surgery, Division of Surgery, Cancer and Transplantation Medicine, Oslo University Hospital Ullevål, Oslo, Norway), Dr. Anita Langerød, PhD (Department of Cancer Genetics, Institute for Cancer Research, Oslo University Hospital Radiumhospitalet, Oslo, Norway), Dr. Ellen Schlichting, MD (Section for breast- and endocrine surgery, Department of oncology, Oslo university hospital), Dr. Marit Muri Holmen, MD (Department of Radiology and Nuclear Medicine, Oslo University Hospital, Oslo, Norway), Prof. Toril Sauer, MD (Institute of Clinical Medicine, Faculty of Medicine, University of Oslo, Oslo, Norway and Department of Pathology at Akershus University hospital, Lørenskog, Norway), Dr. Vilde Haakensen, MD (Department of Cancer Genetics, Institute for Cancer Research, Oslo University Hospital Radiumhospitalet, Oslo, Norway), Dr. Olav Engebråten, MD (Institute of Clinical Medicine, Faculty of Medicine, University of Oslo, Oslo, Norway, Department of Tumor Biology, Institute for Cancer Research, Oslo University Hospital Radiumhospitalet, Oslo, Norway and Department of Oncology, Division of Surgery and Cancer and Transplantation Medicine, Oslo University Hospital Radiumhospitalet, Oslo, Norway), Prof. Bjørn Naume, MD (Department of Oncology, Oslo University Hospital, Oslo, Norway and Institute of Clinical Medicine, Faculty of Medicine, University of Oslo, Oslo, Norway), Dr. Alexander Fosså, MD (National Advisory Unit on Late Effects after Cancer Treatment, Department of Oncology, Oslo University Hospital, Oslo, Norway and Department of Oncology, Oslo University Hospital Ullevål, Oslo, Norway), Dr. Cecile E. Kiserud, MD (National Advisory Unit on Late Effects after Cancer Treatment, Department of Oncology, Oslo University Hospital, Oslo, Norway and Department of Oncology, Oslo University Hospital Ullevål, Oslo, Norway), Dr. Kristin V. Reinertsen, MD (National Advisory Unit on Late Effects after Cancer Treatment, Department of Oncology, Oslo University Hospital, Oslo, Norway and Department of Oncology, Oslo University Hospital Ullevål, Oslo, Norway), Assoc. Prof. Åslaug Helland, MD (Department of Cancer Genetics, Institute for Cancer Research, Oslo University Hospital Radiumhospitalet, Oslo, Norway), Dr. Margit Riis, MD (Department of Breast- and Endocrine Surgery, Division of Surgery, Cancer and Transplantation Medicine, Oslo University Hospital Ullevål, Oslo, Norway), Prof. Jürgen Geisler, MD (Institute of Clinical Medicine, Faculty of Medicine, University of Oslo, Oslo, Norway and Department of Oncology, Akershus University Hospital, Oslo, Norway), Prof. Per Eystein Lønning, MD (Section of Oncology, Institute of Medicine, University of Bergen and Department of Oncology, Haukeland University Hospital, Bergen, Norway), Grethe I. Grenaker Alnæs, M.Sc. (Department of Cancer Genetics, Institute for Cancer Research, Oslo University Hospital Radiumhospitalet, Oslo, Norway) and OSBREAC (Oslo Breast Cancer Research Consortium, Oslo University Hospital, Oslo, Norway)

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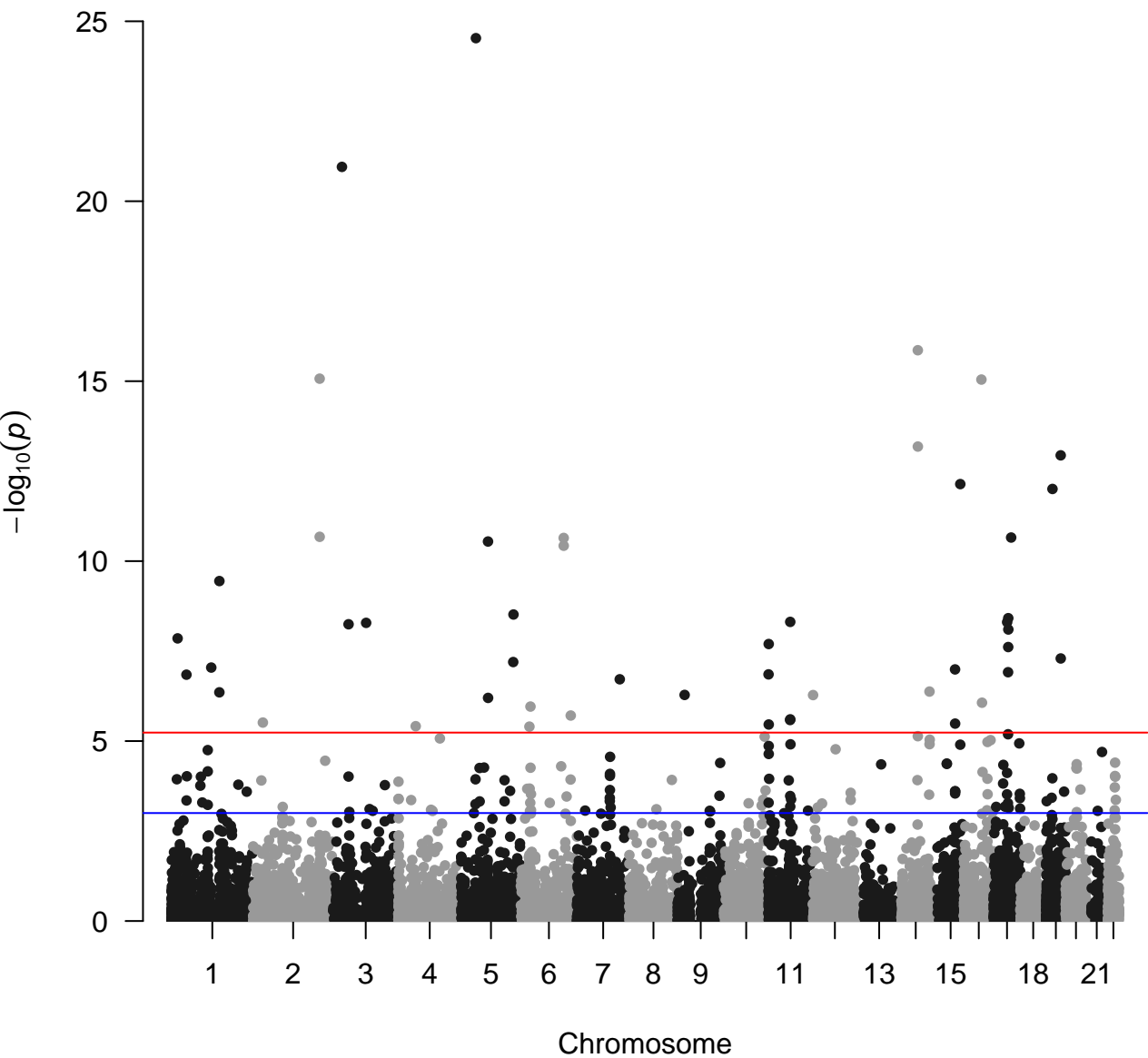
Australian Ovarian Cancer Study (AOCS Management Group):

D Bowtell (Peter MacCallum Cancer Centre), G Chenevix-Trench, A Green, P Webb (QIMR Berghofer), A deFazio (Westmead Institute for Cancer Research, WMI), D Gertig (Victorian Cervical Cytology Registry)

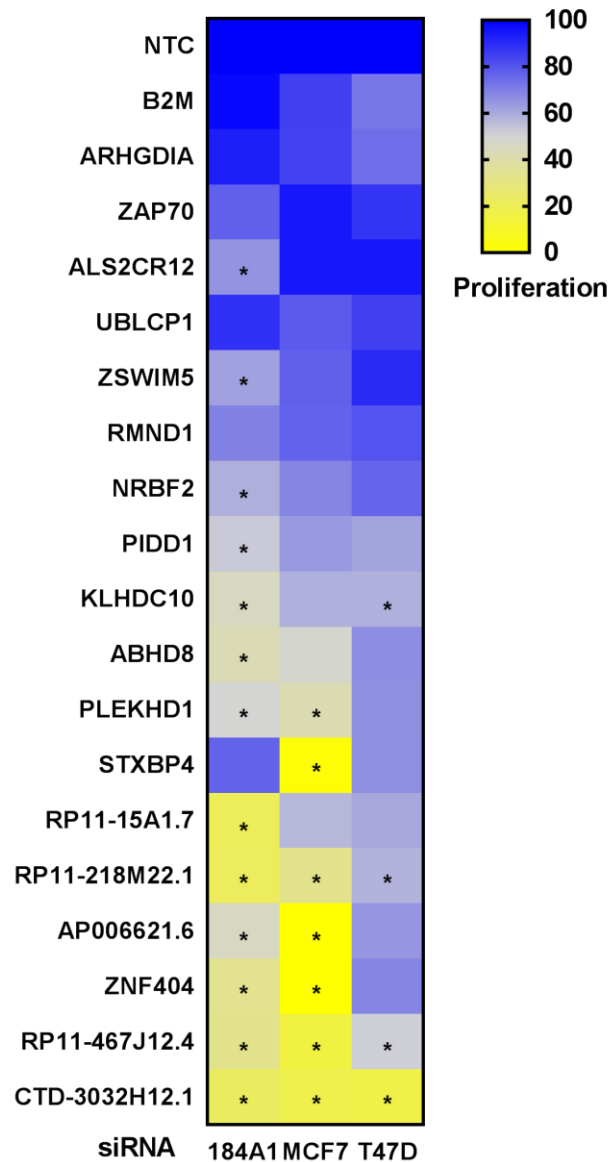
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