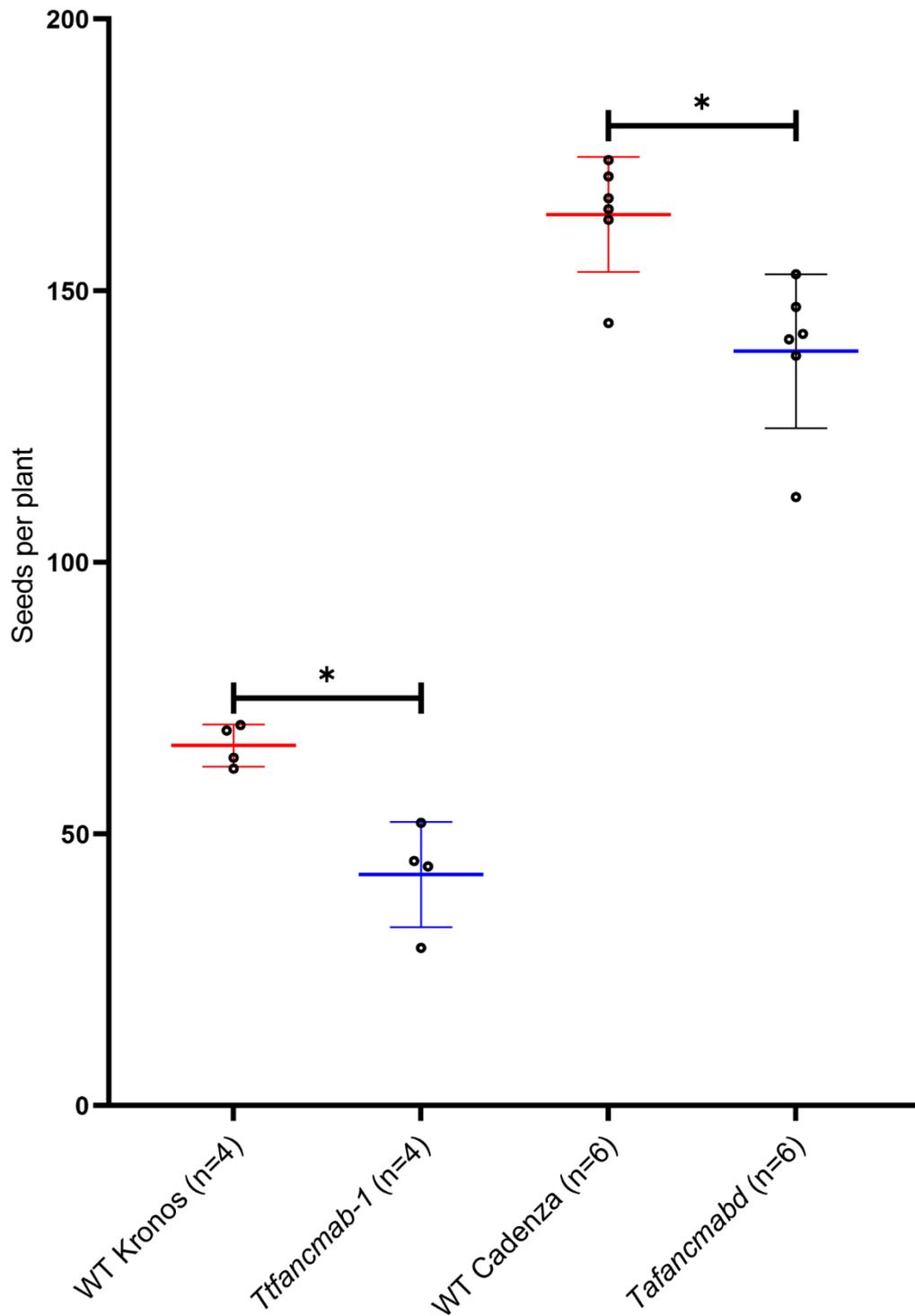
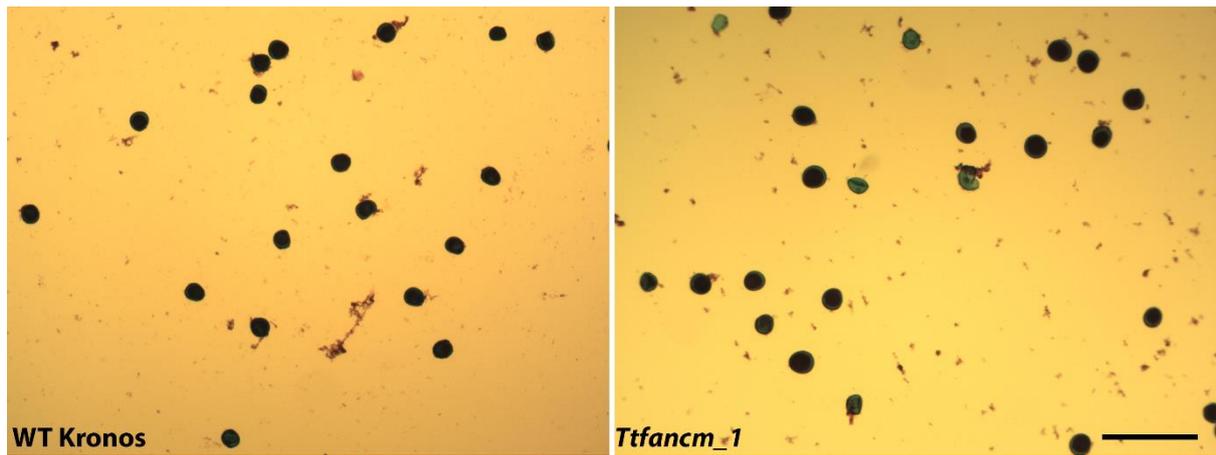


**FANCM promotes class I interfering crossovers and suppresses class II non-interfering crossovers in wheat meiosis**

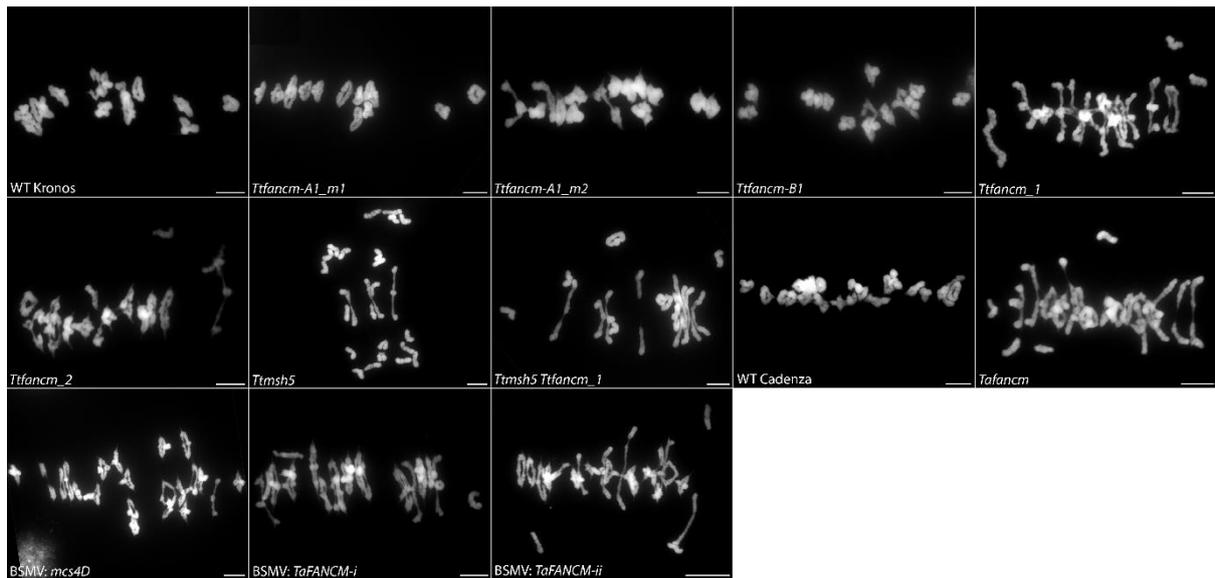
Desjardins *et al.*



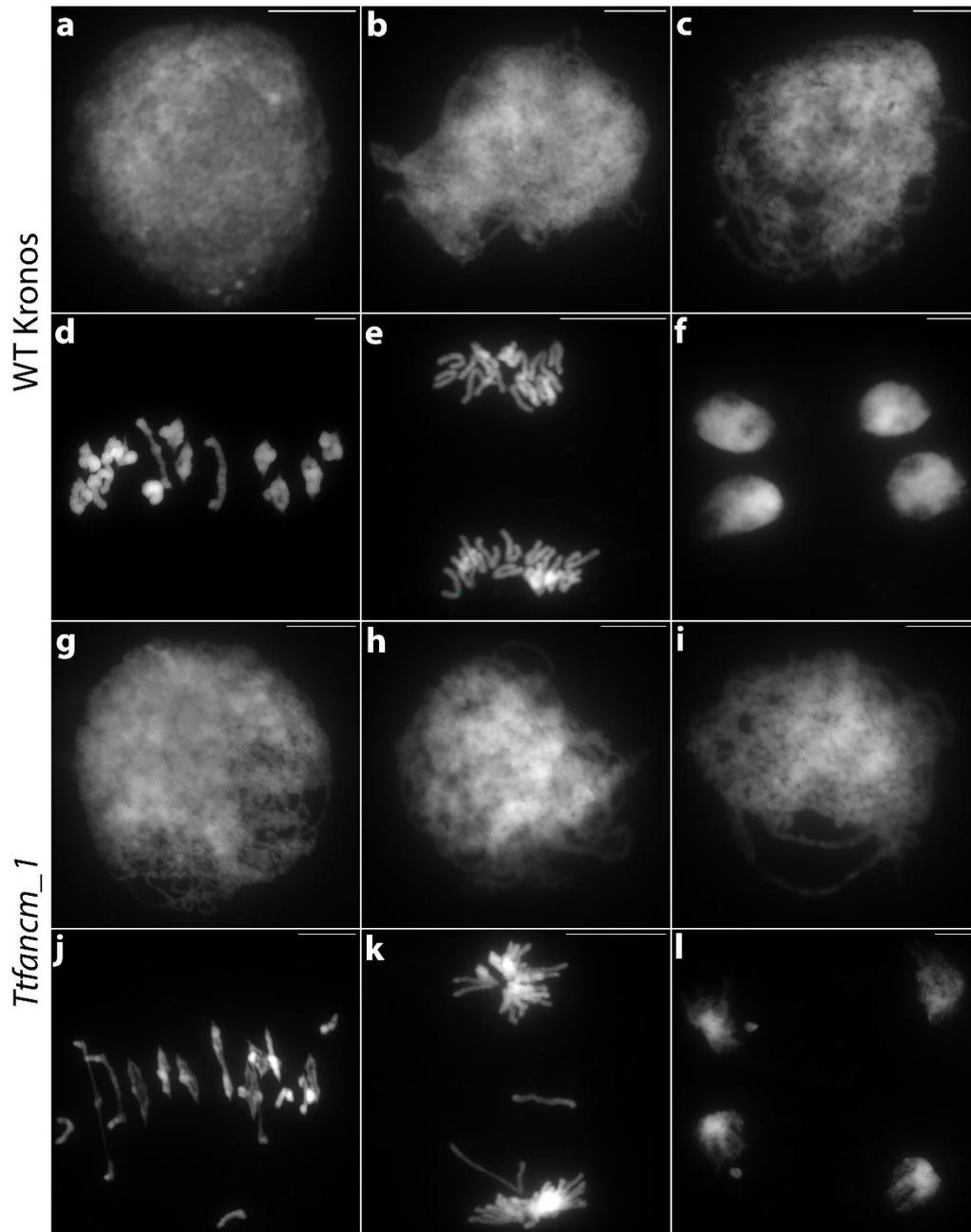
**Supplementary Figure 1. Reduced seed set in *fancm* null mutants.** Counts of total seed per plant with mean values  $\pm$  SD. The number of plants sampled for each line is shown in brackets. \* =  $p < 0.05$  (Mann-Whitney U Test). Exact  $p$ -values are as follows: 1 vs 2 ( $p = 0.03$ ) and 3 vs 4 ( $p = 0.013$ ). Source data are provided as a Source Data file.



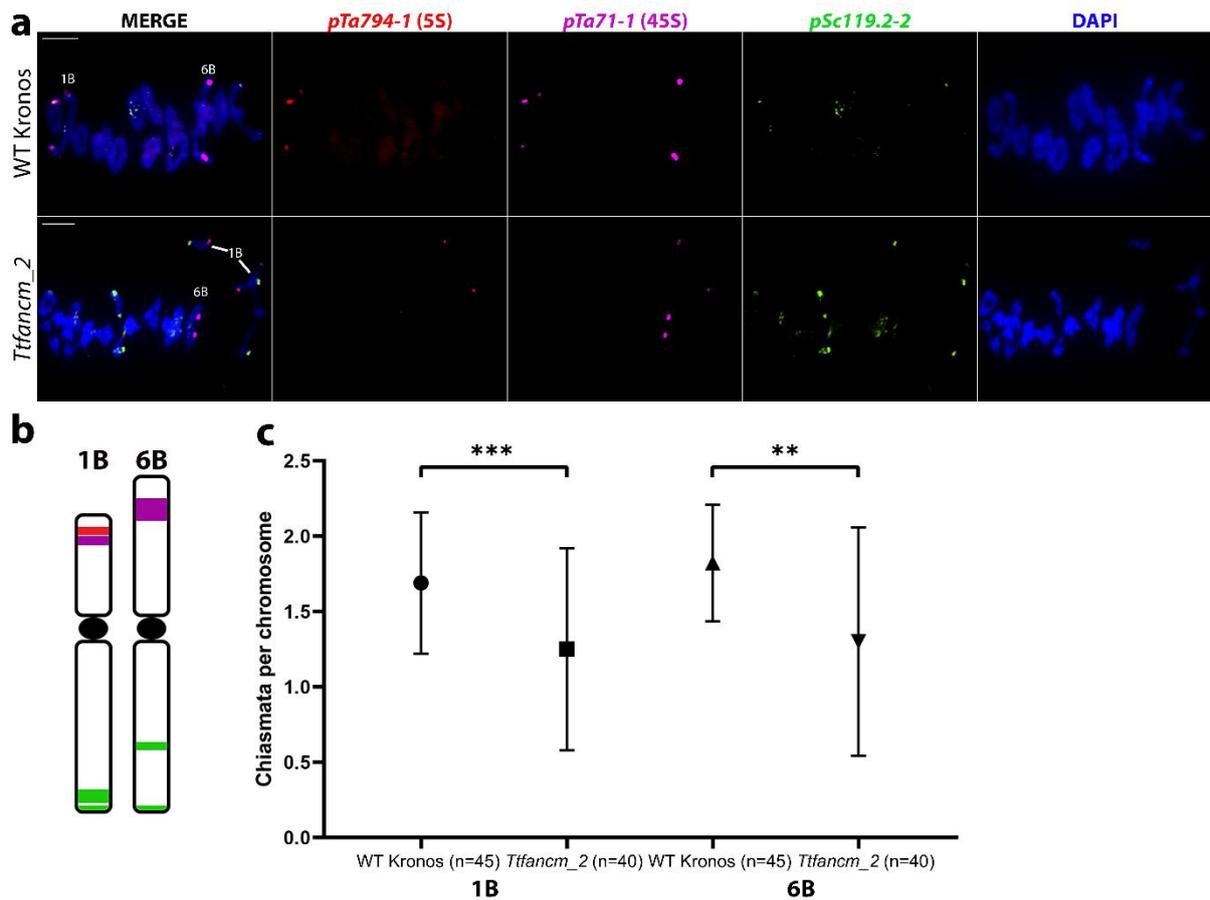
**Supplementary Figure 2. Reduced pollen viability in *Tfancm\_1* null mutant.** Alexander staining of fresh pollen grains. Representative pollen grains are shown from replicates, wild type ( $n = 2042$ ) and *Tfancm\_1* ( $n = 1994$ ). Scale bar = 200  $\mu\text{m}$ .



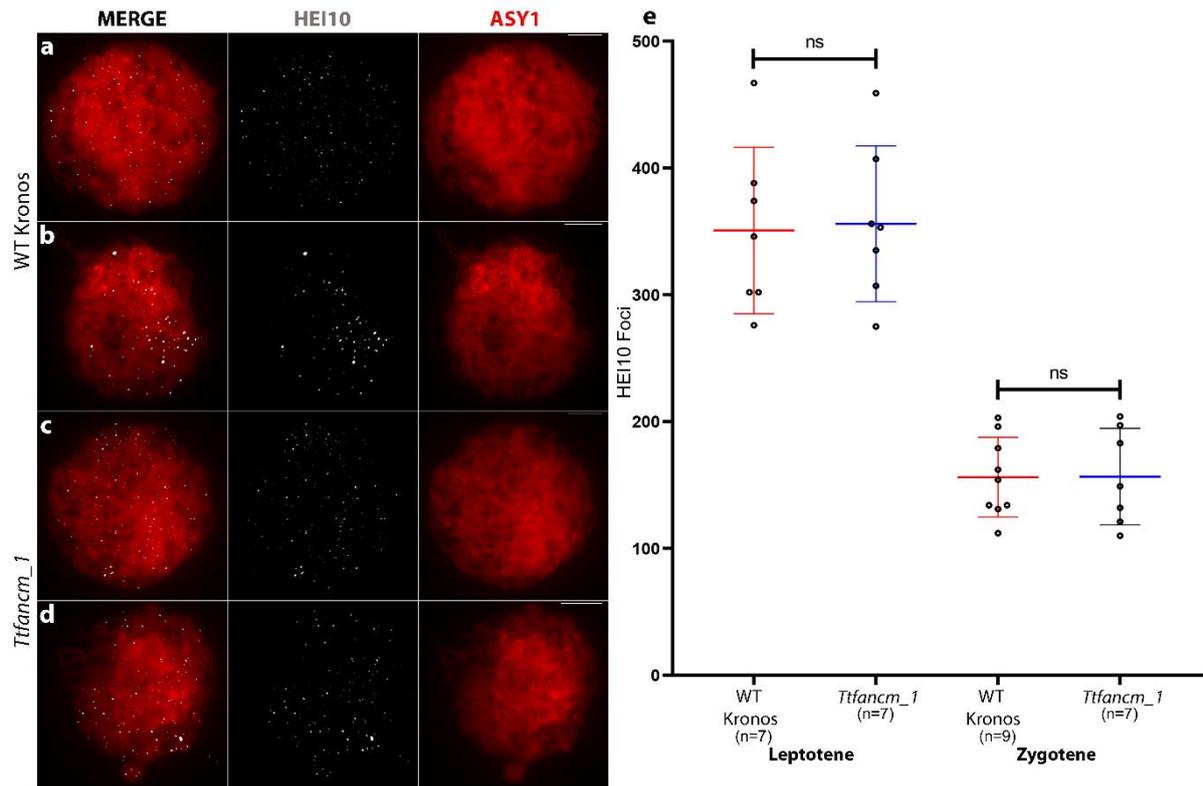
**Supplementary Figure 3. FANCM promotes formation of obligate chiasma.** DAPI-stained meiotic chromosome spreads at metaphase I. Representative micrographs are shown from replicates, WT Kronos ( $n = 60$ ), *Ttfancm-A1\_m1* ( $n = 64$ ), *Ttfancm-A1\_m2* ( $n = 60$ ), *Ttfancm-B1* ( $n = 44$ ), *Ttfancm\_1* ( $n = 104$ ), *Ttfancm\_2* ( $n = 30$ ), *Ttmsh5* ( $n = 194$ ), *Ttmsh5 Ttfancm\_1* ( $n = 86$ ), WT Cadenza ( $n = 76$ ), *Tafancm* ( $n = 81$ ), BSMV: *msc4D* ( $n = 50$ ), BSMV: *TaFANCM-i* ( $n = 33$ ) and BSMV: *TaFANCM-ii* ( $n = 43$ ). Scale bars = 10  $\mu$ m.



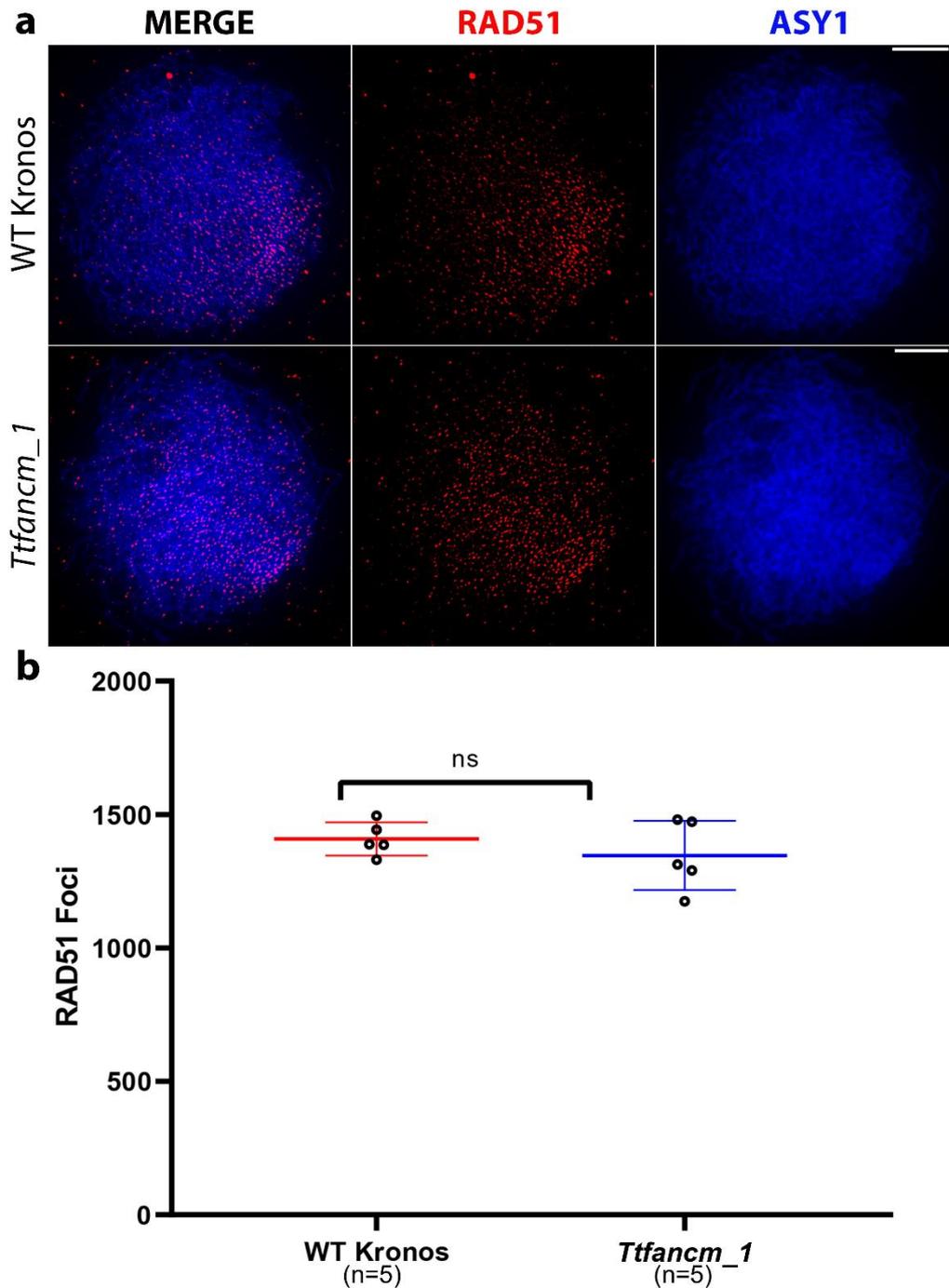
**Supplementary Figure 4. Univalents and mis-segregation observed in *Ttfancm\_1*, resulting in unbalanced gametes. a-l** Meiotic atlas of DAPI-stained chromosome spreads. Representative micrographs are shown from replicates, **a** ( $n = 10$ ), **b** ( $n = 10$ ), **c** ( $n = 10$ ), **d** ( $n = 60$ ), **e** ( $n = 40$ ), **f** ( $n = 10$ ), **g** ( $n = 10$ ), **h** ( $n = 10$ ), **i** ( $n = 10$ ), **j** ( $n = 60$ ), **k** ( $n = 40$ ), and **l** ( $n = 10$ ). **a** and **g** Leptotene. **b** and **h** Zygotene. **c** and **i** Pachytene. **d** and **j** Metaphase I. **e** and **k** Dyad. **f** and **l** Tetrad. Scale bars = 10  $\mu\text{m}$ .



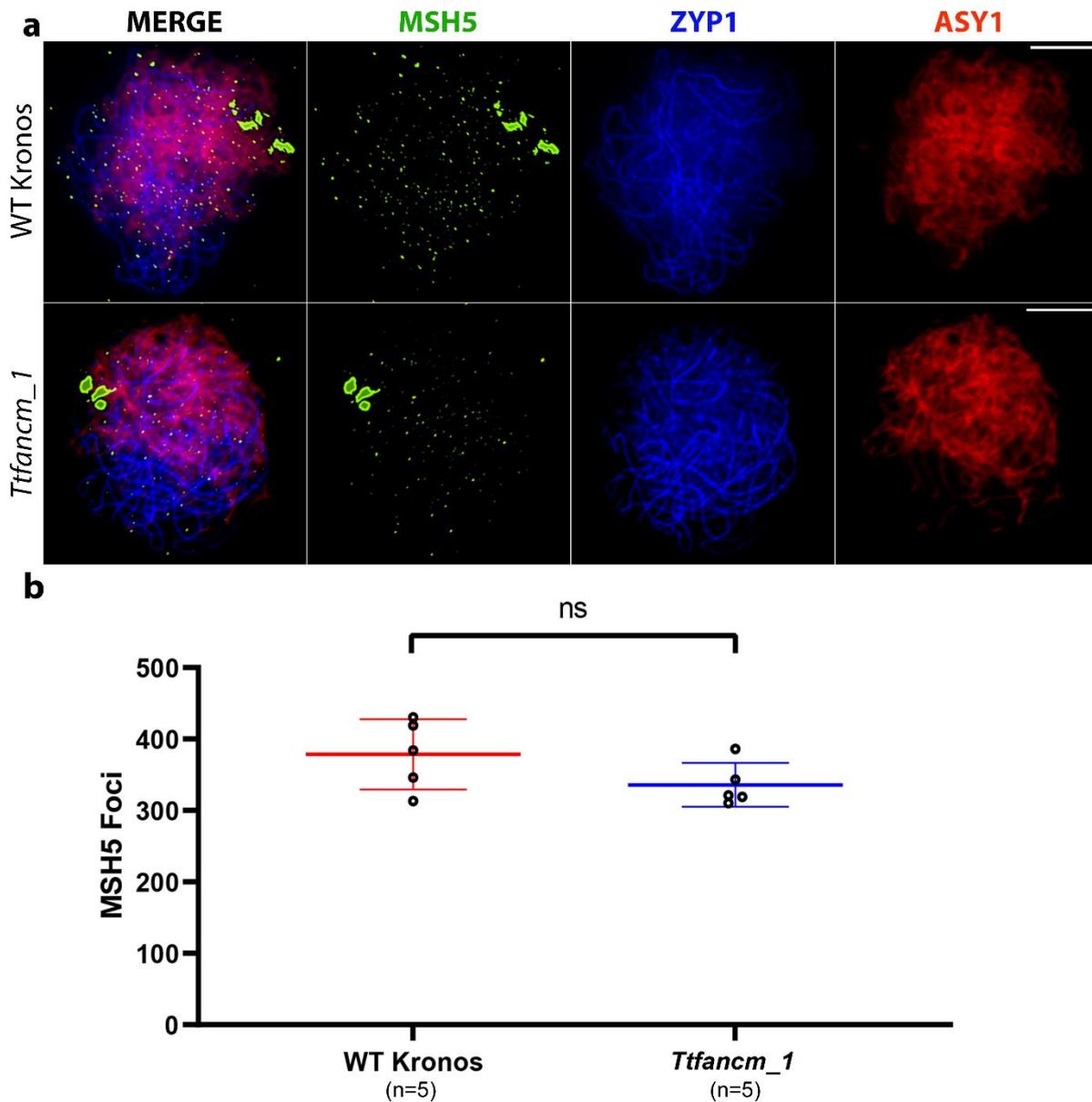
**Supplementary Figure 5. Chiasmata number reduced in chromosomes 1B and 6B in *Ttfancm\_2*.** **a** Fluorescence *in situ* hybridization (FISH) of pTa794-1 (5S), pTa71-1 (45S) and pSc119.2-2 probes on meiotic metaphase I chromosome spreads. Representative micrographs are shown from replicates, WT Kronos ( $n = 45$ ) and *Ttfancm\_2* ( $n = 40$ ). Scale bars = 10  $\mu$ m. **b** Hybridization signal patterns of chromosomes 1B and 6B. **c** Mean  $\pm$  SD chiasmata frequency per chromosome. The number of meicytes sampled for each line is shown in brackets. \*\* =  $p < 0.01$ . \*\*\* =  $p < 0.001$  (Mann-Whitney U Test). Exact p-values are as follows: 1 vs 2 ( $p = 0.002$ ), 3 vs 4 ( $p = 3.34 \times 10^{-4}$ ). Source data are provided as a Source Data file.



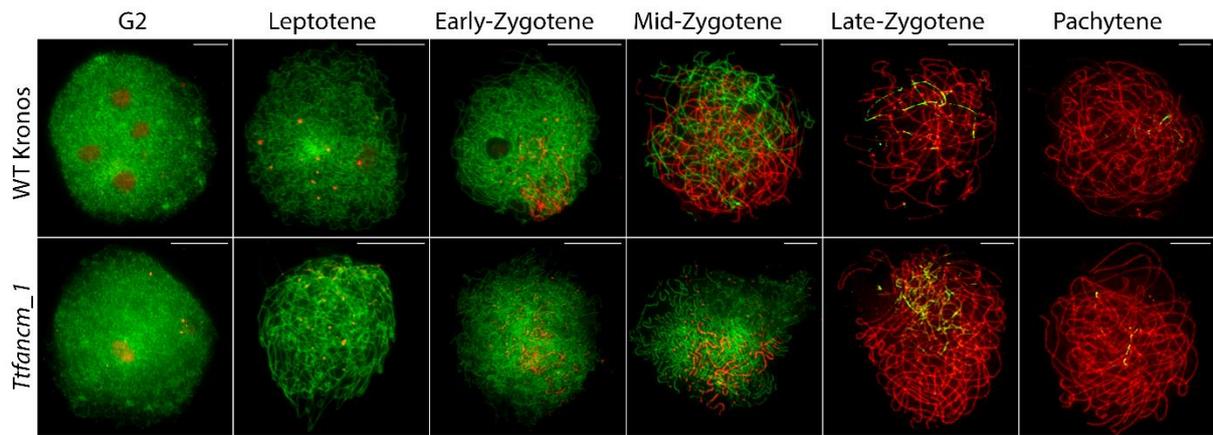
**Supplementary Figure 6. Class I crossover recombination protein HEI10 is unaffected in the *Ttfancm\_1* null mutant at early prophase I.** **a-e** Co-immunofluorescence of HEI10 (white) and ASY1 (red) on meiotic prophase I chromosome spreads. Representative micrographs are shown from replicates, **a** ( $n = 7$ ), **b** ( $n = 7$ ), **c** ( $n = 9$ ) and **d** ( $n = 7$ ). Leptotene (**a** and **c**) and Zygotene (**b** and **d**). Scale bars = 10  $\mu\text{m}$ . **e** Counts of HEI10 foci per cell with mean values  $\pm$  SD. The number of cells sampled for each line is shown in brackets. n.s. =  $p > 0.05$  (Mann-Whitney U Test). Exact  $p$ -values are as follows: 1 vs 2 ( $p = 1$ ), 3 vs 4 ( $p = 1$ ). Source data are provided as a Source Data file.



**Supplementary Figure 7. Early recombination protein RAD51 is unaffected in the *Ttfancm\_1* null mutant at leptotene.** **a** Co-immunofluorescence of RAD51 (red) and ASY1 (blue) on meiotic chromosome spreads at leptotene. Representative micrographs are shown from replicates, WT Kronos ( $n = 5$ ) and *Ttfancm\_1* ( $n = 5$ ). Scale bars = 10  $\mu$ m. **b** Counts of RAD51 foci per cell with mean values  $\pm$  SD. The number of cells sampled for each line is shown in brackets. n.s. =  $p > 0.05$  (Mann-Whitney U Test). Exact  $p$ -value is  $p = 0.403$ . Source data are provided as a Source Data file.

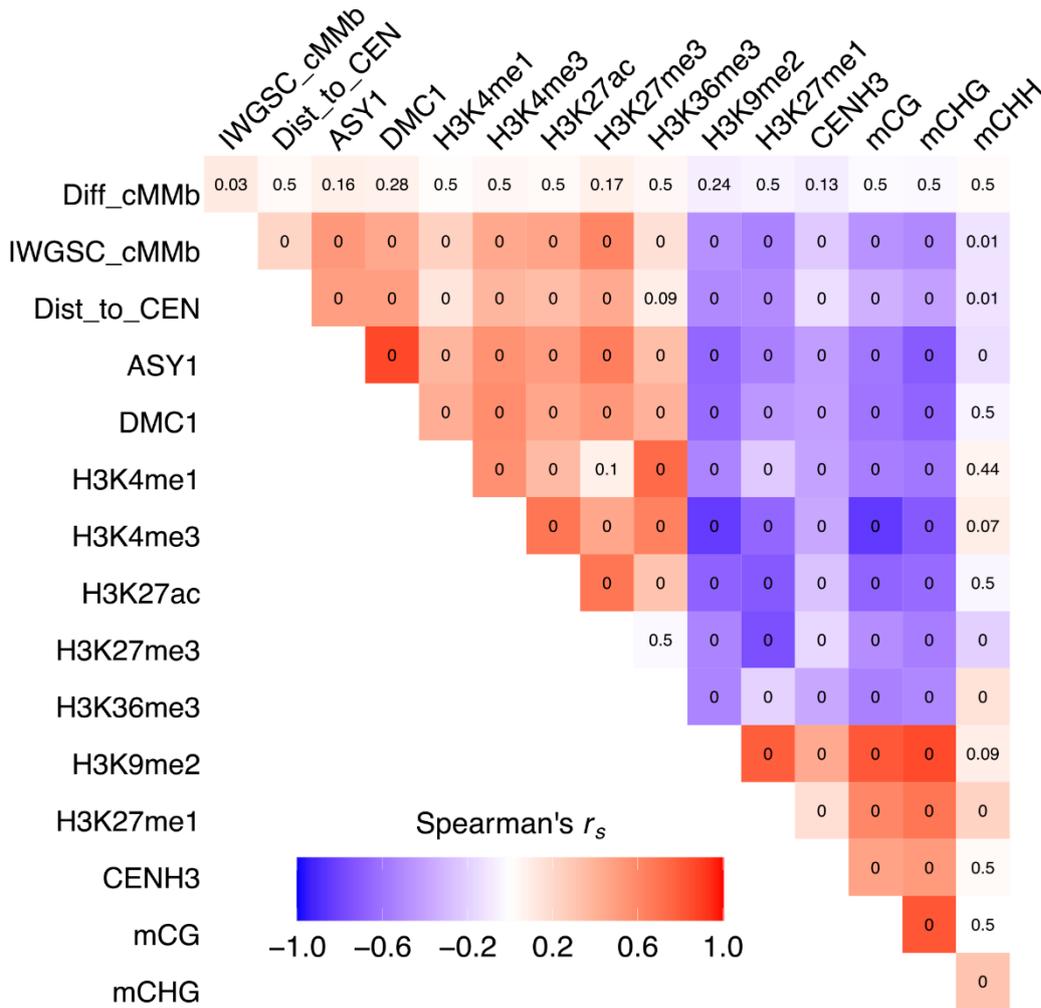


**Supplementary Figure 8. Class I crossover recombination protein MSH5 is unaffected in the *Ttfancm\_1* null mutant at mid-zygotene.** **a** Co-immunofluorescence of MSH5 (green), ZYP1 (blue) and ASY1 (red) on meiotic chromosome spreads at mid-zygotene. Representative micrographs are shown from replicates, WT Kronos ( $n = 5$ ) and *Ttfancm\_1* ( $n = 5$ ). Scale bars = 10  $\mu\text{m}$ . **b** Counts of MSH5 foci per cell with mean values  $\pm$  SD. The number of cells sampled for each line is shown in brackets. n.s. =  $p > 0.05$  (Mann-Whitney U Test). Exact  $p$ -values is  $p = 0.066$ . Source data are provided as a Source Data file.



**Supplementary Figure 9. Axis formation and synapsis are unaffected in the *Ttfanm\_1* null mutant.** Co-immunofluorescence of ASY1 (green) and ZYP1 (red) on meiotic prophase I chromosome spreads. Representative micrographs are shown from replicates: for WT Kronos G2 ( $n = 5$ ), leptotene ( $n = 53$ ), early-zygotene ( $n = 18$ ), mid-zygotene ( $n = 6$ ), late-zygotene ( $n = 9$ ), pachytene ( $n = 11$ ); for *Ttfanm\_1* G2 ( $n = 5$ ), leptotene ( $n = 20$ ), early-zygotene ( $n = 17$ ), mid-zygotene ( $n = 21$ ), late-zygotene ( $n = 3$ ), pachytene ( $n = 16$ ). Scale bars = 10  $\mu\text{m}$ .

Spearman's  $r_s$  for AxC mapped marker intervals



**Supplementary Figure 10. Spearman's rank-order correlation coefficients ( $r_s$ ) for the indicated parameter pairs computed within each genetic marker interval.** Correlation coefficients are indicated by cell colour.  $P$ -values for  $r_s$  correlation coefficients were standardised to represent those based on pairwise values across 100 marker intervals and are indicated within each cell. Included data sets are differential CO rate (*fanm* cM/Mb minus wild type cM/Mb; "Diff\_cMMb"), wild type CO rate derived from a Chinese Spring x Renan genetic map ("IWGSC\_cMMb"), ASY1, DMC1, H3K4me3, H3K9me2 and H3K27me1 ChIP-seq, H3K4me1 and H3K27ac ChIP-seq, H3K27me3 and H3K36me3 ChIP-seq, CENH3 ChIP-seq, whole-genome bisulfite sequencing-derived DNA methylation (mCG, mCHH and mCHG proportions), and the distance between the midpoint of each marker interval and the midpoint of previously defined centromeric coordinates ("Dist\_to\_CEN").