



Brain Cells Release Calreticulin That Attracts and Activates Microglia, and Inhibits Amyloid Beta Aggregation and Neurotoxicity

Kyle M. Reid[†], Emily J. A. Kitchener[†], Claire A. Butler, Tom O. J. Cockram and Guy C. Brown^{*}

Department of Biochemistry, University of Cambridge, Cambridge, United Kingdom

OPEN ACCESS

Edited by:

Carla Marie Cuda,
Northwestern University,
United States

Reviewed by:

Raymond B. Birge,
The State University of New Jersey,
United States
Bijay Parajuli,
University of Yamanashi, Japan

*Correspondence:

Guy C. Brown
gcb3@cam.ac.uk

[†]These authors share first authorship

Specialty section:

This article was submitted to
Inflammation,
a section of the journal
Frontiers in Immunology

Received: 24 January 2022

Accepted: 28 March 2022

Published: 20 April 2022

Citation:

Reid KM, Kitchener EJA, Butler CA,
Cockram TOJ and Brown GC (2022)
Brain Cells Release Calreticulin
That Attracts and Activates Microglia,
and Inhibits Amyloid Beta
Aggregation and Neurotoxicity.
Front. Immunol. 13:859686.
doi: 10.3389/fimmu.2022.859686

Calreticulin is a chaperone, normally found in the endoplasmic reticulum, but can be released by macrophages into the extracellular medium. It is also found in cerebrospinal fluid bound to amyloid beta (A β). We investigated whether brain cells release calreticulin, and whether extracellular calreticulin had any effects on microglia and neurons relevant to neuroinflammation and neurodegeneration. We found that microglia release nanomolar levels of calreticulin when inflammatory-activated with lipopolysaccharide, when endoplasmic reticulum stress was induced by tunicamycin, or when cell death was induced by staurosporine, and that neurons release calreticulin when crushed. Addition of nanomolar levels of extracellular calreticulin was found to chemoattract microglia, and activate microglia to release cytokines TNF- α , IL-6 and IL-1 β , as well as chemokine (C-C motif) ligand 2. Calreticulin blocked A β fibrillization and modified A β oligomerization, as measured by thioflavin T fluorescence and transmission electron microscopy. Extracellular calreticulin also altered microglial morphology and proliferation, and prevented A β -induced neuronal loss in primary neuron-glia cultures. Thus, calreticulin is released by microglia and neurons, and acts: as an alarmin to recruit and activate microglia, as an extracellular chaperone to prevent A β aggregation, and as a neuroprotectant against A β neurotoxicity.

Keywords: microglia, calreticulin, alarmin, brain, amyloid beta, chaperone, neurotoxicity

INTRODUCTION

Calreticulin is a highly-conserved chaperone that is constitutively expressed in virtually all mammalian cell-types (1). Calreticulin typically resides within the endoplasmic reticulum (ER), where it binds to exposed hydrophobic patches and sugar residues of nascent polypeptides, and functions as a molecular chaperone by refolding misfolded proteins (2). During conditions of apoptosis or ER stress, calreticulin can be translocated to the cell surface (3, 4), where it acts as an 'eat-me' signal to local phagocytes, such as macrophages, inducing macrophage phagocytosis of calreticulin-exposed cells by activating the LDL receptor-related protein 1 (LRP1) receptor on phagocytes (3, 5–8). Calreticulin on the surface of phagocytes can also act as a phagocytic co-

receptor together with LRP1, enabling LRP1 to be activated by SP-A and SP-D (9) and/or C1q (10, 11). However, calreticulin is a soluble protein, and therefore, once released onto the cell surface, has the potential to be released extracellularly. Indeed, calreticulin has been shown to be secreted by inflammatory-activated macrophages into the extracellular space, where it can opsonise target cells for phagocytosis (12, 13).

Alarmins are endogenous molecules released by necrotic cells and inflamed immune cells, to recruit and activate immune cells, in order to restore homeostasis of damaged tissues (14). Alarmins are therefore also damage-associated molecular patterns (DAMPs), i.e., molecules (such as nucleotides and HMGB1) released by tissue damage to induce an inflammatory response in immune cells to reduce damage. DAMPs were named in analogy with pathogen-associated molecular patterns (PAMP), which are exogenous molecules (such as lipopolysaccharide) released by pathogens, that induce an inflammatory response to combat pathogens. Extracellular calreticulin may also act as a DAMP to activate immune cells (15), although contradictory findings (16) make this unclear. It is not known whether extracellular calreticulin can recruit and activate microglia, so we aimed to test this here.

Extracellular chaperones (such as clusterin) are proteins in the extracellular space that help fold other proteins into a functional or non-pathogenic form (17, 18). This may be particularly important in Alzheimer's disease, where the extracellular space of the brain becomes clogged up by the aggregation of amyloid beta ($A\beta$) into extracellular plaques, and extracellular chaperones may potentially prevent this (19). Activated microglia release calreticulin into the culture medium (13), and calreticulin is bound to $A\beta$ in human cerebrospinal fluid (20). Thus, it is possible that calreticulin acts as an extracellular chaperone to keep $A\beta$ in solution or promote its degradation (20, 21). We tested here whether calreticulin could prevent $A\beta$ aggregation into oligomers and fibrils, and if so whether it can prevent $A\beta$ -induced neurotoxicity.

MATERIALS AND METHODS

Materials

All cell culture reagents were from Invitrogen (Paisley, UK), unless otherwise indicated. Culture treatments were procured as follows: staurosporine (from *Streptomyces* sp.) and recombinant human calreticulin were from Abcam (Cambridge, UK); peptide synthesized human amyloid-beta (1-42) was purchased from Anaspec (Fremont, CA); adenosine 5'-triphosphate (ATP) disodium salt hydrate, lipopolysaccharide (LPS; from *Salmonella Enterica* serotype typhimurium), polymyxin B sulfate salts and tunicamycin (from *Streptomyces* sp.) were from Sigma-Aldrich (St Louis, MO, USA); recombinant mouse LRPAP-1 protein was from R&D Systems (Minneapolis, USA).

Cell Culture and Treatments

The immortalized cell lines BV-2 (ECACC Cat# 0356, RRID: CVCL_0182) and CHME3 (ATCC Cat# CRL-3304, RRID:

CVCL_I176) were maintained as previously described (22, 23). Neither cell line is listed as a commonly misidentified cell line by the International Cell Line Authentication Committee. Primary microglial and mixed neuron-glia cultures were respectively prepared from the cortex and cerebellum of 3-5 day old Wistar rats (Charles River, RRID: RGD_2312511), following procedures described elsewhere (24, 25). All animal experiments were approved by the Cambridge University Local Research Ethics Committee and undertaken in accordance with the UK Animals (Scientific Procedures) Act (1986).

Cells were treated as follows: calreticulin was used at 1 nM, 10 nM, 100 nM or 170 nM for 20 hours; 10 nM, 17 nM or 50 nM for 24 hours; 10 nM or 50 nM for 48 hours; and 2.5 nM, 10 nM or 50 nM for 72 hours as indicated. Monomeric $A\beta$ was used at 250 nM over 72 hours. LPS was added at 100 ng/mL over 20 or 24 hours where indicated. Calreticulin or LPS were pre-treated with 10 U/mL polymyxin B for 1 hour where indicated. Polymyxin B was used at 10 U/mL for 20 hours. LRPAP-1 was used at 100 nM for 20 hours. Tunicamycin was used at 0.2 and 2 μ g/mL for 24 hours. Staurosporine was used at 10 nM and 100 nM for 24 hours, and at 1000 nM for 6 or 24 hours as specified. ATP was added at 10 μ M for 24 hours.

Cell Viability

Cell viability, defined as the percentage of non-necrotic cells, was measured at indicated endpoints by differential dye uptake of propidium iodide (identifying necrotic cells) and Hoechst 33342 (identifying all cells) using a fluorescent microscope (EVOS M5000). Alexa Fluor 488-tagged isolectin B4 was used to identify microglia. Apoptotic cells were recognized as non-necrotic cells with nuclear condensation (identified with Hoechst 33342). For pure microglial cultures, the entire well was imaged, and the number of cells quantified using QuPath (version 0.3.0). Two wells were quantified per experiment, with the number of experimental replicates indicated in the figure legends. For neuronal-glia co-cultures, four microscopic fields were quantified per well, with two wells per experiment, and three experimental replicates.

Transwell Migration Assay

BV-2 migration was measured by transwell (Boyden) chamber assay using COSTAR 24-well plate 8.0 μ m pore membrane inserts (Corning). Briefly, 2×10^4 BV-2 cells were seeded into serum-free DMEM on the top chamber membrane. In the bottom chamber, serum-free DMEM, supplemented with calreticulin or ATP, was added. Migration was assessed across 24 hours under incubating conditions (37°C, 5% CO₂). At experimental endpoints, residual microglia found on top of the membrane were removed using a cotton bud. Microglia that had migrated to the underside of the membrane were fixed in 4% paraformaldehyde, washed in PBS, and stained with Hoechst 33342. Migrated microglia were imaged using the EVOS M5000 (4x Magnification, DAPI Channel).

Measurement of Calreticulin Release

Calreticulin release from BV-2 microglia and mixed neuron-glia were measured by quantitative western blot. Briefly, 5×10^4 BV-2 cells were cultured in serum-free DMEM and treated

accordingly. Neuron-glia culture medium was replaced with PBS supplemented with proteinase inhibitors, and cultures were compressed with the flat end of a syringe plunger for one minute to rupture the cells. Proteins from the conditioned mediums of treated cultures were resolved on NuPAGE 4-12% Bis-Tris gels (Invitrogen) and transferred onto PVDF membranes. Blots were probed with a polyclonal antibody to calreticulin (1:500, EnzoLifeSciences) followed by IRDye® 800CW donkey anti-rabbit IgG secondary antibody (1:5000, Licor). Antibody binding was detected using the LI-COR Odyssey® CLX. Quantification of calreticulin loaded was attained by comparing band intensity values for each condition, quantified by Licor Image Studio™, to a standard curve created using 1, 2.5 and 5 ng of recombinant calreticulin loaded within each gel.

Calreticulin release from BV-2 microglia treated with LPS was measured by ELISA. Briefly, 5×10^4 BV-2 cells were cultured in low-serum DMEM. After 24 hours, supernatants were assessed for calreticulin using a calreticulin ELISA (Abbea), following the supplied protocol. Absorbances were read at 450nm by the FLUOstar OPTIMA spectrophotometer (BMG Labtech, Ortenberg, Germany).

Measurement of Inflammatory Molecules

Release of inflammatory molecules from BV-2 microglia and primary rat microglia were measured by ELISA. Briefly, 5×10^4 BV-2 cells were seeded in serum-free DMEM, 5×10^4 primary rat microglia were seeded in appropriate medium, and microglia were treated accordingly for 20 hours. Following treatments, supernatants were removed, and protein detection was achieved using mouse ELISAs for TNF- α , IL-6 and CCL2, and a rat ELISA for TNF- α , as per the manufacturer's instructions (all BioLegend). Absorbances were read at 450nm by a spectrophotometer.

Release of inflammatory molecules from CHME3 microglia were assessed using a custom pre-coated meso scale discovery plex human pro-inflammatory panel 2 (Meso Scale Discovery). Briefly, 3×10^4 CHME3 cells were seeded in serum-free DMEM and treated with calreticulin. After 20 hours, supernatants were removed, and protein detection was achieved by electrochemiluminescence for human TNF- α , IL-6 and IL-1 β as per the manufacturer's instructions.

Measurement of Endotoxin

Endotoxin (LPS) levels were measured from recombinant calreticulin preparations using the Pierce™ limulus amoebocyte lysate (LAL) chromogenic endotoxin quantification kit (Thermo Fisher Scientific), as per manufacturer's protocol. Endotoxin levels are expressed as endotoxin unit per milliliter (EU/mL).

Proliferation Assay

2,000 BV-2 cells per well were seeded in DMEM and treated with $\pm$ calreticulin. After 24, 48 and 72-hour periods, microglial count and viability were measured through differential dye uptake of Hoechst 33342 and propidium iodide by fluorescence microscopy.

Amyloid- β Preparation

Peptide synthesized human amyloid-beta 1-42 was prepared as previously described (26). Briefly, A β was dissolved in

1,1,1,3,3,3-hexafluoroisopropanol (HFIP, Sigma) and dried under a stream of nitrogen. Prior to the aggregation assay, A β was resolubilized by DMSO (Sigma) and resuspended in DMEM.

In Vitro Amyloid- β Aggregation Assay

A β aggregation was measured by thioflavin T fluorescence as previously described (27). Briefly, thioflavin T (Sigma) and A β (10 μ M final concentration for both) were added to DMEM $\pm$ 0.1 μ M or 1 μ M calreticulin. A β was let to aggregate at 31.5°C, with orbital shaking for 10 seconds, every 10 minutes and fluorescence (440 nm absorbance, 480 nm emission) was measured at 10 minute intervals within the plate reader. At the end of the assay, aggregates in wells were imaged with a fluorescence microscope (DMI6000; Leica) using 480/40 nm excitation and 527/30 nm emission filters with a 40x objective.

Preparation of Amyloid- β Oligomers for Transmission Electron Microscopy

To generate oligomers, monomeric A β (10 μ M) was incubated $\pm$ 1 μ M calreticulin [18 h at 4°C using a thermocycler (Thermo Fisher Scientific)]. These oligomeric samples (10 μ L) were applied to carbonate coated grids for 1 min and negatively stained with 1% uranyl acetate for 1 minute. Micrographs were obtained on a Tecnai G2 transmission electron microscope.

Statistical Analysis

Statistical analyses were conducted using GraphPad Prism v9. Statistical differences between two groups were analyzed by unpaired and paired t-tests where indicated. Statistical differences between three or more groups were analyzed by one-way repeated measures or mixed model ANOVA followed by Dunnett's or Šidák multiple comparisons *post-hoc* test. Normality of acquired data was tested by Shapiro-Wilk test. Error bars represent the standard error of the mean of experiments (SEM). p-values refer to the probability of the null hypothesis that the means do not differ. $p < 0.05$ was considered significant, and $p \geq 0.05$ not significant.

RESULTS

Microglia and Neurons Release Calreticulin

We have previously shown that primary mouse microglia release calreticulin into the medium when inflammatory activated with lipopolysaccharide, LPS (13), and we confirmed here that BV-2 microglia release calreticulin in response to LPS (Supplementary Figure 1A), without inducing cell death (Supplementary Figure 1B). We then tested whether BV-2 microglia would release calreticulin, measured by western blots of cell culture supernatants, in various other conditions. Tunicamycin, which induces ER stress by inhibiting protein glycosylation (28), induced significant calreticulin release (Figures 1A, B), without inducing significant cell death (Supplementary Figure 2). Staurosporine, which induces apoptosis by inhibiting protein phosphatases (29),

killed the cells (**Supplementary Figure 2**), and induced release of calreticulin into the medium (**Figures 1A, B**).

We then tested whether necrotic neurons would release calreticulin, and found that crushed cerebellar granule neurons significantly released calreticulin into the culture medium, measured by western blots (**Figures 1C, D**). The reason for crushing the neurons was to test whether calreticulin would be released into the medium when rupturing their plasma membrane. Thus, dying microglia and neurons, and ER-stressed or inflammatory-activated microglia, can release calreticulin, resulting in nanomolar levels in the extracellular culture medium.

Calreticulin Activates Microglia

Calreticulin is known to be present in human cerebrospinal fluid, but the levels are unknown (20). Having found that microglia and neurons can release nanomolar levels of calreticulin, we tested whether the addition of nanomolar levels of calreticulin can activate microglia to release pro-inflammatory cytokines. Human, recombinant calreticulin was used here and in all subsequent experiments. First, we added 0 nM, 1 nM, 10 nM or 100 nM concentrations of extracellular calreticulin to mouse BV-2 microglia and measured the amount of extracellular TNF- α (tumor necrosis factor alpha) in the medium 20 hours later by ELISA, and compared the release to that induced by the positive control, LPS. Untreated BV-2 microglia released no detectable TNF- α , while 1 nM, 10 nM and 100 nM calreticulin induced the release of 0.05, 0.12 and 0.25 ng/ml TNF- α , and LPS induced the release of 0.35 ng/ml TNF- α (**Figure 2A**). Thus, nanomolar concentrations of calreticulin can induce TNF- α release from BV-2 microglia, at a similar level to that induced by LPS.

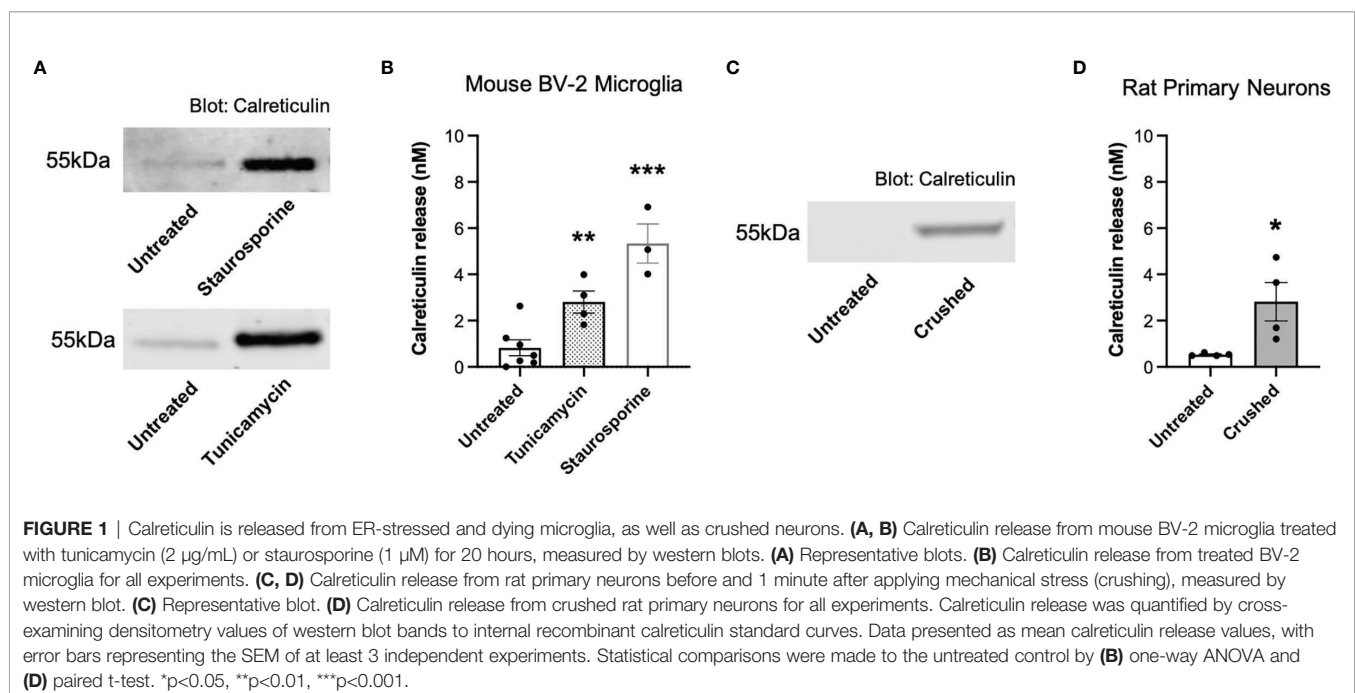
As mouse microglia may differ from human microglia (30), we tested whether calreticulin can activate human microglia to release cytokines, using a human microglial cell line, CHME3

[also known as HMC3 (31)]. By meso scale discovery, we measured TNF- α , IL-6 (interleukin 6) and IL-1 β release from CHME3 microglia treated for 20 hours with calreticulin. Calreticulin induced significant release of TNF- α , IL-6 and IL-1 β from this human microglial cell line (**Figures 2B–D**).

As the calreticulin we used was recombinant, and recombinant proteins are potentially contaminated with LPS (32), we tested whether Polymyxin B, which binds and inactivates LPS (33), could prevent the release of TNF- α induced by calreticulin in BV-2 microglia. Polymyxin B prevented the TNF- α release from BV-2 microglia induced by LPS, but had no effect on the TNF- α release induced by calreticulin (**Figure 3A**). This suggests there was little or no LPS contamination of the calreticulin used. This was confirmed by limulus amebocyte lysate (LAL) assay, which measured LPS levels in the calreticulin preparation to be negligible (0.22 ± 0.01 EU/mL (<1 ng of LPS)).

One potential means by which extracellular calreticulin might activate microglia is *via* the receptor LRP1, as this receptor mediates some actions of extracellular calreticulin (34, 35). LRP1 is inhibited by LRPAP1 (LRP-associated protein (36));, so we tested whether added LRPAP1 could prevent calreticulin inducing TNF- α release from BV-2 microglia, but found that it had no effect on this release (**Figure 3A**).

In order to test whether calreticulin can induce the release of other cytokines or chemokines from BV-2 microglia, we added calreticulin to BV-2 microglia and measured IL-6 and CCL2 in the extracellular medium 20 hours later by ELISA. IL-6 is a pro-inflammatory cytokine, and CCL2 (chemokine (C-C motif) ligand 2) is a chemokine, also known as monocyte chemoattractant protein 1 (MCP1). Calreticulin induced the release of both IL-6 and CCL2, and this was not prevented by Polymyxin B or LRPAP1 (**Figures 3B, C**). Thus, calreticulin induces the release of multiple pro-inflammatory cytokines and a



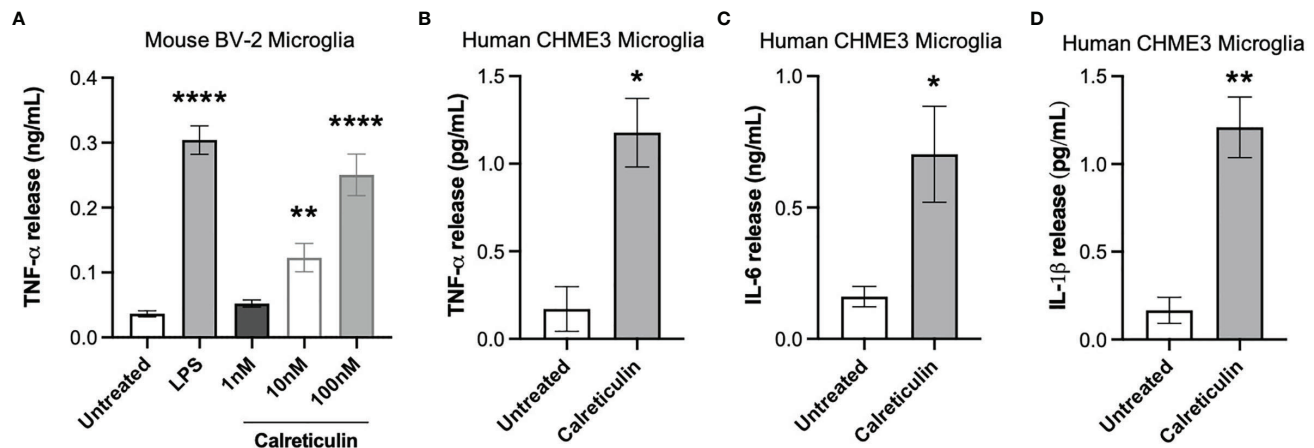


FIGURE 2 | Calreticulin induces cytokine release from mouse and human microglia. **(A)** BV-2 microglia were treated with lipopolysaccharide (LPS, 100 ng/mL) or human, recombinant calreticulin (1 nM, 10 nM or 100 nM) for 20 hours. Supernatants from treated BV-2 were assessed for TNF- α by ELISA. **(B–D)** CHME3 microglia were treated for 20 hours with 170 nM calreticulin (human, recombinant). Supernatants from treated CHME3 were assessed for **(B)** TNF- α , **(C)** IL-6 and **(D)** IL-1 β by meso scale discovery. Data presented as mean cytokine release values, with error bars representing the SEM of 3 independent experiments. Statistical comparisons were made to the untreated control by **(A)** one-way ANOVA and **(B–D)** unpaired t-test. * $p < 0.05$, ** $p < 0.01$, **** $p < 0.0001$.

chemokine from BV-2 microglia, and this does not appear to be due to LPS contamination or activation of LRP1.

As BV-2 are a cell line, which may diverge from primary cells as a result of transformation and/or mutations (37), we also tested whether calreticulin could induce TNF- α release from primary microglia isolated from rat brain. Calreticulin did induce TNF- α release from primary microglia, and this was not prevented by Polymyxin B or LRPAP1 (**Figure 3D**), just as BV-2 microglia (**Figure 3A**).

We conclude that nanomolar extracellular calreticulin can activate microglia (mouse, rat and human, primary and cell lines), as measured by the release of pro-inflammatory cytokine and chemokines, and this is not mediated by LPS or LRP1.

Calreticulin Chemoattracts Microglia

Having found that damaged or activated brain cells release calreticulin, and that added calreticulin can activate microglia, we went on to test whether extracellular calreticulin can do anything else relevant to neuroinflammation and neurodegeneration, in particular whether calreticulin can act as an alarmin, extracellular chaperone or neuroprotectant. If extracellular calreticulin acts as an alarmin, then it should chemoattract microglia. We tested whether extracellular calreticulin can chemoattract microglia by placing BV-2 microglia on top of a transwell (Boyden) chamber and culture medium $\pm$ calreticulin or ATP in the lower chamber. ATP is known to chemoattract microglia (38), and acted as a positive control here. After 24 hours, the number of cells that had migrated through the transwell membrane (with 8 μ m pores) were counted, and both calreticulin and ATP were found to significantly stimulate microglial migration (**Figures 4A, B**). Thus, extracellular calreticulin at nanomolar concentrations can chemoattract microglia.

Calreticulin Reduces Microglial Proliferation

If extracellular calreticulin activates microglia, then we might expect it to affect microglial proliferation. Calreticulin can alter the proliferation of a variety of cell types, and is implicated in tumor growth (39), tumor suppression (40, 41), and wound healing (42). We therefore tested whether calreticulin affected the proliferation of microglia. To do this, we seeded the same number of BV-2 cells in culture, to which we added either 0, 10 or 50 nM calreticulin, and then counted the numbers of cells after 24, 48 and 72 hours. Across all timepoints, calreticulin dose-dependently reduced the proliferation of microglia (**Figures 5A–C**), without inducing cell death (**Supplementary Figure 3**). 10 nM calreticulin significantly inhibited proliferation at all times, and 50 nM calreticulin inhibited proliferation more, but, note that even in the presence of 50 nM calreticulin, the BV-2 microglia still proliferated substantially (**Figure 5**). Thus, exogenous calreticulin mildly reduces microglial proliferation.

Calreticulin Alters Aggregation of Amyloid Beta (A β)

Calreticulin is an intracellular chaperone, which can bind A β (21), but it is not known whether calreticulin can act as extracellular chaperone for A β , which would be particularly relevant for Alzheimer's disease. We investigated whether calreticulin affected A β aggregation using an *in vitro* fibrillization assay, incubating pure monomeric A β with thioflavin T, which becomes fluorescent when bound to β -sheet structures within A β aggregates (43). In the absence of calreticulin, A β (10 μ M) fibrillized with standard lag-phase kinetics at 31.5°C (**Figure 6A**). In the presence of 1 μ M calreticulin, fibrillization of A β was almost completely prevented (**Figure 6A**). While in the presence of 0.1 μ M

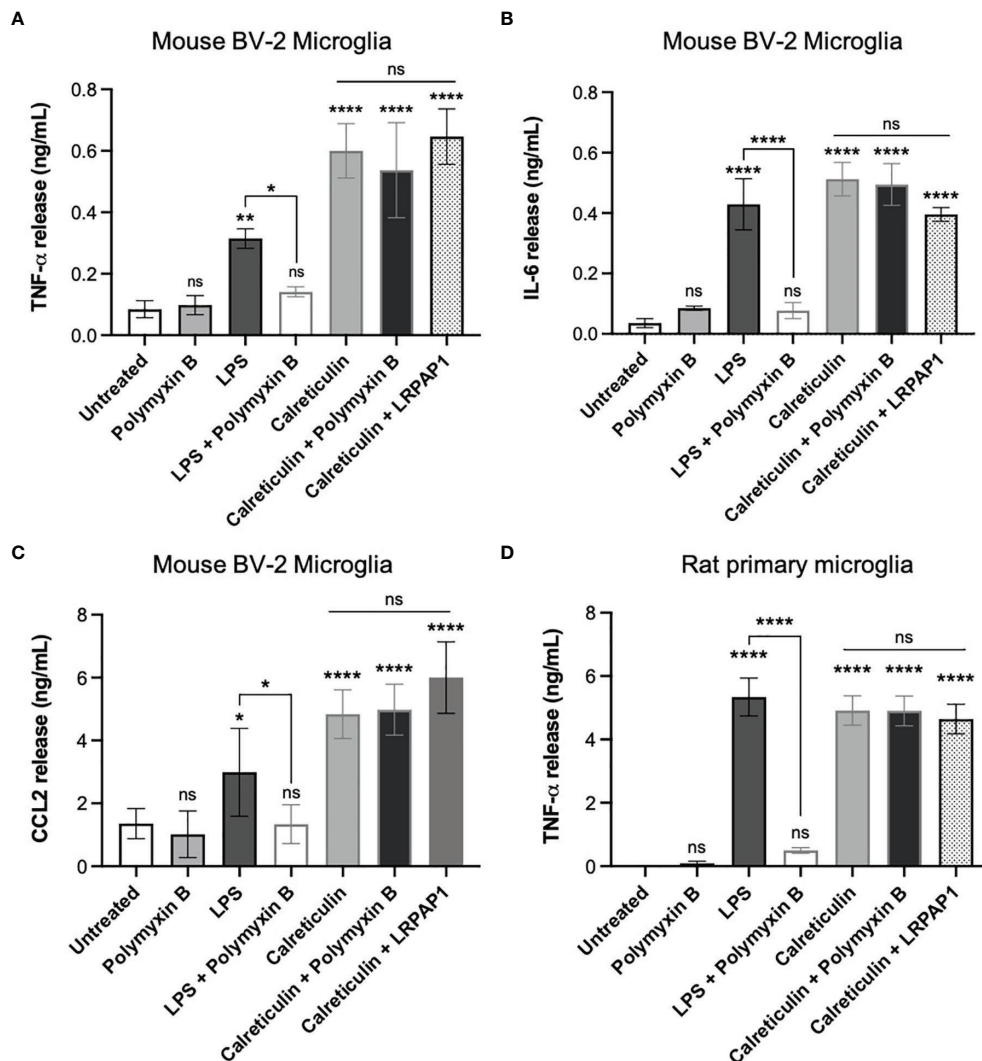


FIGURE 3 | Calreticulin induces cytokine and chemokine release from BV-2 and primary microglia independent of endotoxin and LRP1. BV-2 and rat primary microglia were treated with polymyxin B (10 U/mL), lipopolysaccharide (LPS, 100 ng/mL) $\pm$ polymyxin B, and calreticulin (170 nM) $\pm$ polymyxin B or LRPAP1 (100 nM) for 20 hours. Supernatants from treated BV-2 were assessed for **(A)** TNF- α , **(B)** IL-6 and **(C)** CCL2 by ELISA. **(D)** Supernatants of treated primary microglia were assessed for TNF- α by ELISA. Data presented as mean cytokine or chemokine release values, with error bars representing the SEM of at least 3 independent experiments. Statistical comparisons were made to the untreated control, or as illustrated by a comparison line, by one-way ANOVA. ns: $p \geq 0.05$, * $p < 0.05$, ** $p < 0.01$, **** $p < 0.0001$.

calreticulin, the kinetics of A β fibrillization was little affected, but the maximum fluorescence was reduced (**Figure 6A**). This suggests that calreticulin may help fold A β into forms with less β -sheet structure, consistent with calreticulin acting as a chaperone for A β . Imaging the A β aggregates after the fibrillization assay confirmed that calreticulin reduced the thioflavin T fluorescence of the A β aggregates (**Figure 6B**).

As oligomeric A β is widely considered to be more neurotoxic than A β fibrils (44), we next assessed the effect of calreticulin on A β oligomer formation. We incubated 10 μ M monomeric A β $\pm$ 1 μ M calreticulin for 18 hours at 4°C [conditions known to result in A β oligomers (43)], and imaged the resulting A β oligomers by transmission electron microscopy. A β oligomerized in the

presence of calreticulin resulted in larger oligomeric A β structures compared to A β oligomers formed in the absence of calreticulin (**Figures 6C, D**).

Taken together, this data suggests that calreticulin inhibits A β fibrillization, or folds A β into aggregates with less β -sheet structure, and promoted the formation of larger oligomeric structures.

Calreticulin Prevents A β -Induced Neuronal Loss

As calreticulin affects fibrillization and oligomerization of A β , we next tested whether calreticulin affected the neurotoxicity induced by A β . We have previously shown that nanomolar concentrations of A β induce neuronal loss mediated by

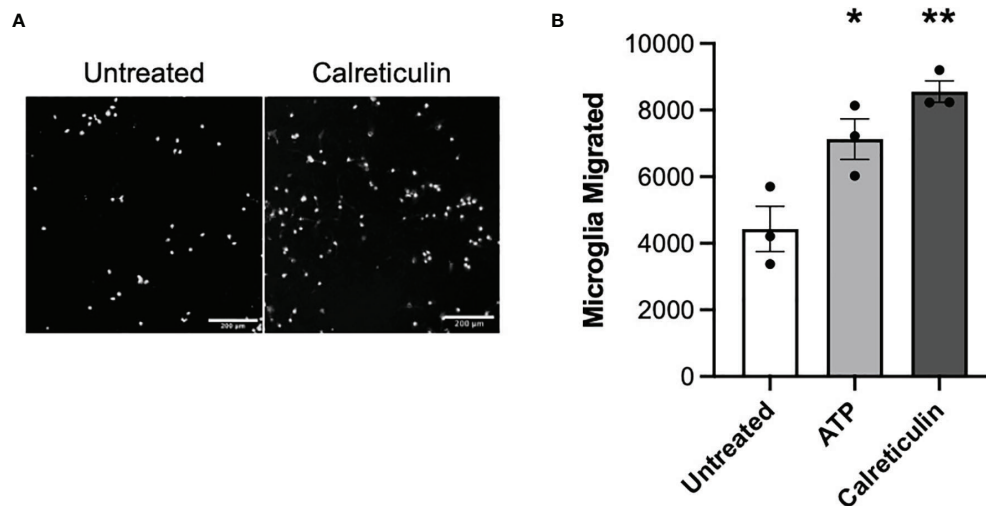


FIGURE 4 | Microglia migrate towards calreticulin. **(A)** Representative images and **(B)** quantified migration of BV-2 microglia migration across a transwell towards 17 nM calreticulin over 24 hours. 10 μM adenosine triphosphate (ATP) acted as the positive control. Microglia were stained with Hoechst 33342. Scale bar = 200 μm. Data presented as mean microglial migration values, with error bars representing the SEM of 3 independent experiments. Statistical comparisons were made to the untreated control by one-way ANOVA. * $p < 0.05$, ** $p < 0.01$.

microglia in mixed neuron–glial co-cultures (26). Here, we found that 250 nM Aβ induced loss of about 35% of the neurons over 72 hours (**Figures 7A, B**). 50 nM calreticulin alone (in the absence of Aβ) had no effect on neuronal or microglial numbers (**Figures 7A–C**), but did dramatically affect microglial morphology, as microglia became flatter with increased area (**Figures 7A, D**). Thus, calreticulin activates microglia in a way that is not detrimental to neurons. Indeed, addition of 10 nM calreticulin significantly inhibited the Aβ-induced neuronal loss, and 50 nM calreticulin almost completely prevented the Aβ-

induced neuronal loss (**Figures 7A, B**). Thus, extracellular calreticulin is neuroprotective at concentrations similar those released by brain cells in a variety of circumstances.

DISCUSSION

We investigated here whether brains cells release calreticulin, and whether extracellular calreticulin had any effects on microglia and neurons relevant to neuroinflammation and neurodegeneration.

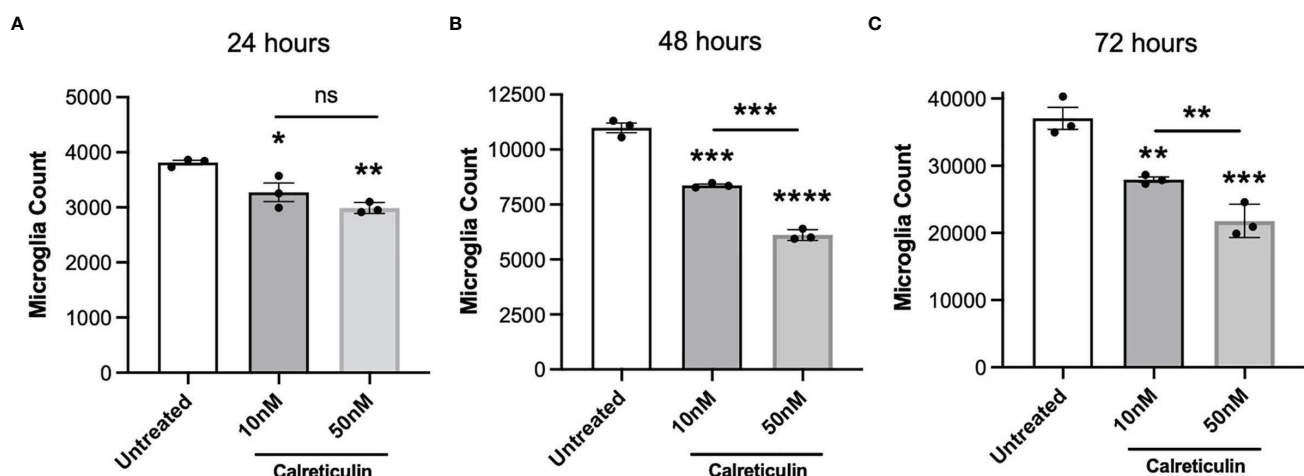


FIGURE 5 | Calreticulin dose-dependently inhibits the proliferation of BV-2 microglia. BV-2 were seeded at a density of 2000 cells/well, then treated with either 0, 10 or 50 nM calreticulin. After incubations for 24 hours **(A)**, 48 hours **(B)** or 72 hours **(C)**, microglia were stained with Hoechst 33342 and microglial cell numbers were quantified by fluorescence microscopy. Data presented as mean microglial cell numbers, with error bars representing the SEM of 3 independent experiments. Statistical comparisons were made to the untreated control, or as illustrated by a comparison line, by one-way ANOVA. ns: $p \geq 0.05$, * $p < 0.05$, ** $p < 0.01$, *** $p < 0.001$, **** $p < 0.0001$.

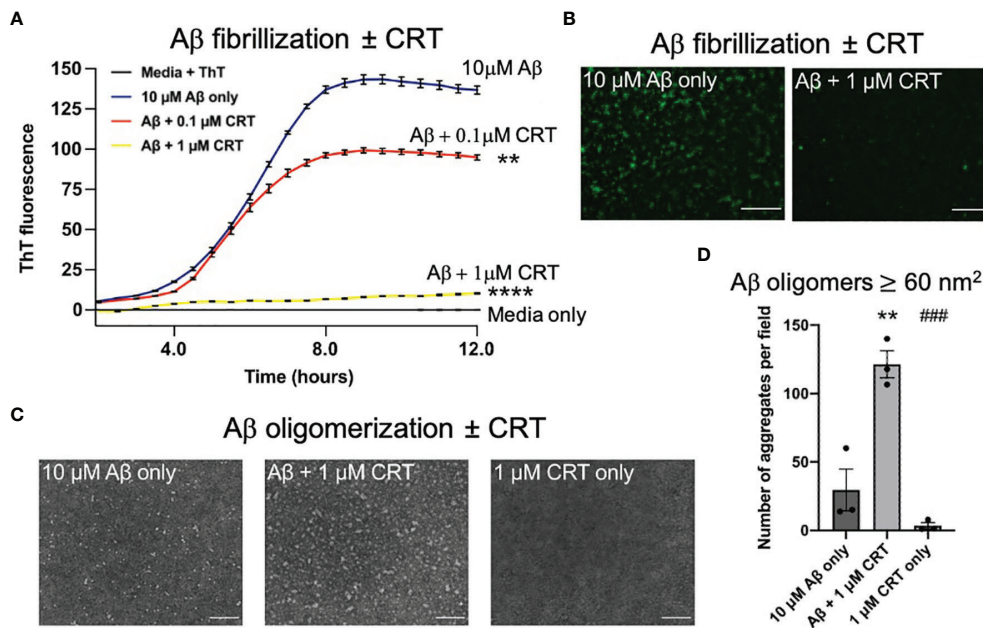


FIGURE 6 | Calreticulin prevents fibrillization and promotes formation of large oligomeric structures of amyloid beta. **(A)** 10 μ M of monomeric $A\beta$ was incubated at 31.5°C in DMEM with 10 μ M thioflavin T $\pm$ 0.1 μ M or 1 μ M calreticulin (CRT), and the fluorescence recorded over time. **(B)** At the end of the assay, each well was imaged using a fluorescence microscope. Scale bar = 50 μ m. **(C)** 10 μ M of monomeric $A\beta$ was incubated for 18 h at 4°C $\pm$ 1 μ M CRT to generate $A\beta$ oligomers imaged by transmission electron microscopy. Representative images shown. Scale bar = 100 nm. **(D)** Quantification of aggregate ≥ 60 (nm²) quantified from **(C)**. Data presented as mean final fluorescence or average aggregate size values, with error bars representing the SEM of at least 3 independent experiments. Statistical comparisons were made by one-way ANOVA. Asterisk (*) indicate significance (** $p < 0.01$, **** $p < 0.0001$) to 10 μ M $A\beta$ only. Hash (#) indicate significance (### $p < 0.001$) to 10 μ M $A\beta$ + 1 μ M CRT.

We found that nanomolar levels of calreticulin were released from crushed primary neurons, apoptotic BV-2 microglia killed with staurosporine, and BV-2 under ER stress from tunicamycin. This extends our previous finding that LPS can induce calreticulin release from primary mouse microglia (13). As calreticulin can be released from neurons and microglia in a variety of conditions, we tested whether extracellular calreticulin could act as an alarmin. We found that nanomolar calreticulin can chemoattract microglia and activate microglia to release proinflammatory cytokines. This indicates that calreticulin can act as an alarmin for microglia.

Alarmins are endogenous molecules released by necrotic cells and inflamed immune cells, to recruit and activate immune cells, in order to restore homeostasis of damaged tissues (14). Calreticulin is recognized as a damage-associated molecular pattern (DAMP), expressed on the surface of dead or dying cells, where it may induce phagocytosis and antigen presentation by dendritic cells [so called: immunogenic cell death (45, 46)]. These pro-phagocytic and immunogenic functions of calreticulin are dependent on being bound to the surface of dead or dying cells. However, calreticulin can also be released from activated neutrophils (47) and macrophages (12), and extracellular calreticulin can induce activation of monocytes, B cells and dendritic cells (15, 48). Thus, it is possible that calreticulin may act as an alarmin to immune cells other than microglia. Exogenous calreticulin has been shown to stimulate wound healing by recruiting macrophages, stimulating the proliferation of keratinocytes, fibroblasts and endothelial cells, and

increasing extracellular matrix production (42). Thus, if calreticulin is endogenously released by damaged tissues in the body or brain, it may help repair that tissue.

We did not investigate the mechanism by which calreticulin activates microglia, other than to show that chelation of LPS by polymyxin B, or inhibition of LRP1 by LRPAP1, did not prevent calreticulin-induced activation of microglia, which suggests that the activation is not mediated by LPS or LRP1. However, it has previously been shown that nanomolar calreticulin can induce the release of soluble LRP1 (sLRP1) from microglia, and that sLRP1 alone can induce cytokine release by an unknown mechanism (49). So, it is possible that calreticulin binds to LRP1 to induce the release of sLRP1, which then induces cytokine release, although one might expect that LRPAP1 would block such a mechanism (49). Alternatively, as a proteolytic fragment of calreticulin has previously been shown to activate myeloid cells *via* TLR4/CD14 (50), it is possible that full-length calreticulin binds to TLR4 or CD14 to activate microglia.

We found that calreticulin affected $A\beta$ aggregation. In conditions of $A\beta$ oligomerization, calreticulin increased the formation of larger $A\beta$ oligomers. In conditions of $A\beta$ fibrillization, calreticulin folded $A\beta$ into aggregates with less β -structure and $A\beta$ fibrillization. This is consistent with calreticulin acting as an extracellular chaperone for $A\beta$. Within the ER, calreticulin is an intracellular chaperone, binding proteins with a hydrophobic surface, enabling refolding (51). Calreticulin can directly bind $A\beta$ *via* the polypeptide binding site of calreticulin

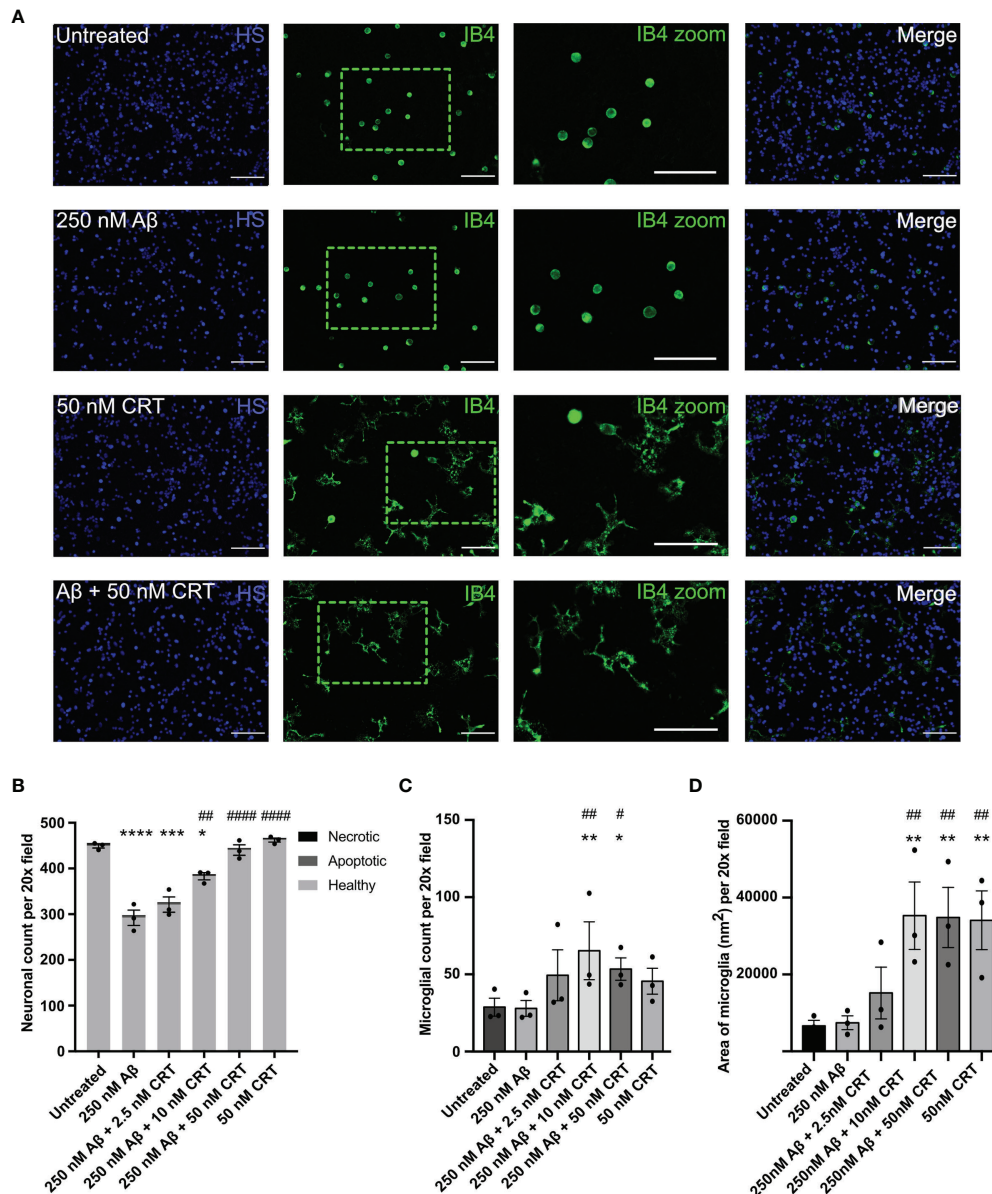


FIGURE 7 | Calreticulin protects against amyloid beta induced neuronal loss. Mixed neuron-glial co-cultures were treated $\pm$ monomeric 250 nM A β and $\pm$ 2.5 nM, 10 nM or 50 nM calreticulin (CRT) for 72 h at 37 °C. **(A)** Cultures were stained with isolectin B4 (green, to identify microglia), propidium iodide (red, to identify necrotic cells, not shown), Hoechst 33342 (blue, to identify nuclei and apoptotic cells) and representative, imaged fields are shown. Scale bars = 100 μ m. **(B)** Neuronal cell numbers and cell death were quantified by fluorescence microscopy. **(C)** Microglial cell numbers were quantified by fluorescence microscopy. **(D)** Microglial area was quantified as area of field stained with IB4. Data presented as mean neuronal count, microglial count, or area of microglia, with error bars representing the SEM of 3 independent experiments. Statistical comparisons were made by one-way ANOVA. Asterisk (*) indicate significance compared to untreated control (* p <0.05, ** p <0.01, *** p <0.001, **** p <0.0001). Hash (#) indicate significance compared to A β only (# p <0.05, ## p <0.01, ### p <0.0001).

and the hydrophobic C-terminus of A β (21), and a significant fraction of A β in human cerebrospinal fluid is found bound to calreticulin (20), suggesting that extracellular calreticulin may significantly impact A β aggregation in human brain. It has previously been shown that five different chaperones: α B-crystallin, heat shock protein 70, clusterin, haptoglobin and α_2 -macroglobulin, each block A β neurotoxicity by binding to A β oligomers, promoting their assembly into larger oligomeric

species, with consequent shielding of the reactive surfaces and reduced fibrilization (52, 53). This fits our data for calreticulin, therefore suggesting that calreticulin inhibits A β neurotoxicity by acting as an A β chaperone.

As calreticulin induced microglial activation and interfered with A β aggregation, we tested whether it affected A β -induced neurotoxicity. Nanomolar levels of calreticulin prevented A β -induced neurotoxicity. We did not investigate the mechanism of

this protection, but possible mechanisms include calreticulin interfering with A β aggregation or microglial activation. Another possibility is that calreticulin increases microglial uptake and degradation of A β .

Calreticulin alone induced a strong morphological transition of microglia in primary neuron-glial co-cultures, from an unattached, spheroidal morphology to an attached and partially ramified morphology. This may appear contrary to the finding that microglial activation *in vivo* is accompanied by a transition from a highly ramified morphology to a more amoeboid morphology (30). However, in culture, untreated microglia are mainly spheroidal and relatively unattached (on uncoated culture plates), but become attached and flatten onto the culture plate in response to LPS (54, 55), and become ramified in response to ATP (55) or TNF- α and INF γ (54).

Calreticulin alone had no significant effect on microglial numbers in primary neuron-glial co-cultures, and microglial numbers increased in the presence of both calreticulin and A β , but calreticulin alone caused a small decrease in proliferation of BV-2 cells. BV-2 cells are an immortalized microglial cell line (37), so their proliferative response may not be representative of primary cells. Microglial activation generally increases proliferation (56). However, calreticulin has been found to inhibit the proliferation of a variety of cell types (40, 41, 57).

It used to be thought that microglia could be activated into only two main states, M1 and M2, but there are now known to be multiple forms of microglial activation (58, 59), some neurotoxic and some neuroprotective (60, 61). Calreticulin appears to recruit and activate microglia without neurotoxicity, consistent with calreticulin being an alarmin.

In summary, neurons and microglia can release calreticulin in multiple conditions, resulting in nanomolar levels of extracellular calreticulin. Nanomolar calreticulin can induce microglial migration and activation, and can interfere with A β aggregation and neurotoxicity. As calreticulin has been found in human cerebrospinal fluid (20), it may be protecting the brain by acting as an alarmin, extracellular chaperone and neuroprotectant. If so, increasing extracellular calreticulin levels might be beneficial in brain pathologies.

REFERENCES

- Coppolino MG, Dedhar S. Calreticulin. *Int J Biochem Cell Biol* (1998) 30:553–8. doi: 10.1016/S1357-2725(97)00153-2
- Hammond C, Braakman I, Helenius A. Role of N-Linked Oligosaccharide Recognition, Glucose Trimming, and Calnexin in Glycoprotein Folding and Quality Control. *Proc Natl Acad Sci USA* (1994) 91:913–7. doi: 10.1073/pnas.91.3.913
- Gardai SJ, McPhillips KA, Frasch SC, Janssen WJ, Starefeldt A, Murphy-Ullrich JE, et al. Cell-Surface Calreticulin Initiates Clearance of Viable or Apoptotic Cells Through Trans-Activation of LRP on the Phagocyte. *Cell* (2005) 123:321–34. doi: 10.1016/j.cell.2005.08.032
- Scholnik-Cabrera A, Oldak B, Juárez M, Cruz-Rivera M, Flisser A, Mendlovic F. Calreticulin in Phagocytosis and Cancer: Opposite Roles in Immune Response Outcomes. *Apoptosis* (2019) 24:245–55. doi: 10.1007/s10495-019-01532-0
- Martins I, Kepp O, Galluzzi L, Senovilla L, Schlemmer F, Adjemian S, et al. Surface-Exposed Calreticulin in the Interaction Between Dying Cells and

DATA AVAILABILITY STATEMENT

The original contributions presented in the study are included in the article/**Supplementary Material**. Further inquiries can be directed to the corresponding author.

ETHICS STATEMENT

The animal study was reviewed and approved by University of Cambridge Animal Welfare and Ethical Review Body.

AUTHOR CONTRIBUTIONS

KR, EK, CB, and TC performed and analyzed experiments. GB conceived and managed the research. GB and KR wrote most of the manuscript. All authors reviewed and approved the manuscript.

FUNDING

This work received funding from the Biotechnology and Biological Sciences Research Council UK (BBSRC BB/M011194/1).

ACKNOWLEDGMENTS

We wish to thank David Allendorf for assistance in the laboratory.

SUPPLEMENTARY MATERIAL

The Supplementary Material for this article can be found online at: <https://www.frontiersin.org/articles/10.3389/fimmu.2022.859686/full#supplementary-material>

Phagocytes. *Ann N Y Acad Sci* (2010) 1209:77–82. doi: 10.1111/j.1749-6632.2010.05740.x

- Hong C, Qiu X, Li Y, Huang Q, Zhong Z, Zhang Y, et al. Functional Analysis of Recombinant Calreticulin Fragment 39–272: Implications for Immunobiological Activities of Calreticulin in Health and Disease. *J Immunol* (2010) 185:4561–9. doi: 10.4049/jimmunol.1000536
- Garg AD, Krysko DV, Verfaillie T, Kaczmarek A, Ferreira GB, Marysael T, et al. A Novel Pathway Combining Calreticulin Exposure and ATP Secretion in Immunogenic Cancer Cell Death. *EMBO J* (2012) 31:1062–79. doi: 10.1038/emboj.2011.497
- Païdassi H, Tacnet-Delorme P, Verneret M, Gaboriaud C, Houen G, Duus K, et al. Investigations on the C1q-Calreticulin-Phosphatidylserine Interactions Yield New Insights Into Apoptotic Cell Recognition. *J Mol Biol* (2011) 408:277–90. doi: 10.1016/j.jmb.2011.02.029
- Vandivier RW, Ogden CA, Fadok VA, Hoffmann PR, Brown KK, Botto M, et al. Role of Surfactant Proteins a, D, and C1q in the Clearance of Apoptotic Cells *In Vivo* and *In Vitro*: Calreticulin and CD91 as a Common Collectin Receptor Complex. *J Immunol* (2002) 169:3978–86. doi: 10.4049/jimmunol.169.7.3978

10. Stuart GR, Lynch NJ, Day AJ, Schwaeble WJ, Sim RB. The C1q and Collectin Binding Site Within C1q Receptor (Cell Surface Calreticulin). *Immunopharmacology* (1997) 38:73–80. doi: 10.1016/S0162-3109(97)00076-3
11. Ogden CA, DeCathelineau A, Hoffmann PR, Bratton D, Fadok B, Ghebrehiwet VA, et al. C1q and Mannose Binding Lectin Engagement of Cell Surface Calreticulin and CD91 Initiates Macropinocytosis and Uptake of Apoptotic Cells. *J Exp Med* (2001) 194:781–95. doi: 10.1084/jem.194.6.781
12. Feng M, Marjon KD, Zhu F, Weissman-Tsukamoto R, Levett A, Sullivan K, et al. Programmed Cell Removal by Calreticulin in Tissue Homeostasis and Cancer. *Nat Commun* (2018) 9:1–15. doi: 10.1038/s41467-018-05211-7
13. Cockram TOJ, Puigdemílv M, Brown GC. Calreticulin and Galectin-3 Opsonise Bacteria for Phagocytosis by Microglia. *Front Immunol* (2019) 10:2647. doi: 10.3389/fimmu.2019.02647
14. Bianchi ME. Damps, Pamps and Alarmins: All We Need to Know About Danger. *J Leukoc Biol* (2007) 81:1–5. doi: 10.1189/jlb.0306164
15. Bajor A, Tischer S, Figueiredo C, Wittmann M, Immenschuh S, Blasczyk R, et al. Modulatory Role of Calreticulin as Chaperone for Dendritic Cell-Based Immunotherapy. *Clin Exp Immunol* (2011) 165:220–34. doi: 10.1111/j.1365-2249.2011.04423.x
16. Bak SP, Amiel E, Walters JJ, Berwin B. Calreticulin Requires an Ancillary Adjuvant for the Induction of Efficient Cytotoxic T Cell Responses. *Mol Immunol* (2008) 45:1414–23. doi: 10.1016/j.molimm.2007.08.020
17. Dabbs RA, Wyatt AR, Yerbury JJ, Ecroyd H, Wilson MR. *Extracellular Chaperones*. Berlin, Heidelberg: Springer (2010) p. 1–34. doi: 10.1007/128_2010_85
18. Wyatt AR, Yerbury JJ, Ecroyd H, Wilson MR. Extracellular Chaperones and Proteostasis. *Annu Rev Biochem* (2013) 82:295–322. doi: 10.1146/annurev-biochem-072711-163904
19. Wyatt AR, Yerbury JJ, Dabbs RA, Wilson MR. Roles of Extracellular Chaperones in Amyloidosis. *J Mol Biol* (2012) 421:499–516. doi: 10.1016/j.jmb.2012.01.004
20. Erickson RR, Dunning LM, Olson DA, Cohen SJ, Davis AT, Wood WG, et al. In Cerebrospinal Fluid ER Chaperones Erp57 and Calreticulin Bind β -Amyloid. *Biochem Biophys Res Commun* (2005) 332:50–7. doi: 10.1016/j.bbrc.2005.04.090
21. Houen G, Duus K, Hansen P. Interaction of Calreticulin With Amyloid Beta Peptide 1–42. *Protein Pept Lett* (2008) 15:103–7. doi: 10.2174/092986608783330459
22. Blasi E, Barluzzi R, Bocchini V, Mazzolla R, Bistoni F. Immortalization of Murine Microglial Cells by a V-Raf / V-Myc Carrying Retrovirus. *J Neuroimmunol* (1990) 27:229–37. doi: 10.1016/0165-5728(90)90073-V
23. Shen X, Burguillos MA, Osman AM, Frijhoff J, Carrillo-Jiménez A, Kanatani S, et al. Glioma-Induced Inhibition of Caspase-3 in Microglia Promotes a Tumor-Supportive Phenotype. *Nat Immunol* (2016) 17:1282–90. doi: 10.1038/ni.3545
24. Carrillo-Jimenez A, Puigdemílv M, Vilalta A, Venero JL, Brown GC, StGeorge-Hyslop P, et al. Effective Knockdown of Gene Expression in Primary Microglia With Sirna and Magnetic Nanoparticles Without Cell Death or Inflammation. *Front Cell Neurosci* (2018) 12:313. doi: 10.3389/fncel.2018.00313
25. Allendorf DH, Puigdemílv M, Brown GC. Activated Microglia Desialylate Their Surface, Stimulating Complement Receptor 3-Mediated Phagocytosis of Neurons. *Glia* (2020) 68:989–98. doi: 10.1002/glia.23757
26. Neniskyte U, Neher JJ, Brown GC. Neuronal Death Induced by Nanomolar Amyloid β is Mediated by Primary Phagocytosis of Neurons by Microglia. *J Biol Chem* (2011) 286:39904–13. doi: 10.1074/jbc.M111.267583
27. Naiki H, Higuchi K, Hosokawa M, Takeda T. Fluorometric Determination of Amyloid Fibrils *In Vitro* Using the Fluorescent Dye, Thioflavine T. *Anal Biochem* (1989) 177:244–9. doi: 10.1016/0003-2697(89)90046-8
28. Abdullahi A, Stanojic M, Parousis A, Patsouris D, Jeschke MG. Modeling Acute Er Stress *In Vivo* and *In Vitro*. *Shock* (2017) 47:506–13. doi: 10.1097/SHK.0000000000000759
29. Simenc J, Lipnik-Stangelj M. Staurosporine Induces Different Cell Death Forms in Cultured Rat Astrocytes. *Radiol Oncol* (2012) 46:312–20. doi: 10.2478/v10019-012-0036-9
30. Torres-Platas SG, Comeau S, Rachalski A, Bo GD, Cruceanu C, Turecki G, et al. Morphometric Characterization of Microglial Phenotypes in Human Cerebral Cortex. *J Neuroinflamm* (2014) 11:12. doi: 10.1186/1742-2094-11-12
31. Dello Russo C, Cappoli N, Coletta I, Mezzogori D, Paciello F, Pozzoli G, et al. The Human Microglial HMC3 Cell Line: Where do We Stand? A Systematic Literature Review. *J Neuroinflamm* (2018) 15:1–24. doi: 10.1186/s12974-018-1288-0
32. Wakelin SJ, Sabroe I, Gregory CD, Poxton IR, Forsythe JLR, Garden OJ, et al. “Dirty Little Secrets”—Endotoxin Contamination of Recombinant Proteins. *Immunol Lett* (2006) 106:1–7. doi: 10.1016/j.imlet.2006.04.007
33. Cardoso LS, Araujo MI, Góes AM, Pacífico LG, Oliveira RR, Oliveira SC. Polymyxin B as Inhibitor of LPS Contamination of Schistosoma Mansoni Recombinant Proteins in Human Cytokine Analysis. *Microb Cell Fact* (2007) 6:1. doi: 10.1186/1475-2859-6-1
34. Fricker M, Oliva-Martin MJ, Brown GC. Primary Phagocytosis of Viable Neurons by Microglia Activated With LPS or $A\beta$ is Dependent on Calreticulin/LRP Phagocytic Signalling. *J Neuroinflamm* (2012) 9:697. doi: 10.1186/1742-2094-9-196
35. Orr AW, Pedraza CE, Pallero MA, Elzie CA, Goicoechea S, Strickland DK, et al. Low Density Lipoprotein Receptor-Related Protein is a Calreticulin Coreceptor That Signals Focal Adhesion Disassembly. *J Cell Biol* (2003) 161:1179–89. doi: 10.1083/jcb.200302069
36. Williams SE, Ashcom JD, Argraves WS, Strickland DK. A Novel Mechanism for Controlling the Activity of Alpha 2-Macroglobulin Receptor/Low Density Lipoprotein Receptor-Related Protein. Multiple Regulatory Sites for 39-Kda Receptor-Associated Protein. *J Biol Chem* (1992) 267:9035–40. doi: 10.1016/S0021-9258(19)50384-2
37. Henn A, Lund S, Hedtjörn M, Schrattenholz A, Pörzgen P, Leist M. The Suitability of BV2 Cells as Alternative Model System for Primary Microglia Cultures or for Animal Experiments Examining Brain Inflammation. *ALTEX* (2009) 26:83–94. doi: 10.14573/alte.2009.2.83
38. Ohsawa K, Irino Y, Nakamura Y, Akazawa C, Inoue K, Kohsaka S. Involvement of P2X4 and P2Y12 Receptors in ATP-Induced Microglial Chemotaxis. *Glia* (2007) 55:604–16. doi: 10.1002/glia.20489
39. Lu Y-C, Weng W-C, Lee H. Functional Roles of Calreticulin in Cancer Biology. *BioMed Res Int* (2015) 2015:1–9. doi: 10.1155/2015/526524
40. Alur M, Nguyen MM, Eggen SE, Jiang F, Dadras SS, Stern J, et al. Suppressive Roles of Calreticulin in Prostate Cancer Growth and Metastasis. *Am J Pathol* (2009) 175:882–90. doi: 10.2353/ajpath.2009.080417
41. Weng WC, Lin KH, Wu PY, Lu YC, Weng YC, Wang BJ, et al. Calreticulin Regulates VEGF-a in Neuroblastoma Cells. *Mol Neurobiol* (2015) 52:758–70. doi: 10.1007/s12035-014-8901-8
42. Gold LI, Rahman M, Blechman KM, Greives MR, Churgin S, Michaels J, et al. Overview of the Role for Calreticulin in the Enhancement of Wound Healing Through Multiple Biological Effects. *J Invest Dermatol Symp Proc* (2006) 11:57–65. doi: 10.1038/sj.jidsymp.5650011
43. Stine WB, Jungbauer L, Yu C, LaDu MJ. Preparing Synthetic $A\beta$ in Different Aggregation States. *Methods Mol Biol* (2010) 670:13–32. doi: 10.1007/978-1-60761-744-0_2
44. Amin L, Harris DA. $A\beta$ Receptors Specifically Recognize Molecular Features Displayed by Fibril Ends and Neurotoxic Oligomers. *Nat Commun* (2021) 12:3451. doi: 10.1038/s41467-021-23507-z
45. Obeid M, Tesniere A, Ghiringhelli F, Fimia GM, Apetoh L, Perfettini JL, et al. Calreticulin Exposure Dictates the Immunogenicity of Cancer Cell Death. *Nat Med* (2007) 13:54–61. doi: 10.1038/nm1523
46. Galluzzi L, Buqué A, Kepp O, Zitvogel L, Kroemer G. Immunogenic Cell Death in Cancer and Infectious Disease. *Nat Rev Immunol* (2017) 17:97–111. doi: 10.1038/nri.2016.107
47. Eggleton P, Lieu TS, Zappi EG, Sastry K, Coburn J, Zaner KS, et al. Calreticulin is Released From Activated Neutrophils and Binds to C1q and Mannan-Binding Protein. *Clin Immunol Immunopathol* (1994) 72:405–9. doi: 10.1006/clin.1994.1160
48. Pike SE, Yao L, Jones KD, Cherney B, Appella E, Sakaguchi K, et al. Vasostatin, a Calreticulin Fragment, Inhibits Angiogenesis and Suppresses Tumor Growth. *J Exp Med* (1998) 188:2349. doi: 10.1084/JEM.188.12.2349
49. Brifault C, Gilder AS, Laudati E, Banki M, Gonias SL. Shedding of Membrane-Associated LDL Receptor-Related Protein-1 From Microglia Amplifies and Sustains Neuroinflammation. *J Biol Chem* (2017) 292:18699–712. doi: 10.1074/JBC.M117.798413
50. Li Y, Zeng X, He L, Yuan H. Dendritic Cell Activation and Maturation Induced by Recombinant Calreticulin Fragment 39–272. *Int J Clin Exp Med* (2015) 8:7288–96.

51. Williams DB. Beyond Lectins: The Calnexin/Calreticulin Chaperone System of the Endoplasmic Reticulum. *J Cell Sci* (2006) 119:615–23. doi: 10.1242/jcs.02856
52. Mannini B, Cascella R, Zampagni M, Van Waarde-Verhagen M, Meehan S, Roodveldt C, et al. Molecular Mechanisms Used by Chaperones to Reduce the Toxicity of Aberrant Protein Oligomers. *Proc Natl Acad Sci USA* (2012) 109:12479–84. doi: 10.1073/pnas.1117799109
53. Narayan P, Meehan S, Carver JA, Wilson MR, Dobson CM, Klenerman D. Amyloid- β Oligomers are Sequestered by Both Intracellular and Extracellular Chaperones. *Biochemistry* (2012) 51:9270–6. doi: 10.1021/bi301277k
54. Lively S, Schlichter LC. Microglia Responses to Pro-Inflammatory Stimuli (LPS, $\text{Ifn}\gamma$ +Tnf α) and Reprogramming by Resolving Cytokines (IL-4, IL-10). *Front Cell Neurosci* (2018) 12:215/FULL. doi: 10.3389/FNCEL.2018.00215/FULL
55. Wollmer MA, Lucius R, Wilms H, Held-Feindt J, Sievers J, Mentlein R. ATP and Adenosine Induce Ramification of Microglia *In Vitro*. *J Neuroimmunol* (2001) 115:19–27. doi: 10.1016/S0165-5728(01)00257-0
56. Fügen P, Hefendehl JK, Veeraghavalu K, Wendeln AC, Schlosser C, Obermüller U, et al. Microglia Turnover With Aging and in an Alzheimer's Model *via* Long-Term *In Vivo* Single-Cell Imaging. *Nat Neurosci* (2017) 20:1371–6. doi: 10.1038/nn.4631
57. Pike SE, Yao L, Setsuda J, Jones KD, Cherney B, Appella E, et al. Calreticulin and Calreticulin Fragments Are Endothelial Cell Inhibitors That Suppress Tumor Growth. *Blood* (1999) 94:2461–8. doi: 10.1182/blood.v94.7.2461.419a26_2461_2468
58. Town T, Nikolic V, Tan J. The Microglial “Activation” Continuum: From Innate to Adaptive Responses. *J Neuroinflamm* (2005) 2:1–10. doi: 10.1186/1742-2094-2-24
59. Dubbelaar ML, Kracht L, Eggen BJL, Boddeke EWGM. The Kaleidoscope of Microglial Phenotypes. *Front Immunol* (2018) 9:1753. doi: 10.3389/fimmu.2018.01753
60. Chen Z, Trapp BD. Microglia and Neuroprotection. *J Neurochem* (2016) 136:10–7. doi: 10.1111/jnc.13062
61. Bellver-Landete V, Bretheau F, Mailhot B, Vallières N, Lessard M, Janelle ME, et al. Microglia are an Essential Component of the Neuroprotective Scar That Forms After Spinal Cord Injury. *Nat Commun* (2019) 10:1–18. doi: 10.1038/s41467-019-08446-0

Conflict of Interest: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's Note: All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

Copyright © 2022 Reid, Kitchener, Butler, Cockram and Brown. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.