

Your manuscript has been given the ID JEXBIO/2017/161463.

Take-off mechanisms in parasitoid wasps

by

Burrows, M., and Dorosenko, M.

Department of Zoology, University of Cambridge,
Cambridge CB2 3EJ, England

With 8 Figures and 5 videos as Supplementary Material

Keywords: jumping, flying, kinematics, high speed imaging, escape movements, biomechanics

Running Title: take-off in parasitoid wasps

Please send correspondence to Professor M Burrows at the above address

Telephone +44 (0)1223 336628

Email mb135@hermes.cam.ac.uk

1 ABSTRACT

2 High speed video analyses of the natural behaviour of parasitoid wasps revealed three
3 strategies used to launch themselves into the air. Which strategy is the most energy
4 efficient? In *Pteromalus puparum*, 92% of take-offs by were propelled entirely by
5 movements of the middle and hind legs which were depressed at their coxo-
6 trochanteral and extended at their femoro-tibial joints. The front legs left the ground
7 first, followed by the hind legs, so that the middle legs provided the final propulsion.
8 Second, in other species of a similar mass, *Cotesia glomerata* and *Leptopilina*
9 *boulardi*, all take-offs were propelled by a mean of 2.8 and 3.8 wingbeats respectively
10 with little or no contribution from the legs. The first strategy resulted in take-off times
11 that were four times shorter (5 versus 22.8 ms) and take-off velocities that were four
12 times faster (0.8 versus 0.2 m s⁻¹). Calculations from the kinematics indicate that
13 propulsion by the legs was the most energy efficient strategy, because more energy is
14 put into propulsion of the body, whereas in take-off propelled by repetitive wing
15 movements energy is lost to generating these movements and moving the air. In
16 heavier species such as *Netelia testacea* and *Amblyteles armatorius*, take-off was
17 propelled by the combined movements of the middle and hind legs and wingbeats. In
18 *A. armatorius*, this resulted in the longest mean take-off time of 33.8 ms but an
19 intermediate take-off velocity of 0.4 m s⁻¹. In all three strategies the performance
20 could be explained without invoking energy storage and power amplification
21 mechanisms.

22

23

24 INTRODUCTION

25 Three different strategies are used by a wide diversity of insects to launch a take-off
26 from the ground and into the air. First, rapid movements of the legs propel jumping to
27 achieve take-off, without any contribution from the wings. Second, repeated
28 movements of the wings generate take-off with no obvious thrust contributed by the
29 legs. The third strategy is to combine propulsive movements of both the legs and the
30 wings. This paper analyses the dynamics of these different strategies for take-off and
31 compares their energy efficiencies. We propose that generating take-off by a rapid
32 jump propelled by the legs is more energy efficient than a take-off generated by
33 repetitive flapping movements of the wings and could provide the basis of an
34 explanation as to why so many winged insects jump to take-off. This hypothesis is
35 tested by analysing the take-off mechanisms and performance of a five species of
36 parasitoid wasps (Hymenoptera).

37

38 The most well-documented species of the Hymenoptera that jump are wingless ants.
39 Trap jaw ants, *Odontomachus bauri*, use rapid closing movements of the mandibles to
40 strike a hard substrate and propel a backwards escape jump (Patek et al., 2006) that
41 increase the chance of surviving encounters with predatory insects such as antlions
42 (Neuroptera) (Larabee and Suarez, 2015). They also use leg movements to jump
43 forwards (Sorger, 2015). Ants such as *Polyrhachis laboriosa* that live in trees can
44 jump downwards to escape predators or find new foraging sites (Mercier and Lenoir,
45 1999). Others like the wingless workers of *Cephalotes atratus* live higher in the
46 canopy and either jump or fall but adjust their glide path by asymmetric movements of
47 the hind legs and gaster to regain contact with the trunk of the same tree lower down
48 (Yanoviak et al., 2005; Yanoviak et al., 2010). Other ants like the Indian ant
49 *Harpegnathos saltator* jump by more conventional propulsive movements of the legs.

1 to escape predators, to catch prey in flight and to perform group movements that may
2 aid in prey detection or predator avoidance (Musthak Ali et al., 1992). The jumping
3 mechanism used is unclear. In one description the hind legs push first and then the
4 rapid movements of the middle legs generate the final propulsion to modest take-off
5 velocities of 0.7 m s^{-1} (Tautz et al., 1994). A second description indicates that
6 propulsive movements of the middle and hind legs are synchronous, a conclusion that
7 is said to be supported by the electrical activity of muscles in the middle and hind legs
8 also being synchronous during “fictive” jumping movements (Baroni Urbani et al.,
9 1994). Another species, *Myrmecia nigrocincta* is also reported to move its middle and
10 hind legs synchronously to propel jumping (Tautz et al., 1994). *Gigantiops destructor*
11 combines movements of the middle and hind legs with a forward rotation of the gaster
12 that shifts the centre of mass of the body and could reduce body spin once airborne
13 (Tautz et al., 1994).

14
15 Some larval wasps have abrupt movements that resemble jumping. The larvae of
16 *Neuroterus saltatorius* (Cynipidae) develop in small galls on the leaves of oak trees.
17 When these galls fall to the ground rapid contractions of the U-shaped larva within
18 make the gall bounce 10 mm high (Manier and Deamer, 2014). Similarly, a larva of
19 *Bathyplectes anurus* (Ichneumonidae) spins a cocoon within that of its larval weevil
20 host and its twitch-like movements propel jumps some 50 mm high (Saeki et al.,
21 2016).

22
23 Adult, winged wasps of a few species within the family (Eupelmidae) are more
24 conventional but able jumpers propelled by movements of the legs (Gibson, 1986). No
25 measurements of their performance have been reported, and mechanisms have instead
26 been inferred from their thoracic morphology and musculature. The middle legs are
27 thought to propel jumping and the necessary power is suggested to be generated by
28 different mesothoracic muscles in males and females, although jumping appears to be
29 similar in both sexes (Gibson, 1986). A pad of material onto which one of these
30 muscles attaches has some of the properties of the elastic protein resilin, leading to the
31 inference that the jumping mechanism must involve the storage of energy (Gibson,
32 1986). A mechanism, however, has not been demonstrated that would explain how
33 energy is stored. These wasps unlike the wingless ants, but like many other insects
34 such as moths (Burrows and Dorosenko, 2015a) and butterflies (Bimbard et al.,
35 2013; Sunada et al., 1993), may use jumping as a means of launching into flight. This
36 take-off objective might also be met by repetitive beating of the wings acting alone, or
37 in concert with propulsive legs movements as in whiteflies (Ribak et al., 2016).

38
39 Why take-off with a jump powered by rapid leg movements rather than simply
40 flapping the wings? To address this question, this paper analyses from high speed
41 videos the take-off strategies and mechanisms of parasitoid wasps, selected for two
42 reasons. First, these species fall into two groups based on their mass; three have
43 masses of 1 mg or less, while the others have masses that are 50 times greater. We ask
44 of all these species whether take-off is propelled by leg movements and if so which
45 legs are used. Alternatively is take-off propelled by wing movements or by a
46 combination of legs and wing movements? From these analyses we calculate which
47 mechanisms are the most energy efficient and whether energy storage with
48 concomitant power amplification has to be invoked to explain the observed take-off
49 times and velocities.

50
51

1 MATERIALS AND METHODS

2 Five species of parasitoid wasps were analysed. 1. *Pteromalus puparum* (Linnaeus,
3 1758) (family Pteromalidae) (Fig. 1A). 2. *Cotesia glomerata* (Linnaeus, 1758),
4 formerly *Apanteles glomeratus* (family Braconidae) (Fig. 1B). Both species were
5 found parasitizing a culture of the cabbage white butterfly. 3. *Leptopilina boulandi*,
6 (Barbotin, Carton & Kelner-Pillault, 1979) (family Figitidae) (Fig. 1C,D) was found in
7 a *Drosophila* culture. 4. *Amblyteles armatorius* (Forster, 1771) (Fig. 1E) and 5.
8 *Netelia testacea* (Gravenhorst, 1829) (family Ichneumonidae) were caught amongst
9 garden plants in Girton, Cambridge, UK. The phylogenies of these wasps within the
10 Hymenoptera have recently been analysed (Branstetter et al., 2017; Quicke et al.,
11 2012).

12
13 Live wasps were photographed with a Nikon D7200 camera fitted with a 100 mm
14 Nikon macro lens. The anatomy of the legs was examined in intact wasps, and in those
15 fixed and stored in 70% alcohol, or 50% glycerol. Leg lengths were measured to an
16 accuracy of 0.1 mm from images taken with a GX CAM 5-C camera (GT Vision Ltd.,
17 Stansfield, Suffolk, UK) attached to a Leica MZ16 microscope (Wetzlar, Germany)
18 and projected onto a monitor (Table 1). Body masses were determined to an accuracy
19 of 0.1 mg with a Mettler Toledo AB104 balance (Beaumont Leys, Leicester, UK).

20
21 Sequential images of take-offs were captured with a single Photron Fastcam SA3
22 camera (Photron (Europe) Ltd, High Wycombe, Bucks., UK) fitted with a 100 mm
23 micro Tokina lens. Frame rates of 1000 s⁻¹ with an exposure time of 0.2 ms and
24 5000 s⁻¹ with an exposure time of 0.1 ms were used. Several of the three smaller
25 species were placed at the same time in a chamber made of optical quality glass that
26 was 25 mm wide, 25 mm tall and 12 mm deep. The two larger species were placed
27 individually in a larger chamber measuring 80 mm wide, 80 mm tall and 25 mm deep.
28 The camera pointed directly at the middle of these chambers and focused on an
29 individual wasp. The floor, side walls and ceiling of each chamber were made of 12
30 mm thick, closed cell foam (Plastazote, Watkins and Doncaster, Cranbrook, Kent,
31 UK) from which the wasps would take-off spontaneously. Tracks of the movements of
32 specific body parts were made manually frame by frame with Tracker software
33 (<http://physlets.org/tracker/>); auto-tracking failed to follow the movements of small
34 body parts reliably. The frame at which a particular leg lost contact with the ground
35 was determined by playing the video backwards and forwards frame by frame. An
36 abrupt shift in position of a tarsus indicated that it was no longer load bearing and had
37 lost contact with the ground. Take-off was indicated by the last leg losing contact with
38 the ground so that the insect was airborne; it was designated as time $t = 0$ ms. The
39 acceleration time was defined as the period from the first detectable movement of the
40 propulsive legs, or wings, until take-off. Peak velocity was calculated as the distance
41 moved in a rolling 3 point average of successive images before take-off. A point on
42 the body that could be recognized in successive frames and was close to the centre of
43 mass was selected for measurements of the trajectory. The angle subtended by a line
44 joining these positions after take-off, relative to the natural horizontal, gave the
45 trajectory angle. The body angle was defined as the angle subtended by the
46 longitudinal axis of the wasp relative to the natural horizontal at take-off. The results
47 are based on high speed videos of 292 take-offs by 35 wasps from at temperatures of
48 23-25° C. At least three jumps were analysed in detail for each individual wasp. The
49 number of specimens of *N. testacea* was small, so that no quantitative analysis was
50 performed. Data are presented as means \pm standard error of the mean (s.e.m.) for an

1 individual insect and as grand means (mean of means) for all the individuals of a
2 particular species.

3 4 **RESULTS**

5 **Body Form**

6 The parasitoid wasps fell into two distinct groups on the basis of their mass and size.
7 First were three small species *L. boulandi*, *P. puparum* and *C. glomerata* with body
8 masses of respectively 0.4 ± 0.03 , 1.0 ± 0.1 and 1.0 ± 0.07 mg and body lengths of 1.8
9 ± 0.06 , 2.7 ± 0.01 and 3.1 ± 0.05 mm (N=7 for each species) (Table 1). By contrast,
10 the second group contained two heavier and larger species, *A. armatorius* and *N.*
11 *testacea*, with *A. armatorius* having a mass of 56.9 ± 0.83 mg and a body length of
12 19.5 ± 0.5 mm (N=7).

13 In all species the middle legs were either the same length as the front legs (as in *L.*
14 *boulandi*), or they were just 20% longer. The middle legs were also short relative to
15 the body length, ranging from 46% in *A. armatorius* to 68% in *P. puparum* (Table 1).
16 The hind legs in *L. boulandi* and *P. puparum* were 40% longer than the front legs, in
17 *C. glomerata* they were 70% longer and in *A. armatorius* they were 80% longer.
18 Relative to body length, the hind legs were longest at 87% in *C. glomerata* and
19 shortest at 70% in *A. armatorius* (Table 1). The middle femora were 12% wider than
20 those of the front legs in *C. glomerata* but in *A. armatorius* they were only 3% wider
21 (Fig. 1F,G). By contrast, the hind femora of these two species were 74-75% wider
22 than those of the front legs.

23 **Kinematics of jumping and take-off**

24 ***Pteromalus puparum***

25 In *P. puparum* 65 of 71 jumps (91.55%) performed by 11 individuals were propelled
26 by movements of the middle legs alone (Figs 2,3, Supplementary material Movie 1).
27 The wings remained closed so that the forward and upward trajectory were determined
28 solely by the forces generated by the legs during the jump. In some jumps, the wings
29 opened after take-off and flapping flight ensued, but this rarely occurred in the
30 experimental conditions.

31 In a selected jump of *P. puparum* viewed from the side (Fig. 2), the hind legs were the
32 first to move some 4 ms before take-off and their contribution to propulsion was brief
33 because they were the first pair of legs to lose contact with the ground 0.6 ms later
34 followed by the front legs 2 ms before take-off (Fig. 2A). During this time the two
35 middle legs moved symmetrically by depressing about their coxo-trochanteral joints
36 and extending about their femoro-tibial joints. These movements continued until take-
37 off so that the middle legs alone were then responsible for propulsion. After take-off,
38 the left and right middle legs crossed because the forces they generated now acted
39 only against the air. Clear differences in when the three pairs of legs lost contact with
40 the ground were revealed by plotting their movements against time (Fig. 2B). The
41 sequence in which the legs moved and lost contact with the ground varied, however,
42 both between successive jumps of an individual and between jumps of different
43 individuals; the hind legs were usually the first to lose contact with the ground, but
44 sometimes the front legs were the first. Different sequences did not correlate with the
45 angle of the body relative to the ground, or with the angle of the jump trajectory after
46 take-off. The action of the middle legs was a constant feature in all jumps; their joints

1 depressed and extended progressively to provide upwards and forwards propulsion,
2 and their tarsi were always the last to lose contact with the ground.

3 Six jumps by *P. puparum* (8.45% of the total jumps by this species) were propelled by
4 the same leg movements but were also accompanied before take-off by a single
5 depression of the wings (Fig. 3, Supplementary material Movie 2). The wings opened
6 30 ms before take-off and reached their fully open and elevated position before the
7 first leg movement occurred. The wings then began to depress while the front legs
8 were the first to lose contact with the ground, followed by the hind legs. At take-off,
9 the wings were completing their first depression movement. After take-off the wings
10 began to beat so that there was a seamless transition to powered flight. The mean
11 wingbeat frequency in these wasps once airborne was 185.4 ± 3.2 Hz so that the
12 period from one full elevation to the next was 5.4 ± 0.1 (N = 10 individuals) (Table 2).

13 *Cotesia glomerata*

14 Take-off by *C. glomerata* was propelled by a series of wingbeats with only small
15 movements of the middle and hind legs occurring (Fig. 4, Supplementary material
16 Movie 3). Preparation for take-off was marked by the adoption of a steep angle of the
17 body relative to the ground (69.1 ± 8.0 degrees) (Table 3) so that the front legs were
18 lifted from the ground. The wings then opened and executed 2.8 ± 0.3 wingbeats
19 before take-off (21 take-offs, N=3 individuals) (Fig. 4A). The mean wingbeat
20 frequency was 126.1 ± 6.7 Hz (period 8.2 ± 0.3 ms) (Table 3). The first wingbeat
21 before take-off was often of small amplitude but thereafter all wingbeats were of a full
22 amplitude during both the acceleration phase of take-off and once airborne (Fig. 4B-
23 D). At the start of the first wing movement the middle and hind legs began to show
24 small changes in the angles between the body and the femora and between the femora
25 and tibiae (Fig. 4A,D). These changes continued after take-off as the legs sagged
26 beneath the body under the influence of gravity, suggesting that the initial small
27 changes observed before take-off might be due to the lift generated by the wings
28 rather than active propulsion by the legs. The hind legs were the next to lose contact
29 with the ground so that as in *P. puparum*, it was the middle legs that finally lost
30 contact at take-off while the wings were being depressed (Fig. 4D).

31 *Leptopilina boulardi*

32 In *L. boulardi*, the lightest of all the species analysed, propulsion for take-off shifted
33 further towards a greater reliance on the wings. Preparation for take-off was initiated
34 by tilting of the front of the body upwards so that the front legs lost contact with the
35 ground and thus made no further contribution. The wings then opened and executed
36 3.8 ± 0.1 wingbeats (28 take-offs, N=4 individuals) with take-off again being
37 accomplished during wing depression (Figs 5, 6). The mean wingbeat frequency was
38 the highest at 198.9 ± 5.4 Hz (mean period 5.0 ± 0.1 ms (N= 4 individuals)) (Table 2).
39 No contribution to propulsion could be discerned by the middle and hind legs as
40 viewed from directly in front (Fig. 5A) or from the side (Fig. 6, Supplementary
41 material Movie 4). The angles between the body and femora, and between the femora
42 and tibiae in both pairs of these legs did not change before take-off, but once airborne
43 they increased as the legs sagged beneath the body (Fig. 5B, C). The wing movements
44 described symmetrical paths leading to take-off while the movements of the body
45 described an upward trajectory with a mean angle of 91.2 ± 31.8 degrees but with
46 much variation (Fig. 5B, Table 3). The rhythmic wing movements continued after
47 take-off so that there was a smooth transition to powered flight.

1 **Ichneumonidae**

2 *A. armatorius* and *N. testacea*, were the largest wasps analysed and both used a
3 combination of leg and wing movements to propel take-off (Figs 7, 8). The legs were,
4 however, used in a different sequence to that described for the preceding wasps. The
5 wings opened first and then before they started to beat (Fig. 7A, Supplementary
6 material Movie 5), or during their first depression (Fig. 8A), the hind legs began their
7 propulsive movements. The front legs were the first to leave the ground followed by
8 the middle legs (Figs 7B, 8B). The hind legs were thus the only legs adding to the
9 forces during the latter part of the propulsive phase of the take-off. The wings
10 generated a mean of 2.6 ± 0.2 wingbeats (27 take-offs) before take-off lowest mean
11 frequency for any of the wasps recorded here of 80 ± 3.7 Hz (mean period 13.1 ± 0.5)
12 (N= 9 individuals) (Table 2). Both the angle between the body and the hind femora
13 and between the femora and the tibiae changed progressively to depress and extend
14 the hind legs to take-off while the wings were beating (Fig. 7B). For *N. testacea*
15 plotting the trajectories described by the tip of the right front wing before take-off
16 together with the movements of the tarsi of the right middle and hind legs showed that
17 take-off occurred during the depression phase of a wingbeat and that the middle legs
18 lost contact with the ground well before the hind legs (Fig. 8B).

19 **Jumping and take-off performance**

20 Jumping and take-off performance were derived from measurements taken from the
21 high speed videos, and from subsequent calculations (Table 3). The time taken to
22 accelerate to take-off varied by a factor of six between the different species. It was
23 shortest at 5.0 ± 0.3 ms (three jumps by each of 11 wasps) in *P. puparum*, which was
24 propelled by leg movements. The acceleration times of take-offs by this species that
25 were assisted by a single depression of the wings all fell within the range of times
26 shown in jumps propelled only by the legs, suggesting only a small contribution of the
27 wings to lift. In other species where take-off was propelled by a few wingbeats, the
28 take-off time was longer at 19.3 ± 1.1 ms (N=4 individuals) in *L. bouleari* and 22.8
29 ± 2.8 ms (N=3 individuals) in *C. glomerata*. It was longest at 33.8 ± 1.9 ms (N=9
30 individuals) in the heaviest wasp *A. armatorius* that was propelled by combined leg
31 and wing movements.

32
33 Take-off velocity varied by a factor of four between the different species. The fastest
34 velocities, mean (0.8 ± 0.1 m s⁻¹) and best (0.99 m s⁻¹), were achieved by *P. puparum*.
35 Both *L. bouleari* and *C. glomerata* which have a similar mass, but were propelled by
36 wing movements, could only achieve mean take-off velocities that were a quarter of
37 this at 0.2 m s⁻¹ (Table 3). The heaviest wasp *A. armatorius* that used leg and wing
38 movements had an intermediate take-off velocity with a mean of 0.4 ± 0.03 m s⁻¹ and
39 best value of 0.6 m s⁻¹.

40
41 The trajectory of the take-off and the angle of the body relative to the horizontal were
42 correlated with the propulsive strategy that was used. Where the legs provided the sole
43 or main propulsion, as in *P. puparum*, the mean trajectory angle was 65.8 ± 7.8
44 degrees and where leg movements were combined with wing movements, as in the
45 much heavier *A. armatorius*, the angle was shallower at 54.4 ± 4.9 degrees (Table3).
46 The body angle relative to the ground was also low at 35.1 ± 4.6 and 25.4 ± 2.6
47 degrees respectively. In contrast, where wing movements were the sole or dominant
48 propulsive force, the trajectories were much steeper; in *L. bouleari* they were almost
49 vertical (mean 91.2 ± 5.4 degrees), and in *C. glomerata* they were backwards (mean

1 110.7 ± 14.6 degrees). The angle of the body at take-off was also steeper at 63.3
2 ± 10.2 degrees and 69.1 ± 8.0 degrees respectively.

3
4 The acceleration experienced at take-off was highest at 17 g in *P. puparum* which had
5 the fastest take-off velocity and shortest acceleration time (Table 3). By contrast, in
6 species with both slower take-off velocities and longer acceleration times, the values
7 ranged only between 1-2 g.

8
9 The energy for take-off (E) was calculated in the same way (Equation 1) for each of
10 the three strategies used by these parasitoid wasps

11
12 (1) $E = 0.5 m v^2$

13
14 Where m = mass, kg
15 v = velocity, m s⁻¹

16
17 In *P. puparum* the mean energy required for a jump propelled by leg movements was
18 0.3 µJ and the mean power was 0.06 mW. In the two other species of similar mass, but
19 in which take-off was propelled by wing movements the mean energy required was
20 only 0.01- 0.02 µJ and the mean power 0.0003 - 0.001 mW. In the much heavier *A.*
21 *armatorius*, these values were 10.9 µJ and 0.15 mW in its best jumps.

22
23 The power requirements for take-off in its best jumps by *P. puparum* were 766 W kg⁻¹
24 muscle whereas in the remaining three species they were dramatically lower at 13 - 24
25 W kg⁻¹ muscle in their best jumps. This calculation assumes that the muscles used to
26 propel take-off comprised 10% of body mass.

27 28 29 **DISCUSSION**

30 **Consequences of different strategies for take-off**

31 This paper has shown that different species of parasitoid wasps use different strategies
32 to generate take-off. In three species of similar mass (0.4 – 1.0 mg), two distinct
33 strategies were found even in species that parasitise the same host. *P. puparum*
34 propelled 92% of its jumps by rapid leg movements alone with no accompanying
35 movements of the wings. In the small number of their remaining jumps, the wings
36 executed just one depression movement of the wings before take-off which did not
37 increase take-off velocity above that achieved by the propulsive leg movements alone.
38 By contrast, in *C. glomerata* and *L. boulandi* wings movements were respectively
39 either the main, or only contributors of force to take-off. In the much heavier *A.*
40 *armatorius* (mass 50 times greater) a combination of propulsive leg movements and
41 wing beating generated take-off. Only one species showed evidence of being able to
42 change strategies. In a few take-offs, *P. puparum* added one cycle of wing depression
43 to the same sequence of leg movements. The measurements were made in a limited
44 behavioural context in which the take-offs could not be related directly to a particular
45 sensory stimulus. Some take-offs appeared to be “spontaneous” but the possibility
46 remains that others might have been triggered by the presence of members of the same
47 species in the chamber at the same time. In insects such as *Drosophila* different take-
48 off strategies are clearly used in different behavioural contexts (Card and Dickinson,
49 2008; Trimarchi and Schneiderman, 1995; von Reyn et al., 2014).

1 The following conclusions can be drawn about the effectiveness of the strategies for
2 take-off, particularly by comparing the performance of the three species with similar,
3 low masses. The shortest acceleration time of 5 ms and a mean take-off velocity of
4 0.8 m s^{-1} , with the fastest take-off reaching almost 1 m s^{-1} , was achieved by take-off
5 propelled by movements of the legs in *P. puparum*. By contrast, in two other species
6 of similar mass, take-off generated by beating the wings a mean of 2.8 times in *C.*
7 *glomerata* and 3.8 times in *L. boulandi* resulted in acceleration times that were four
8 times longer at 19-23 ms and take-off velocities that were four times slower at
9 0.2 m s^{-1} . The take-off angles and the subsequent trajectories of these two species once
10 airborne were also much steeper than those of *P. puparum* which was propelled only
11 by jumping movements of the legs. These three species of wasps had bodies of similar
12 size, shape and mass. The legs of *P. puparum* and *L. boulandi* were also of similar
13 proportions relative to each other and to body length. The exception was *C. glomerata*
14 the hind legs of which were 70% longer than the front legs and the longest relative to
15 body, but this increased leverage did not apparently contribute to take-off.

16
17 To calculate the energy expended in translation of the body, the same method was
18 used for take-offs generated by both strategies. This method calculates the energy put
19 into propelling the body into the air. Take-offs of *P. puparum* generated by leg
20 movements resulted in $0.3 \mu\text{J}$ of energy being put into achieving this goal, whereas in
21 take-offs generated by flapping movements of the wings, this value fell by 15-30
22 times to 0.02 or $0.01 \mu\text{J}$. The reason for this dramatic difference is that in the latter
23 strategy, energy is lost in generating repetitive movements of the wings and in moving
24 air rather than in propelling the body upwards and forwards. These data therefore
25 support the hypothesis of this paper that jumping as a mechanism to propel take-off is
26 more energy efficient than propulsion generated by flapping movements of the wings.
27 This energy efficiency is thus a further advantage conferred by jumping in addition to
28 a shorter acceleration time to take-off and a faster take-off velocity.

29
30 This conclusion is even more striking when consideration is given to the mechanical
31 power that is transmitted to the body during take-off. In all strategies, the relevant
32 muscles must generate much power but the amount seen in the output kinematics is
33 very different for the different strategies. In take-offs by *P. puparum* that are
34 propelled by leg movements, this power is almost completely transferred to the final
35 movement of the wasp (Table 3). The high power outputs of this species may also
36 indicate a contribution from hind leg muscles to take-off. By contrast, in *C. glomerata*,
37 *L. boulandi*, and *A. armatorius* in which take-offs are propelled by wing movements,
38 less than 10% of the power generated by the muscles is transferred directly to the final
39 movement of the wasp (Table 3). Most of the power is used in accelerating the wings
40 and the air beneath the wings, and not into accelerating the wasp itself.

41
42 In the heavier species such as *A. armatorius*, both leg and wing movements were
43 combined to propel take-off in the longest acceleration time of 34 ms and to a take-off
44 with a velocity of 0.4 m s^{-1} that is half generated by *P. puparum* when jumping.
45 Lifting the heavy body must contribute to this long acceleration time and lower take-
46 off velocity and offers a possible explanation of why a combination of leg and wing
47 movements are needed to effect take-off.

48 **Take-off performance**

49 How does the jumping performance of the wasps analysed here compare with other
50 insects that also use a power generating mechanism that depends on direct
51 contractions of leg muscles and four propulsive legs? The take-off velocity of *P.*

1 *puparum* matched that of lacewings (Neuroptera) $0.5 - 0.6 \text{ m s}^{-1}$ (Burrows and
2 Dorosenko, 2014) and moths (Lepidoptera) $0.6 - 0.9 \text{ m s}^{-1}$ (Burrows and Dorosenko,
3 2015a), overlapped with some caddis flies (Trichoptera) $0.7 - 1.0 \text{ m s}^{-1}$ (Burrows and
4 Dorosenko, 2015b), but fell short of the velocities achieved by a praying mantis
5 (Mantodea) 1.0 m s^{-1} (Burrows et al., 2015) and the fly *Hydrophorus albofloreus*
6 (Diptera) 1.6 m s^{-1} (Burrows, 2013a). Two of the species of wasps analysed here, *C.*
7 *glomerata* and *L. boulandi*, used the wings alone to propel take-off and reached low
8 velocities of 0.2 m s^{-1} . Their take-offs were almost vertical or sometimes slightly
9 backwards so their trajectories were much steeper than those of wasps that are
10 propelled by the legs alone or assisted by the wings. Caddis flies also differ in their
11 take-off trajectory depending on the strategy they use (Burrows and Dorosenko,
12 2015b).

13 **Use of two pairs of legs for take-off**

14 A number of jumping solutions have evolved in insects particularly when the legs are
15 used for propulsion. When one pair of legs is used by true flies (Diptera) it is the
16 middle legs (Hammond and O'Shea, 2007; Trimarchi and Schneiderman, 1995b), but
17 in other insects it is more frequently the hind legs. In bush crickets these legs are long
18 relative to the body and allow jumps to be propelled by direct contractions of the
19 muscles acting on these long levers (Burrows and Morris, 2003). Higher take-off
20 velocities have been achieved by catapult mechanisms involving the hind legs in
21 insects such as grasshoppers, fleas and froghoppers (Bennet-Clark, 1975; Bennet-
22 Clark and Lucey, 1967; Burrows, 2003). In catapult mechanisms, energy can be stored
23 in advance of a jump to enable power to be amplified with a concomitant reduction of
24 the acceleration time. The stored energy is then released suddenly to generate fast leg
25 movements that propel a jump. So rapid are the leg movements of frog-, plant- and
26 tree-hoppers (Burrows, 2006; Burrows, 2009; Burrows, 2013b) using this mechanism
27 that additional specialisation are needed to ensure that the leg movements are
28 synchronised (Burrows, 2010; Burrows and Sutton, 2013) so as not to result in
29 rotation of the body and the loss of energy for forward momentum.
30

31 In other insects, such as the wasps described here, both the middle and hind legs can
32 potentially be used to propel jumping or assist in take-off. The front legs can be
33 excluded as in none of the recorded take-offs do they move in ways that are consistent
34 with generating propulsion. In snow fleas (Burrows, 2011), praying mantis (Burrows
35 et al., 2015) and the Dolichopodid fly *Hydrophorus albofloreus* (Burrows, 2013a) the
36 middle and the hind legs leave the ground at the same time so both are able to
37 contribute thrust during the last part of the acceleration phase of take-off. In caddis
38 flies (Trichoptera) (Burrows and Dorosenko, 2015b) the middle legs are the last to
39 leave the ground, but in moths (Lepidoptera) (Burrows and Dorosenko, 2015a),
40 lacewings (Neuroptera) (Burrows and Dorosenko, 2014) and mirid bugs (Hemiptera)
41 (Burrows and Dorosenko, 2017) it is the hind legs. In the wasp *P. puparum*, the
42 middle legs are the last to leave the ground so that it is using the same strategy as
43 caddis flies (Burrows and Dorosenko, 2015b). By contrast, the hind legs of
44 *A. armatorius* are the last to lose contact with the ground so that it is using the same
45 strategy as ants (Baroni Urbani et al., 1994; Tautz et al., 1994), moths (Burrows and
46 Dorosenko, 2015a) and lacewings (Burrows and Dorosenko, 2014).
47

48 What advantages does the use of two propulsive pairs of legs confer? First, four
49 propulsive legs ensure that forces needed for take-off are distributed over a larger
50 surface area of the substrate. This could allow take-off from more compliant surfaces.
51 Adding thrust from wing movements should further reduce the forces transmitted

1 directly to the ground, while at the same time allowing a smooth transition into
2 forward, powered flight.

3
4 A second advantage is that the muscles of both pairs of legs power the movement,
5 effectively almost doubling the muscle mass available for take-off. Estimates that the
6 jumping muscles represent about 10% of total body mass are based on measurements
7 from insects using just one pair of propulsive legs. On this basis, the calculated energy
8 requirements for take-off by the wasps studied here are low and only in *P. puparum*
9 do they reach the high end of values shown for muscle from a range of animals
10 (Askew and Marsh, 2002; Ellington, 1985; Josephson, 1993; Weis-Fogh and
11 Alexander, 1977). Increasing the muscle mass used for jumping would thus reduce the
12 expected energy demands to well within these limits and allow all the wasp jumps and
13 take-offs to be explained by direct contractions of the muscles. Mechanisms involving
14 energy storage have been invoked (Gibson, 1986) to explain the take-offs of some
15 wasps, but the data presented here indicates that such mechanisms are not required for
16 the performance of the parasitoid wasps studied here. A clear disadvantage of using
17 four legs is that it precludes achieving the high take-off velocities that catapult
18 mechanisms can generate, because no neural or mechanical mechanisms have been
19 found that could synchronise all these legs with the necessary precision.
20
21

22 **ACKNOWLEDGEMENTS**

23 Many thanks to Roger Northfield for supplying *C. glomerata* and *P. puparum* from
24 his culture of cabbage white butterflies and to Sophie Smith for supplying *L. boulandi*
25 from *Drosophila* cultures. We thank William Foster for help with the identification
26 and classification of these wasps and Peter Lawrence for many helpful suggestions
27 during the experimental work.
28

29 **COMPETING INTERESTS**

30 No competing interests declared.
31

32 **FUNDING**

33 This research received no specific grant from any funding agency in the public,
34 commercial, or not-for-profit sectors.
35

36 **AUTHOR CONTRIBUTIONS**

37 M.B. initiated the project, carried out the experimental work and prepared the first
38 draft of the paper. M.D. analysed the videos to determine take-off performance and
39 contributed to subsequent versions of the manuscript.
40
41
42

Table 1. Body form of parasitoid wasps

Body length and mass, and lengths of the hind femora and tibiae (grand means \pm s.e.m.) of four species of parasitoid wasps (N=7 for each species). The ratios of leg lengths are given relative to the front legs.

	Body Mass, mg	Body Length, mm	Hind leg, Femur, mm	Hind leg, Tibia, mm	Ratio of leg lengths			Middle leg length as % of body length	Hind leg length as % of body length	Hind leg length (mm)/ body mass ^{1/3} (mg)
					Front	Middle	Hind			
<i>Pteromalus puparum</i>	1.0 \pm 0.10	2.7 \pm 0.01	0.6 \pm 0.02	0.7 \pm 0.03	1	1.2	1.4	68	78	2.1
<i>Cotesia glomerata</i>	1.0 \pm 0.07	3.1 \pm 0.05	0.7 \pm 0.03	0.8 \pm 0.03	1	1.2	1.7	59	87	1.6
<i>Leptopilina boulardi</i>	0.4 \pm 0.03	1.8 \pm 0.06	0.4 \pm 0.03	0.5 \pm 0.01	1	1	1.4	61	84	1.6
<i>Amblyteles armatorius</i>	56.9 \pm 0.83	19.5 \pm 0.50	3.4 \pm 0.05	3.9 \pm 0.09	1	1.2	1.8	46	70	3.6

Table 2. Wingbeat frequency and period

The number of wingbeats before take-off, the wingbeat frequency and period for four species of parasitoid wasps analysed in detail. N = number of individuals of each species, n = the total number of wingbeats measured; grand means (\pm s.e.m.) for each species. At least 3 wingbeats were measured after take-off for each individual wasp.

Species	Number of wingbeats before take off	Wingbeat frequency, Hz	Wingbeat period, ms
<i>Pteromalus puparum</i> (N = 10, n = 30)	Single depression	185.4 \pm 3.2	5.4 \pm 0.1
<i>Cotesia glomerata</i> (N = 3, n = 21)	2.8 \pm 0.3	126.1 \pm 6.7	8.2 \pm 0.3
<i>Leptopilina boulardi</i> (N = 4, n = 28)	3.8 \pm 0.1	198.9 \pm 5.4	5.0 \pm 0.1
<i>Amblyteles armatorius</i> (N = 9, n = 27)	2.6 \pm 0.2	80.0 \pm 3.7	13.1 \pm 0.5

Table 3. Jumping performance of parasitoid wasps

Data in columns 2–6 are the grand means (\pm s.e.m.) for the measured jumping performance of each of the four species; the best performance (defined by the fastest take-off velocity) of a particular individual is also given. The values in columns 7–12 on the right are calculated from these measured data. N = number of individuals of each species that were analysed

	Body Mass	Time to take off	Take-off velocity	Take-off angle	Body angle at take-off	Acceleration	g force	Energy	Power	Force	Power/kg muscle
Formula	m		v			$f = v/t$	$g = f/9.81$	$E = 0.5 m v^2$	$=E/t$	$=m f$	$=\text{power} / (.1m)$
Units	mg	ms	$m s^{-1}$	degrees	degrees	$m s^{-2}$	g	uJ	mW	mN	$W kg^{-1}$
<i>Pteromalus puparum</i>											
Average (N=11)	1.0 \pm 0.1	5.0 \pm 0.3	0.8 \pm 0.1	65.8 \pm 7.8	35.1 \pm 4.6	163	17	0.3	0.06	0.2	656
Best	0.7	6.4	0.99	67.8	37.5	155	16	0.3	0.05	0.1	766
<i>Cotesia glomerata</i>											
Average (N=3)	1.0 \pm 0.07	22.8 \pm 2.8	0.2 \pm 0.01	110.7 \pm 14.6	69.1 \pm 8.0	9	1	0.02	0.001	0.01	10
Best	0.8	22.0	0.2			11	1	0.02	0.001	0.01	13
<i>Leptopilina boulardi</i>											
Average (N=4)	0.4 \pm 0.03	19.3 \pm 1.1	0.2 \pm 0.02	91.2 \pm 31.8	63.3 \pm 10.2	9	1	0.01	0.0003	0.003	8
Best	0.4	14.2	0.2			17	2	0.01	0.0008	0.007	20
<i>Amblyteles armatorius</i>											
Average (N=9)	56.9 \pm 0.83	33.8 \pm 1.9	0.4 \pm 0.03	54.4 \pm 4.9	25.4 \pm 2.6	12	1	4.7	0.14	0.7	24
Best	60.6	74.0	0.6	39.1	20.7	8	1	10.9	0.15	0.5	24

FIGURE LEGENDS

Fig. 1

Body and leg structure of four species of parasitoid wasps. (A -E) Photographs of four species analysed. (A) Side view of *Pteromalus puparum*. (B) Ventral view of *Cotesia glomerata*. (C,D) Side views of a female *Leptopilina boulardi* (C) with a large abdomen and a male (D) with long antennae. (E) Side view of *Amblyteles armatorius*. (F,G) Photographs of the right front, middle and hind legs viewed laterally of *C. glomerata* (F) and *A. armatorius* (G).

Fig. 2

Take-off by *Pteromalus puparum* propelled only by movements of the legs. (A) Selected images of a jump from the horizontal and to the right and viewed from the side are arranged in two columns with the timing of the frames given relative to take-off at time = 0 ms. Images were captured at 5000 s^{-1} and with an exposure time of 0.1 ms. In this and subsequent Figures, the front legs (LF, left front; RF, right front) are indicated by arrows with yellow heads, the middle legs (LM, RM) by arrows with white heads, and the hind legs (LH, RH) by arrows with pink heads. The triangles in the bottom left hand corners of each image indicate a constant spatial reference point. (B) Plots of the movements, against time, of the tarsi of the front, middle and hind legs during the same jump. The front and hind legs lost contact with the ground during the acceleration phase of the jump so that only the middle legs provided the final propulsion to take-off. The wings did not move.

Fig. 3

Take-off by *Pteromalus puparum* propelled by the legs and one depression of the wings. Selected images of a jump from the horizontal and viewed from the side were captured at 5000 s^{-1} and with an exposure time of 0.1 ms. They are arranged in three columns.

Fig. 4

Take-off by *Cotesia glomerata* propelled by movements of the wings. (A) Selected images captured at 1000 s^{-1} and with an exposure time of 0.2 ms are arranged in two columns. Take-off was from the horizontal and was viewed from the side. The wingbeats are counted from the peaks of elevation and in this and Figs 5, 6 and 8 are indicated in blue. (B,C) Tracks of the movements of the tip of the right front wing (cyan) and of the tarsi of the right middle (white) and right hind legs (pink). In (B) these tracks are superimposed on the image at the start of the first wing depression and in (C) on an image when airborne. The start positions of each track are marked by a large filled circle and the end positions by an **X**. (D) Plot of the movement of the tip of the right front wing, and of the angular changes in particular joints of the right middle and hind legs, against time.

Fig. 5

Take-off by a male *Leptopilina boulardi* propelled by movements of the wings. (A) Selected images of a take-off captured at 5000 s^{-1} and with an exposure time of 0.1 ms are arranged in four columns. The take-off was from the horizontal and towards the camera so that the wasp is viewed head-on. (B) Tracks of the movements of the tips of

the left (green) and right (cyan) hind wings and of the tarsi of the left (blue) and right (white) middle legs. The tracks are superimposed on an image of the wasp at the start of the acceleration phase. The large coloured circles indicate the initial positions, the black dots the positions at take-off and **X** the positions when airborne. (C) Plots against time of the same body parts during the same jump.

Fig. 6

Take-off by a female *Leptopilina boulardi* propelled by movements of the wings. This take-off was from the horizontal and is viewed from the side. Selected images captured at 5000 s^{-1} and with an exposure time of 0.1 ms are arranged in three columns.

Fig. 7

Take-off by *Amblyteles armatorius* propelled by movements of the wings and legs. (A) Selected images of a take-off viewed from the side were captured at 1000 s^{-1} and with an exposure time of 0.2 ms and are arranged in two columns. (B) Movement of the tip of the right front wing (cyan), and the changes in the angle between the body and the femur (pink triangles), and between the femur and tibia (pink squares) of the right hind leg are plotted against time. The propulsive movements of the hind legs began before the wing movements. Take-off occurred during depression of the second wingbeat.

Fig. 8

Take-off by *Netelia testacea* propelled by movements of the wings and legs. (A) Selected images of a take-off viewed from the side were captured at 1000 s^{-1} and with an exposure time of 0.2 ms and are arranged in two columns. Tracks of the movements of the tip of the right front wing (cyan) and the tarsi of the right middle (white) and right hind (pink) legs are superimposed on the image at take-off. (B) Plot of the movements of the same body parts during the same jump against time. Take-off occurred during depression of the third wingbeat.

REFERENCES

- Askew, G. N. and Marsh, R. L. (2002). Muscle designed for maximum short-term power output: quail flight muscle. *J. Exp. Biol.* **205**, 2153-2160.
- Baroni Urbani, C., Boyan, G. S., Blarer, A., Billen, J. and Musthak Ali, T. M. (1994). A novel mechanism for jumping in the Indian ant *Harpegnathos saltator* (Jerdon) (Formicidae, Ponerinae). *Experientia* **50**, 63-71.
- Bennet-Clark, H. C. (1975). The energetics of the jump of the locust *Schistocerca gregaria*. *J. Exp. Biol.* **63**, 53-83.
- Bennet-Clark, H. C. and Lucey, E. C. A. (1967). The jump of the flea: a study of the energetics and a model of the mechanism. *J. Exp. Biol.* **47**, 59-76.
- Bimbard, G., Kolomenskiy, D., Bouteleux, O., Casas, J. and Godoy-Diana, R. (2013). Force balance in the take-off of a pierid butterfly: relative importance and timing of leg impulsion and aerodynamic forces. *J. Exp. Biol.* **216**, 3551-3563.

Branstetter, M. G., Danforth, B. N., Pitts, J. P., Faircloth, B. C., Ward, P. S., Buffington, M. L., Gates, M. W., Kula, R. R. and Brady, S. G. (2017).

Phylogenomic insights into the evolution of stinging wasps and the origins of ants and bees. *Curr Biol* **27**, 1019-1025.

Burrows, M. (2003). Froghopper insects leap to new heights. *Nature* **424**, 509.

Burrows, M. (2006). Jumping performance of froghopper insects. *J. Exp. Biol.* **209**, 4607-4621.

Burrows, M. (2009). Jumping performance of planthoppers (Hemiptera, Issidae). *J. Exp. Biol.* **212**, 2844-2855.

Burrows, M. (2010). Energy storage and synchronisation of hind leg movements during jumping in planthopper insects (Hemiptera, Issidae). *J. Exp. Biol.* **213**, 469-478.

Burrows, M. (2011). Jumping mechanisms and performance of snow fleas (Mecoptera, Boreidae). *J. Exp. Biol.* **214**, 2362-2374.

Burrows, M. (2013a). Jumping from the surface of water by the long-legged fly *Hydrophorus* (Diptera, Dolichopodidae). *J. Exp. Biol.* **216**, 1973-1981.

Burrows, M. (2013b). Jumping mechanisms of treehopper insects (Hemiptera, Auchenorrhyncha, Membracidae). *J. Exp. Biol.* **216**, 788-799.

Burrows, M., Cullen, D. A., Dorosenko, M. and Sutton, G. P. (2015). Mantises exchange angular momentum between three rotating body parts to jump precisely to targets. *Curr. Biol.* **25**, 786-789.

Burrows, M. and Dorosenko, M. (2014). Jumping mechanisms in lacewings (Neuroptera, Chrysopidae and Hemerobiidae). *J. Exp. Biol.* **217**, 4252-4261.

Burrows, M. and Dorosenko, M. (2015a). Jumping mechanisms and strategies in moths (Lepidoptera). *J. Exp. Biol.* **218**, 1655-1666.

Burrows, M. and Dorosenko, M. (2015b). Jumping mechanisms in adult caddis flies (Insecta, Trichoptera). *J. Exp. Biol.* **218**, 2764-2774.

Burrows, M. and Dorosenko, M. (2017). Jumping performance of flea hoppers and other mirid bugs (Hemiptera, Miridae). *J. Exp. Biol.* **in press**.

Burrows, M. and Morris, O. (2003). Jumping and kicking in bush crickets. *J. Exp. Biol.* **206**, 1035-1049.

Burrows, M. and Sutton, G. P. (2013). Interacting gears synchronize propulsive leg movements in a jumping insect. *Science* **341**, 1254-1256.

Card, G. and Dickinson, M. H. (2008). Visually mediated motor planning in the escape response of *Drosophila*. *Curr. Biol.* **18**, 1300-1307.

Ellington, C. P. (1985). Power and efficiency of insect flight muscle. *J. Exp. Biol.* **115**, 293-304.

Gibson, G. A. P. (1986). Mesothoracic skeletomusculature and mechanics of flight and jumping in Eupelminae (Hymenoptera, Chalcidoidea: Eupelmidae). *Can. Ent.* **118**, 691-728.

Hammond, S. and O'Shea, M. (2007). Ontogeny of flight initiation in the fly *Drosophila melanogaster*: implications for the giant fibre system. *J. Comp. Physiol. A* **193**, 1125-1137.

Josephson, R. K. (1993). Contraction dynamics and power output of skeletal muscle. *Annu. Rev. Physiol.* **55**, 527-546.

Larabee, F. J. and Suarez, A. V. (2015). Mandible-powered escape jumps in trap-jaw ants increase survival rates during predator-prey encounters. *PLoS ONE* **10**, e0124871.

Manier, S. and Deamer, D. (2014). Jumping galls: a novel mechanism for motility. *J. Insect Behav.* **27**, 716-721.

Mercier, J.-L. and Lenoir, A. (1999). A new case of jumping behaviour in ants, as part of the foraging strategy. *C.R. Acad. Sci. Paris* **322**, 661-667.

Musthak Ali, T. M., Baroni Urbani, C. and Billen, J. (1992). Multiple jumping behaviours in the ant *Harpegnathos saltator*. *Naturwissenschaften* **79**, 374-376.

Patek, S. N., Baio, J. E., Fisher, B. L. and Suarez, A. V. (2006). Multifunctionality and mechanical origins: Ballistic jaw propulsion in trap-jaw ants. *Proc. Natn. Acad. Sci. U.S.A.* **103**, 12787-12792.

Quicke, D. L., Smith, M. A., Janzen, D. H., Hallwachs, W., Fernandez-Triana, J., Laurenne, N. M., Zaldivar-Riveron, A., Shaw, M. R., Broad, G. R., Klopstein, S., Shaw, S. R., Hrcek, J., Hebert, P. D., Miller, S. E., Rodriguez, J. J., Whitfield, J. B., Sharkey, M. J., Sharanowski, B. J., Jussila, R., Gauld, I. D., Chesters, D. and Vogler, A. P. (2012). Utility of the DNA barcoding gene fragment for parasitic wasp phylogeny (Hymenoptera: Ichneumonoidea): data release and new measure of taxonomic congruence. *Mol. Ecol. Resour.* **12**, 676-85.

Ribak, G., Dafni, E. and Gerling, D. (2016). Whiteflies stabilize their take-off with closed wings. *J. Exp. Biol.* **219**, 1639-1648.

Saeki, Y., Tani, S., Fukuda, K., Iwase, S., Sugawara, Y., Tuda, M. and Takagi, M. (2016). Costs and benefits of larval jumping behaviour of *Bathyplectes anurus*. *Sci. Nat.* **103**, 1-9.

Sorger, D. M. (2015). Snap! Trap-jaw ants in Borneo also jump using their legs. *Frontiers Ecol. Environment* **13**, 574-575.

Sunada, S., Kawachi, K., Watanbe, I. and Azuma, A. (1993). Performance of a butterfly in take-off flight. *J. Exp. Biol.* **183**, 249-277.

Tautz, J., Holldobler, B. and Danker, T. (1994). The ants that jump: different techniques to take off. *Zoology* **98**, 1-6.

Trimarchi, J. R. and Schneiderman, A. M. (1995a). Flight initiations in *Drosophila melanogaster* are mediated by several distinct motor patterns. *J. Comp. Physiol. [A]* **176**, 355-364.

Trimarchi, J. R. and Schneiderman, A. M. (1995b). Initiation of flight in the unrestrained fly, *Drosophila melanogaster*. *J. Zool. Lond.* **235**, 211-222.

von Reyn, C. R., Breads, P., Peek, M. Y., Zheng, G. Z., Williamson, W. R., Yee, A. L., Leonardo, A. and Card, G. M. (2014). A spike-timing mechanism for action selection. *Nature Neuroscience* **17**, 962-970.

Weis-Fogh, T. and Alexander, R. M. (1977). The sustained power output from striated muscle. In *Scale effects in animal locomotion*, (ed. T. J. Pedley), pp. 511-525. London: Academic Press.

Yanoviak, S. P., Dudley, R. and Kaspari, M. (2005). Directed aerial descent in canopy ants. *Nature* **433**, 624-626.

Yanoviak, S. P., Munk, Y., Kaspari, M. and Dudley, R. (2010). Aerial manoeuvrability in wingless gliding ants (*Cephalotes atratus*). *Proc. R. Soc. B.* **277**, 2199-2204.

Figure 1

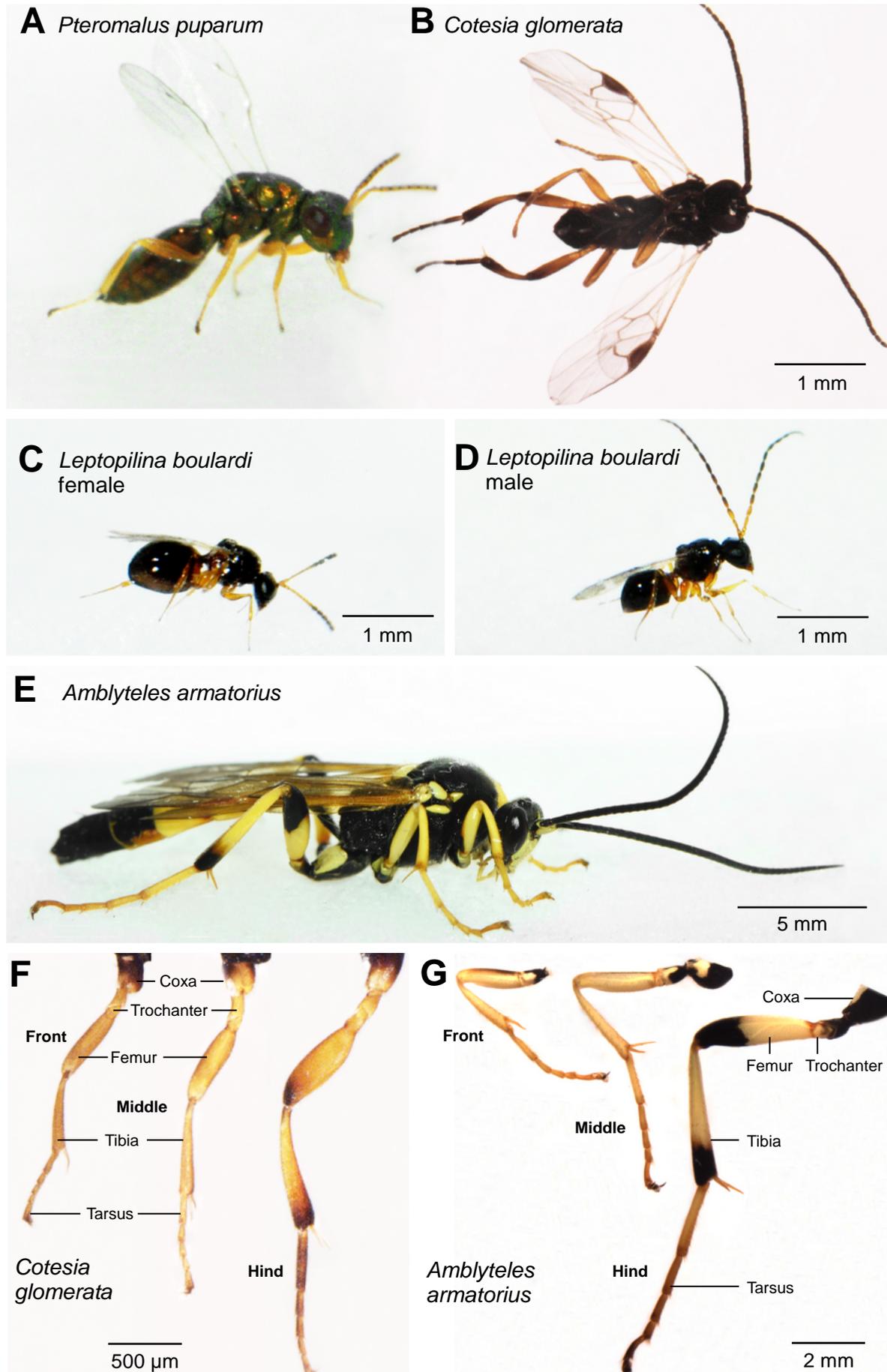


Figure 2

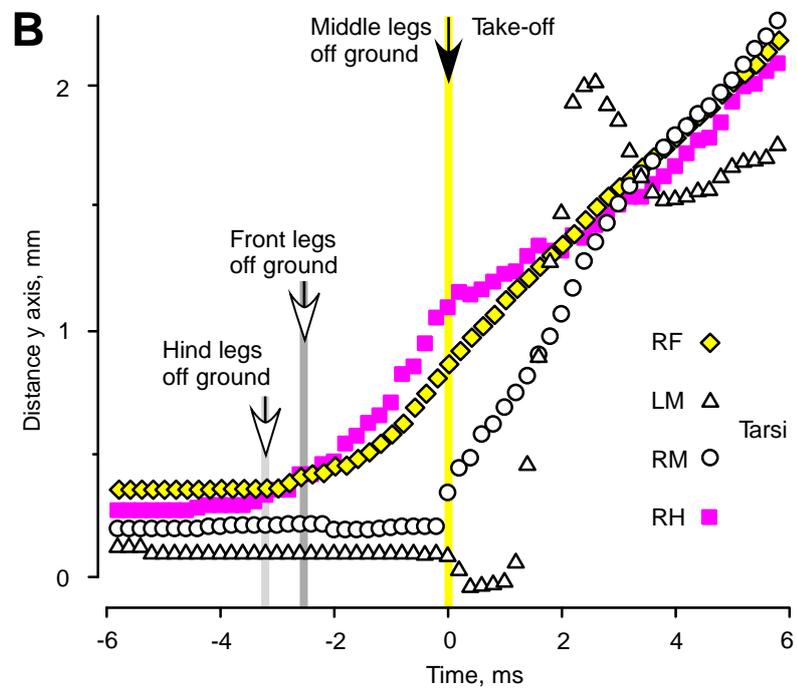
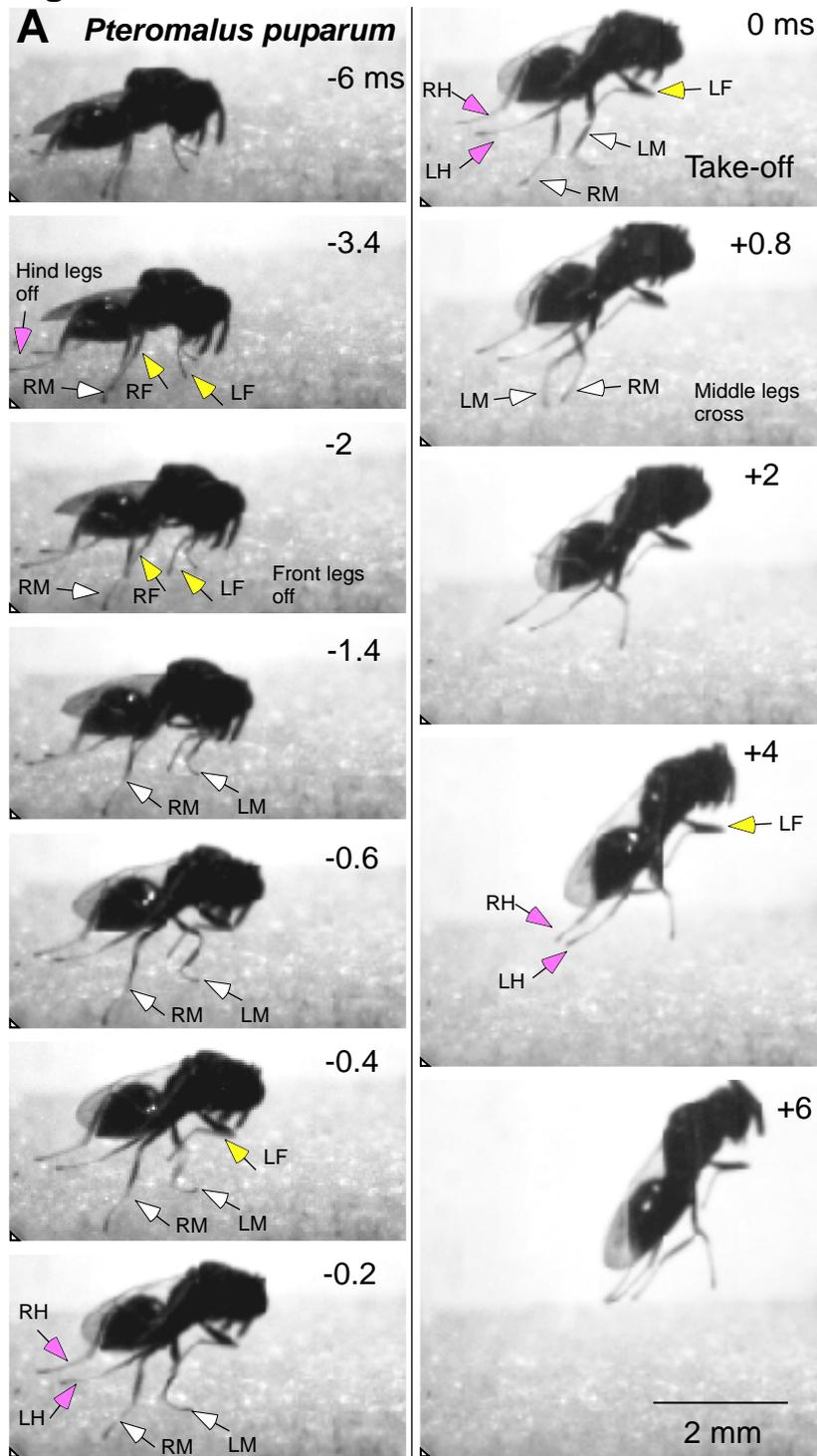


Figure 3

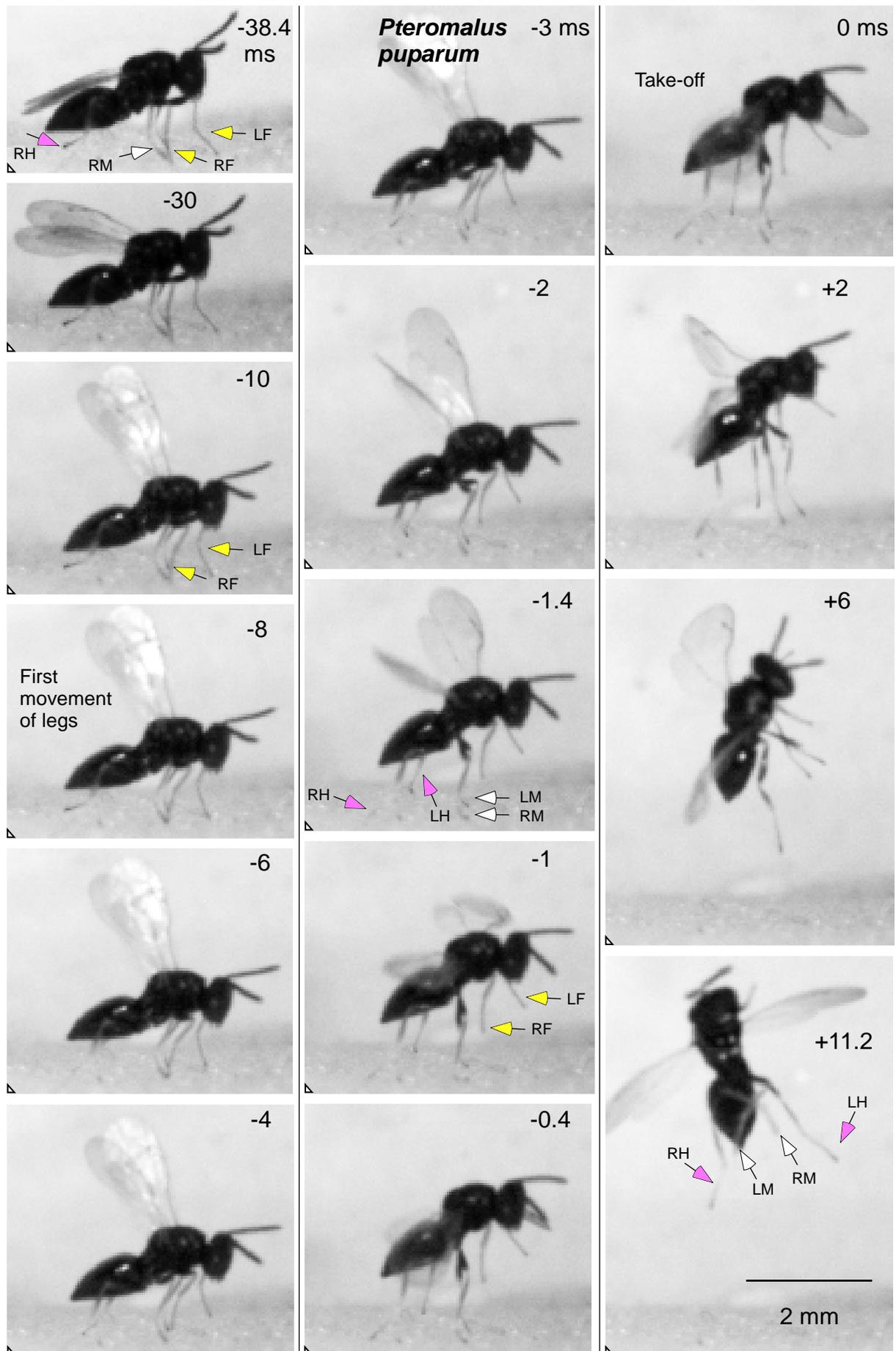


Figure 4

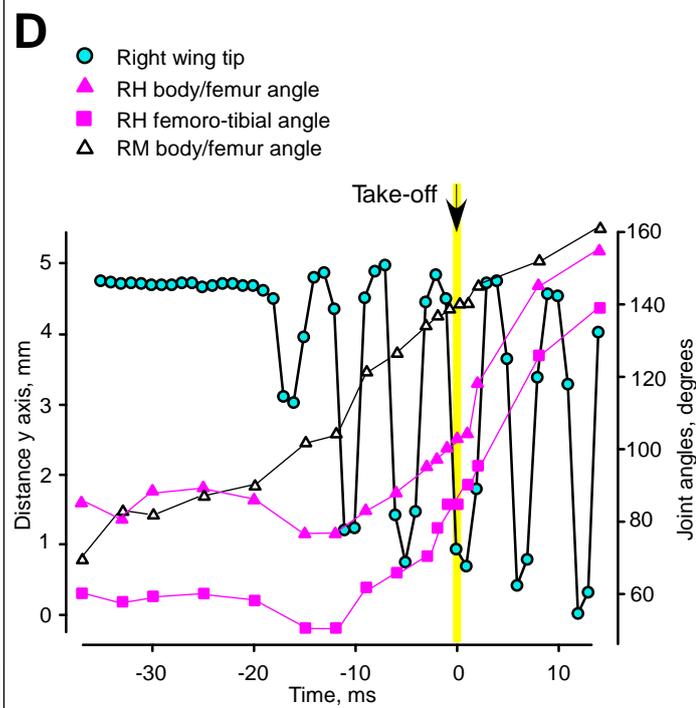
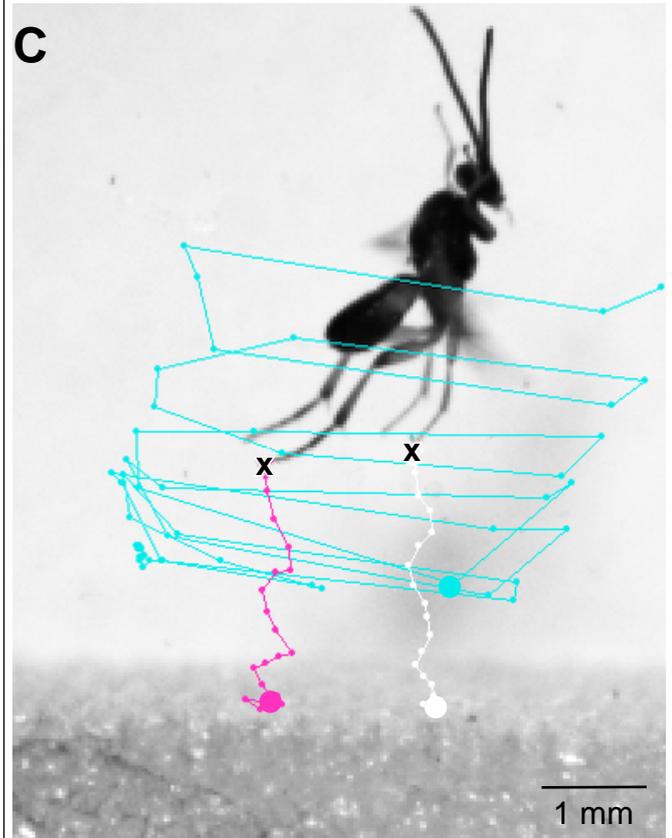
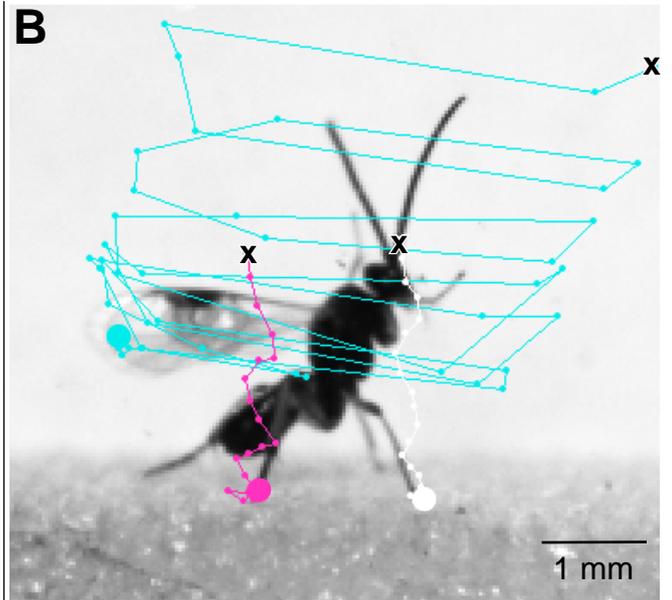
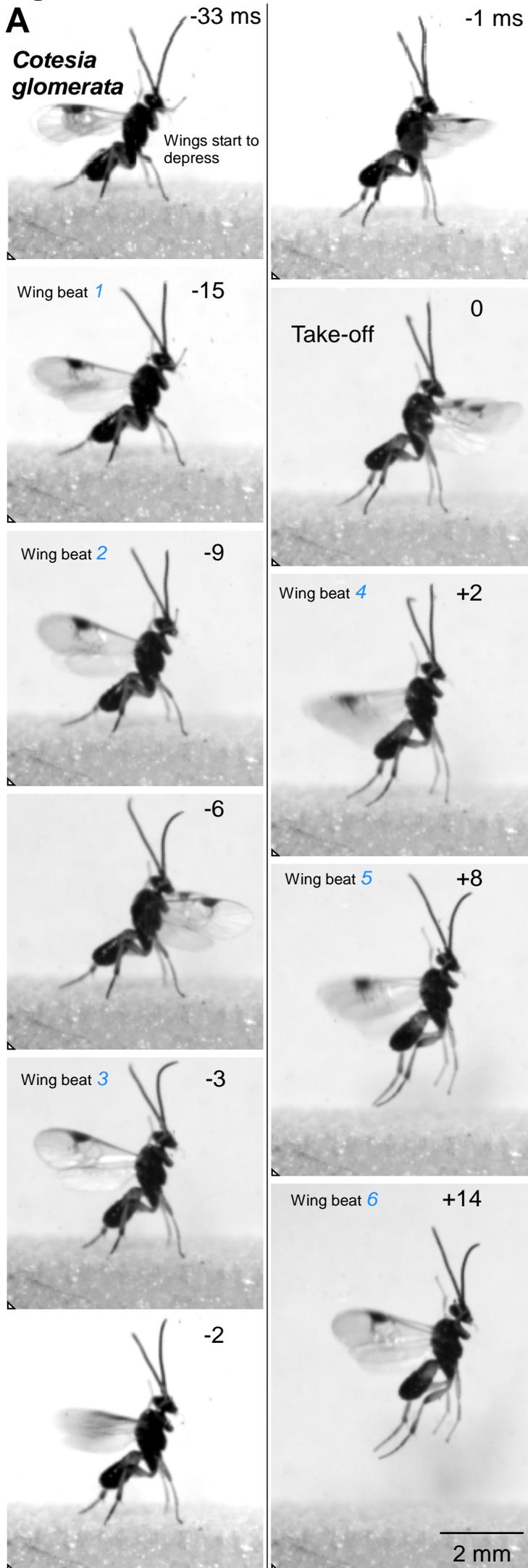


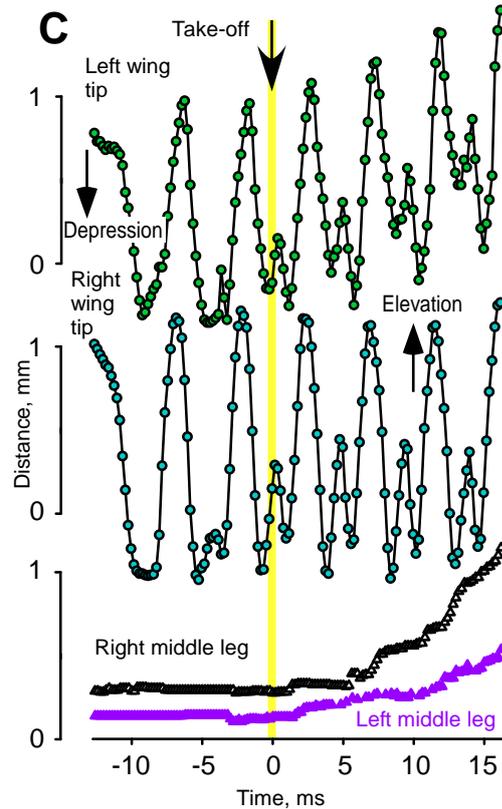
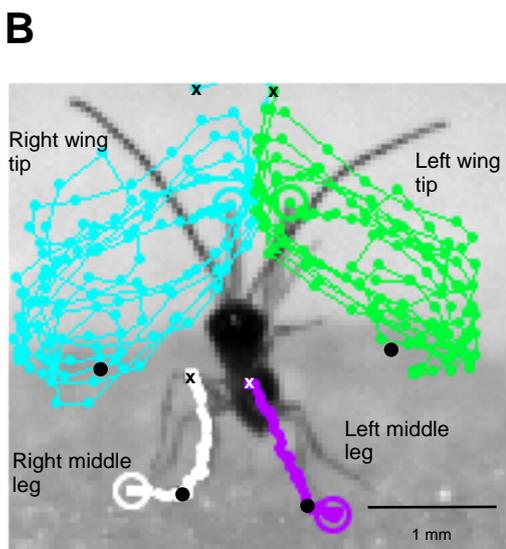
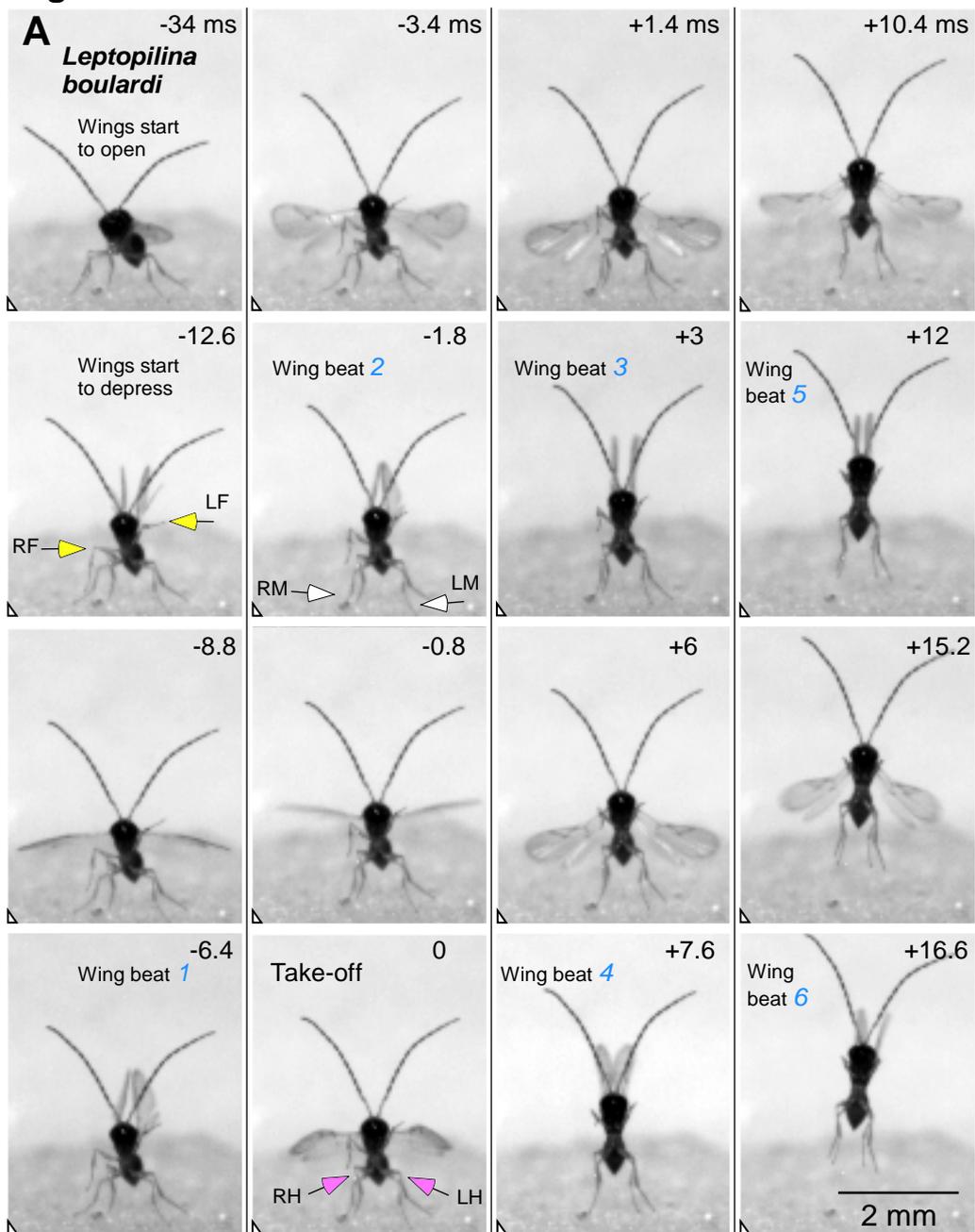
Figure 5

Figure 6

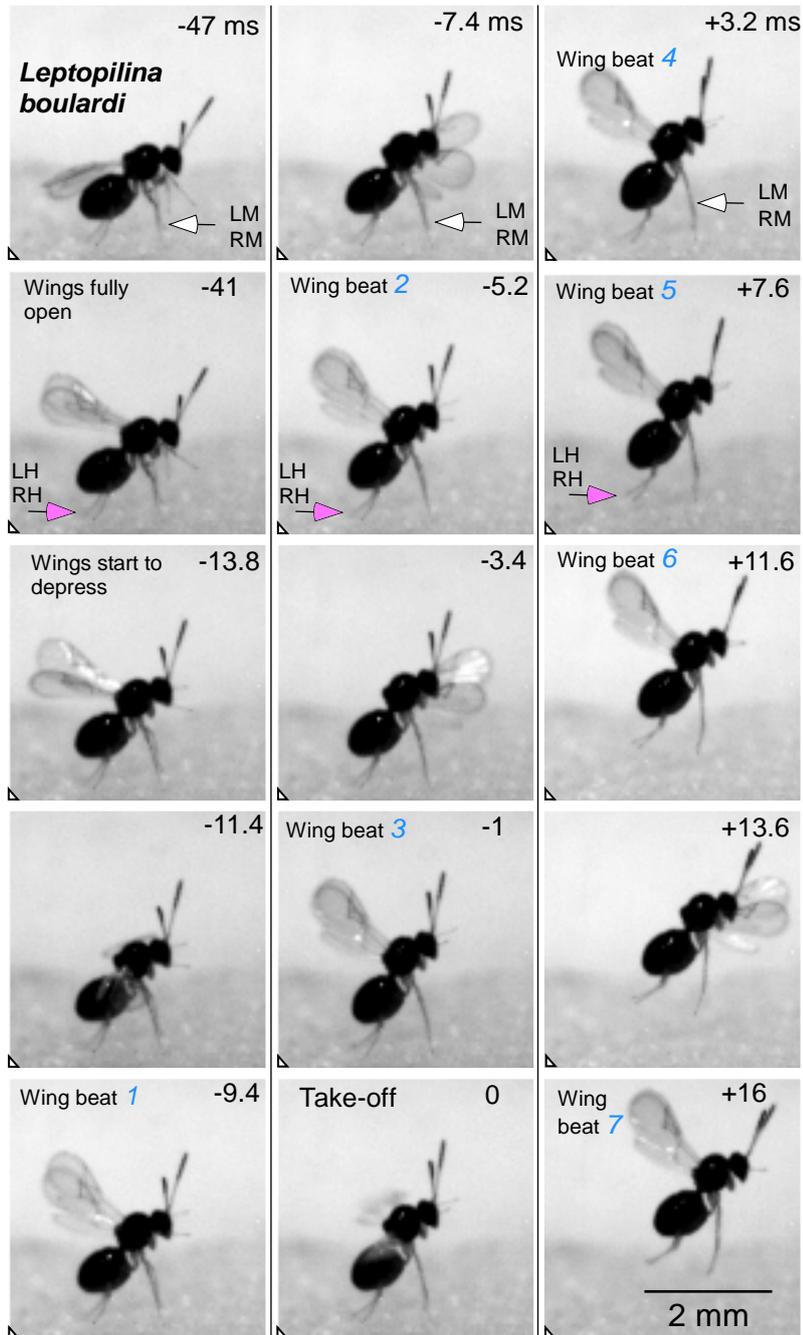


Figure 7

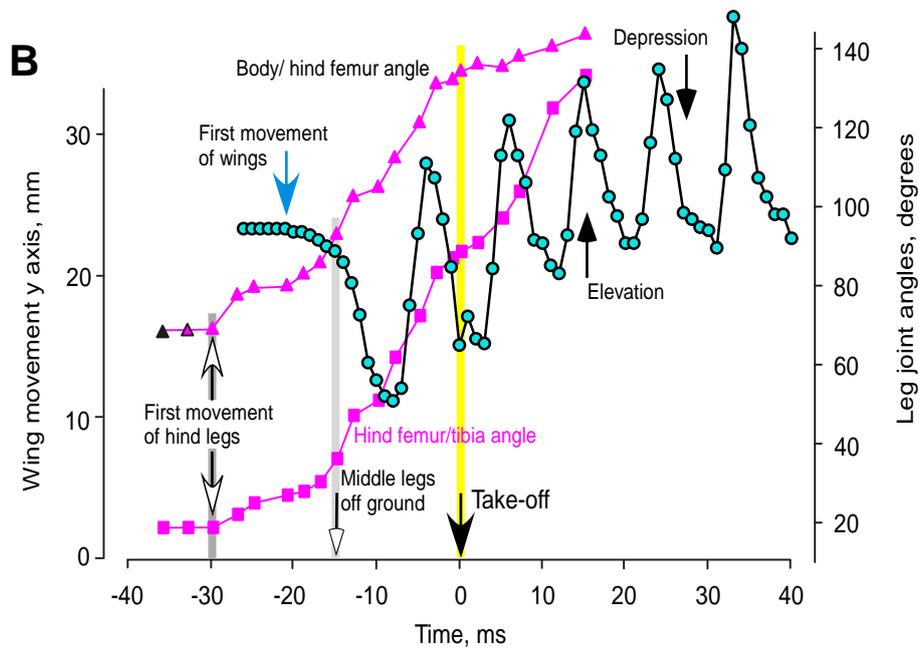
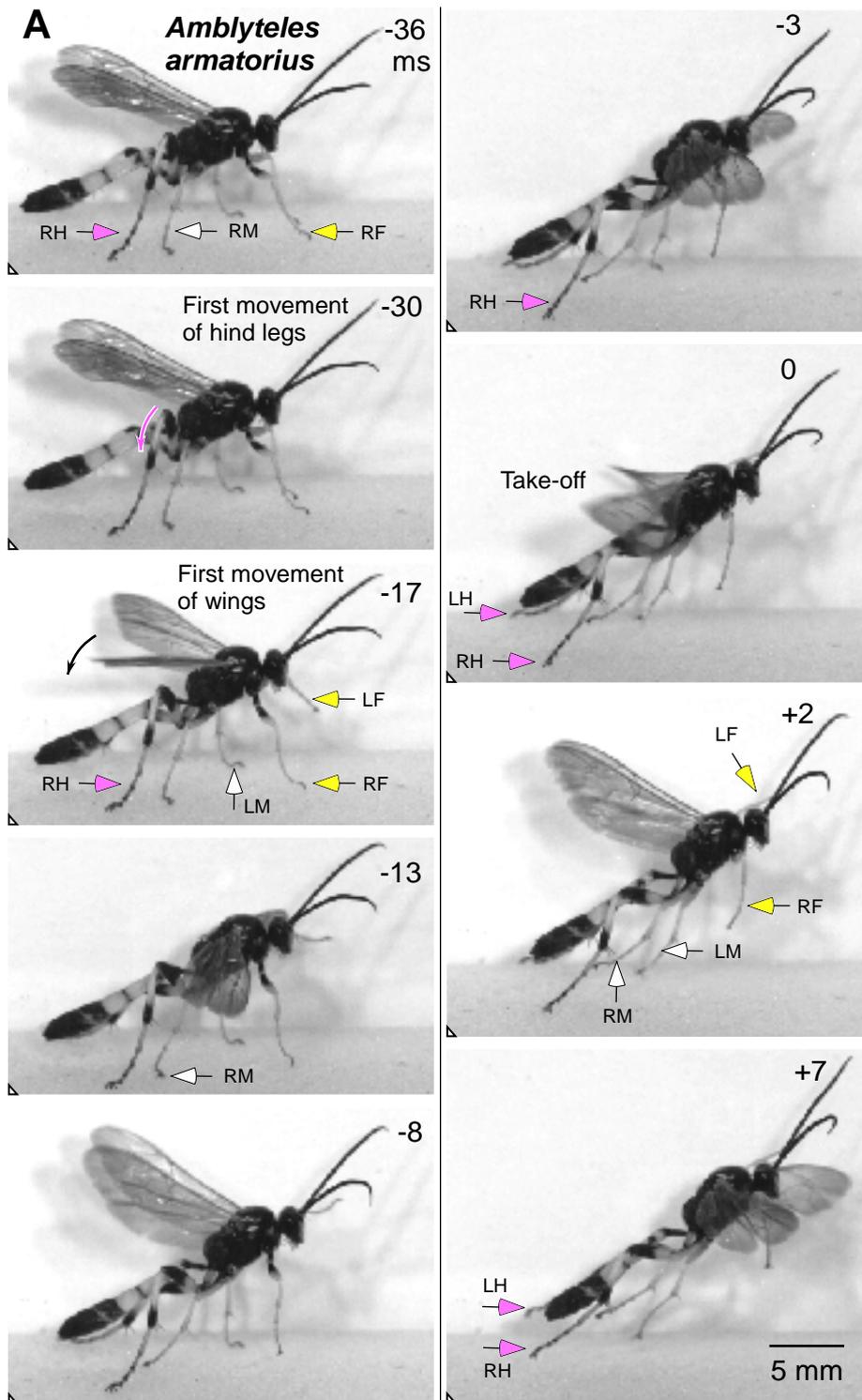


Figure 8

